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**WO 03/103711 A1**

(54) Title: COMBINATIONS OF ANTI-TISSUE FACTOR ANTIBODIES AND ANTICOAGULANT AND/OR ANTIPLATELET AGENTS

(57) Abstract: The invention concerns anti-tissue factor (anti-TF) antibodies with enhanced anticoagulant potency, and methods and means for identifying, producing and using such antibodies. The anti-TF antibodies of the present invention are designed to bind to an epitope comprising the C-terminal macromolecular substrate binding region of TF. The invention also concerns methods of treating TF-VIIa related diseases or disorders comprising administering anti-TF antibodies alone or in combination with at least one additional anticoagulant and/or anti-platelet agent.

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## COMBINATIONS OF ANTI-TISSUE FACTOR ANTIBODIES' AND ANTICOAGULANT AND/OR ANTIPLATELET AGENTS

### Background of the Invention

#### Field of the Invention

This invention concerns methods for engineering anti-tissue factor (anti-TF) antibodies, especially those having enhanced anticoagulant potency. The invention further concerns anti-TF antibodies, methods and means for producing them, compositions comprising the antibodies and their use in the diagnosis, management, prevention and treatment of various diseases and disorders. The invention also concerns combinations of anti-TF antibodies and at least one additional anticoagulant or anti-platelet agent and their use in management, prevention and treatment of various diseases.

#### Description of the Related Art

##### A. Tissue factor

Tissue factor (TF) is the receptor for coagulation factor VIIa (FVIIa) and the zymogen precursor factor VII (FVII). Native human TF (hTF) is a 263 amino acid residue glycoprotein composed of an extracellular domain (residues 1 to 219), a single transmembrane domain (residues 220-242), and a short cytoplasmic domain (residues 243 to 263) (Fisher et al., [1987] Thromb. Res. 48:89-99, Morrissey et al., [1987] Cell 50:129-135). The TF extracellular domain is composed of two immunoglobulin-like fibronectin type III domains of about 105 amino acids each (Huang et al., [1998] J. Mol. Biol. 275:873-894). Each domain is formed by two anti-parallel  $\beta$ -sheets with Ig superfamily type C2 homology.

The protein interaction of FVIIa with TF is mediated entirely by the TF extracellular domain (Muller et al., [1994] Biochem. 33:10864-10870; Gibbs et al., [1994] Biochem. 33:14003-14010; Ruf et al., [1994] Biochem. 33:1565-1572) which has been expressed in *E. coli*, cultured Chinese Hamster Ovary (CHO) cells and *Saccharomyces cerevisiae* (Waxman et al., [1992] Biochemistry 31:3998-4003; Ruf et al., [1991] J. Bio. Chem. 266:2158-2166 and Shigematsu et al., [1992] J. Biol. Chem. 267:21329-21337). The crystal structures of the hTF extracellular domain and its complex with active site inhibited FVIIa have recently been determined by x-ray crystallography (Harlos et al., [1994] Nature 370:662-666; Muller et al., [1994] Biochemistry 33:10864; Muller et al., [1996] J. Mol. Biol. 256:144-159; Banner et al., [1996] Nature 380:41-46).

The hTF extracellular domain has also been extensively characterized by alanine scanning mutagenesis (Kelley et al., [1995] Biochemistry, 34:10383-10392; Gibbs et al., [1994] supra; Ruf et al., [1994] supra). Residues in the area of amino acids 16-26 and 129-

147 contribute to the binding of FVIIa as well as the coagulant function of the molecule. Residues Lys20, Trp45, Asp58, Tyr94, and Phe140 make a large contribution (1 kcal/mol) to the free energy ( $\Delta G$ ) of binding to FVIIa (Kelley et al., (1995) supra). Substitution of Lys20 and Asp58 with alanine residues leads to 78- and 30- fold reductions in FVIIa affinity respectively (Kelley et al., [1995] supra). A set of 17 single-site mutants at other nearby sites that are in contact with FVIIa result in modest decreases in affinity ( $\Delta\Delta G = 0.3 - 1.0$  kcal mol<sup>-1</sup>). Mutations of TF residues Thr17, Arg131, Leu133 and Val207, each of which contact FVIIa in the crystal structure, have no effect on affinity for FVIIa. Lys15Ala and Tyr185Ala mutations result in small increases in affinity ( $\Delta\Delta G = -0.4$  kcal mol<sup>-1</sup>) (Kelley et al., [1995] supra). The 78-fold decrease in affinity imposed by the alanine substitution of Lys20 in hTF can be reversed by substituting a tryptophan for Asp58 (Lee and Kelley, [1998] J. Biol. Chem. 273:4149-4154).

Residues in the area of amino acids 157-168 contribute to the procoagulant function of TF-FVIIa (Kelley et al., [1995] supra; Ruf et al., [1992] J. Biol. Chem. 267:22206-22210) but are not important for FVII/FVIIa binding. It has been shown that lysine residues 165 and 166 are important to TF cofactor function but do not participate in FVIIa complex formation (Roy et al., [1991] J. Biol. Chem. 266:22063; Ruf et al., [1992] J. Biol. Chem. 267:6375). Lysine residues 165 and 166 are located on the C-terminal fibronectin type III domain of TF on the opposite surface of the molecule from residues found to be important for FVIIa binding on the basis of mutagenesis results (Kelley et al., (1995) supra). Alanine substitution of these lysine residues results in a decreased rate of FX activation catalyzed by the TF-FVIIa complex (Ruf et al., (1992) supra). The Lys165Ala-Lys166Ala variant (hTFAA) comprising residues 1-219 of hTF (sTF) inhibits the extrinsic pathway of blood coagulation in vitro through competition with membrane TF for binding to FVIIa. In a rabbit model of arterial thrombosis the variant partially blocks thrombus formation without increasing bleeding tendency (Kelley et al., (1997) Blood 89, 3219-3227). However, high doses of the variant are required for the anti-thrombotic effect, in part because FVIIa binds to cell surface TF approximately 1000-fold more tightly than to sTF (Kelley et al. (1997) supra). The greater apparent affinity is due to interaction of the FVIIa  $\gamma$ -carboxyglutamic acid-containing (Gla) domain with phospholipid.

TF is expressed constitutively on cells separated from plasma by the vascular endothelium (Carson, S. D. and J. P. Brozna, [1993] Blood Coag. Fibrinol. 4:281-292). Its expression on endothelial cells and monocytes is induced by exposure to inflammatory cytokines or bacterial lipopolysaccharide (Drake et al., [1989] J. Cell Biol. 109:389). Upon tissue injury, the exposed extracellular domain of TF forms a high affinity, calcium dependent

complex with FVII. Once bound to TF, FVII can be activated by peptide bond cleavage to yield serine protease FVIIa. The enzyme that catalyzes this step *in vivo* has not been elucidated, but *in vitro* FXa, thrombin, TF-FVIIa and FIXa can catalyze this cleavage (Davie, et al., [1991] Biochemistry 30:10363-10370). FVIIa has only weak activity upon its physiological substrates FX and FIX whereas the TF-FVIIa complex rapidly activates FX and FIX.

The TF-FVIIa complex constitutes the primary initiator of the extrinsic pathway of blood coagulation (Carson, S. D. and Brozna, J. P., (1993) Blood Coag. Fibrinol. 4:281-292; Davie, E. W. et al., [1991] Biochemistry 30:10363-10370; Rapaport, S. I. and L. V. M. Rao, [1992] Arterioscler. Thromb. 12:1111-1121). The complex initiates the extrinsic pathway by activation of FX to Factor Xa (FXa), FIX to Factor IXa (FIXa), and additional FVII to FVIIa. The action of TF-FVIIa leads ultimately to the conversion of prothrombin to thrombin, which carries out many biological functions (Badimon, L. et al., [1991] Trends Cardiovasc. Med. 1:261-267). Among the most important functions of thrombin is the conversion of fibrinogen to fibrin, which polymerizes to form a clot.

The involvement of this plasma protease system has been suggested to play a significant role in a variety of clinical manifestations including arterial and venous thrombosis, septic shock, adult respiratory distress syndrome (ARDS), disseminated intravascular coagulation (DIC), tumor metastasis (Bromberg et al. [1995] Proc. Natl. Acad. Sci. USA 92:8205) and various other disease states (Haskel, E. J. et al., [1991] Circulation 84:821-827; Holst, J. et al., [1993] Haemostasis 23 (suppl. 1):112-117; Creasey, A. A. et al., [1993] J. Clin. Invest. 91:2850-2860; see also, Colman R. W. [1989] N. Engl. J. Med 320:1207-1209; Bone, R. C. [1992] Arch. Intern. Med. 152:1381-1389).

Overexpression and/or aberrant utilization of TF has been linked to the pathophysiology of both thrombosis and sepsis (Taylor et al., [1991] Circ. Shock 33:127; Warr et al., [1990] Blood 75:1481; Pawashe et al., [1994] Circ. Res. 74:56). TF is expressed on cells found in the atherosclerotic plaque (Wilcox et al., [1989] Proc. Natl. Acad. Sci. U.S.A. 86:2839). Disruption of atherosclerotic plaques can result in thrombus formation. Thrombus formation in these circumstances can lead to conditions of acute coronary syndrome such as Unstable Angina (UA) and non-ST-Segment Elevation Myocardial Infarction (NSTEMI). (ACA/AHA Guides for the Management of Patients with Unstable Angina and Non-ST-Segment Elevation Myocardial Infarction, J. Amer. College of Cardiology 36:971-1044 (2000)). These disorders are a major cause of emergency medical care, hospitalization and death in the United States.



Anti-thrombotic therapy is important in the treatment of acute coronary syndrome. Current treatments involve co-administration of heparin, aspirin, and a platelet GP IIb/IIIa inhibitor. Despite improvements in efficacy with the combination therapy, a significant portion of patients have unfavorable outcomes (death or MI). In addition, administration of heparin at therapeutically effective doses is associated with bleeding problems, thrombocytopenia, and in some instances, osteoporosis. (J. Amer. College of Cardiology 36:999-1006 [2000]). Therefore, improvement in clinical outcomes by increasing heparin are limited since more aggressive heparin treatment results in increased bleeding risk. In addition, the combination of heparin and platelet glycoprotein GP IIb/IIIa inhibitors is also associated with high bleeding rates. (Menon et al., Amer. J. Med., 110:641 (2001)). It is desirable to identify novel therapeutic strategies that improve outcomes in thrombotic conditions while minimizing the increase in bleeding risk.

B. Anti-tissue factor antibodies

Monoclonal antibodies in humanized or chimaeric forms are successfully used to treat a variety of diseases (Vaswani and Hamilton, [1998] Ann. Allergy Asthma Immunol. 81: 105-119; Vaughan et al., [1998] Nature Biotechnology 16: 535-539). Antibodies reactive with hTF have been described (Tanaka et al., [1985] Throm. Res., 40:745-756; Tanaka et al., [1986] Chem. Abstracts, 104:366:49211z; Morrissey et al., [1988] Throm. Res., 52:247-260; U.S. Patent No. 5,223,427; Ruf et al., [1992] J. Crystal Growth, 122:253-264; Huang et al., [1998] 275:873-894). Anti-TF monoclonal antibodies have been shown to inhibit tissue factor activity in various primate and non-primate species (Morrissey et al., [1988] supra; Huang et al. [1998] supra). Neutralizing anti-TF monoclonal antibodies have been shown to prevent death in a baboon model of sepsis (Taylor et al., [1991] Circ. Shock 33:127), and attenuate endotoxin-induced DIC in rabbits (Warr et al., [1990], Blood 75:1481).

Inhibition of TF initiated blood coagulation by antibodies reactive with tissue factor has been proposed as a therapeutic modality (European Patent No. 0 266 993 B1), and the use of antibodies that specifically recognize TF at the site of thrombogenesis is currently viewed as a promising strategy for treating various thrombotic disorders. In fact, *in vivo* studies with anti-TF monoclonal antibodies demonstrated efficient anticoagulant activities (Levi et al., [1994] J. Clin. Invest. 93, 114-120; Taylor et al., [1991] Circulatory Shock 33, 127-134; Himber et al., [1997] Thromb Haemostasis 78, 1142-1149; Pawashe et al., [1994] Circ. Res. 74, 56-63; Ragni et al., [1996] Circulation 93, 1913-1918; Jang et al., [1992] Arterioscl. Thromb. 12, 948-954; Thomas et al., [1993] Stroke 24, 847-854; Golino et al., [1996] Nature Med. 2, 35-40). The use of a CDR-grafted anti-hTF antibody has been described for the

attenuation or prevention of tissue factor mediated coagulation (International Publication No. WO 96/40921).

However, the precise TF binding sites of the antibodies used in the foregoing *in vivo* studies, with the exception of the antibody used by Levi et al., *supra*, are not known. The location of the antibody binding epitope may represent a critical factor in determining the inhibitory potencies of antibodies, because the cofactor function of TF involves several defined regions of the TF molecule. As a cofactor for factor VIIa (FVIIa), the cell surface exposed TF immobilizes FVII/FVIIa to the cell membrane thereby stabilizing the overall conformation of FVIIa (Waxman et al., [1993] Biochemistry 32, 3005-3012). The binding to TF also leads to the correct spatial orientation of the catalytic domain and the positioning of the active site in respect to the phospholipid membrane (McCallum et al., [1997] J. Biol. Chem. 272, 30160-30166; Banner et al., [1996] Nature 380, 41-46). Most of the TF-FVIIa contact surface area is provided by the FVIIa light chain interaction with TF. A smaller, yet critical contact surface lies between the N-terminal TF domain and the FVIIa catalytic domain. This contact is thought to play a main role in the enhancement of catalysis towards small synthetic as well as to macromolecular substrates (Dickinson et al., [1996] Proc. Natl. Acad. Sci. USA 93, 14379-14384; Dickinson and Ruf, [1997] J. Biol. Chem. 272, 19875-19879). In addition, TF participates in direct interaction with substrates (Huang et al., [1996] J. Biol. Chem. 271, 21752-21757) via residues K165 and K166 (Huang et al., *supra*; Ruf et al., [1992] J. Biol. Chem. 267, 6375-6381; Roy et al., [1991] J. Biol. Chem. 266, 22063-22066; Kelley et al., [1995] Biochemistry 34, 10383-10392), and neighboring residues (Ruf et al., [1992] J. Biol. Chem. 267: 22206-22210) in the C-terminal domain of TF. To add to this complex cofactor-enzyme-substrate interplay, recent observations suggested that the  $\gamma$ -carboxyglutamic acid-rich (Gla) domain of FVIIa contributes to substrate interaction (Huang et al., [1996] *supra*; Ruf et al., [1991] J. Biol. Chem. 266, 15719-15725; Martin et al., [1993] Biochemistry 32, 13949-13955; Ruf et al., [1999] Biochemistry 38, 1957-1966). Thus, anti-TF antibodies by virtue of their epitope location may interfere with one or several of these TF-mediated processes, which could translate into differences in their anticoagulant effectiveness. Such antibody epitope-dependent differences in potencies could be exacerbated under non-equilibrium conditions, which most likely prevail under therapeutic conditions. In this setting, antibody and the substrates circulating in blood would simultaneously interact with exposed TF.

In view of the limited characterization of most anti-TF antibodies known in the art, and the complexity of the mechanism by which TF exerts its thrombotic activity, it has so far been impossible to reliably engineer anti-TF antibodies with enhanced anticoagulant potency.

It is an objective of the present invention to determine which characteristics of anti-TF antibodies have the most profound effect on their anticoagulant properties. It is another objective, to design anti-TF antibodies with enhanced anticoagulant potency. Another objective is to provide combinations of anti-TF antibodies with at least one additional anticoagulant or anti-platelet agent with enhanced anticoagulant and/or anti-thrombotic efficacy.

#### Summary of the Invention

The present invention is based in part on the experimental finding that potency differences between various anti-TF antibodies can be explained by the location of the TF epitopes to which the antibodies bind and consequently, by the particular mode of inhibition. Anti-TF antibodies which bind to an epitope overlapping with the C-terminal macromolecular substrate-binding region of TF, and thus interfere with the TF-substrate interaction, are the most potent anticoagulant agents. This finding permits, for the first time, the purposeful design of anti - TF antibodies with high potency to treat or inhibit thrombosis.

Another aspect of the invention, involves a method for enhancing antithrombotic efficacy of a treatment comprising administering a dose of an anti-TF antibody and a dose of at least one additional anticoagulant or antiplatelet agent. The administration of the treatment is effective to enhance antithrombotic efficacy and preferably, minimize an increase in bleeding risk of the treatment compared to administration of the antibody or agent alone. Some current treatments of thrombotic conditions, such as acute coronary syndrome, involve administration of multiple agents including aspirin, heparin and a GP IIb/IIIa inhibitor. These agents act to inhibit thrombus formation at later stages in the coagulation pathway by inhibiting either the action of thrombin or platelet aggregation. Therapeutically effective doses of heparin and/or GP IIb/IIIa inhibitors can have adverse consequences, such as increased bleeding risk and/or thrombocytopenia. While not meant to limit the invention, it is believed that anti-TF antibodies act at a different point in the coagulation pathway and inhibit the formation of thrombin. Administration of anti-TF antibodies in combination with at least one additional anticoagulant and/or anti-platelet agent, such as heparin, enhance the efficacy of the combination and preferably, minimize an increase in bleeding risk compared to administration with the antibody or agent alone.

The invention also includes a method for treating a TF-VIIa related disease or disorder comprising administering a treatment comprising a dose of an anti-TF antibody and a dose of at least one additional anticoagulant or anti-platelet agent. Preferably, the anti-TF antibody is an antibody which binds to an epitope comprising at least part of the C-terminal

5 macromolecular substrate-binding region of tissue factor. Preferably, the anticoagulant is an agent that is not an anti-TF antibody and that inhibits formation or growth of a thrombus selected from the group consisting of heparin, low molecular weight heparin, pentasaccharide thrombin inhibitors, direct thrombin inhibitors and mixtures thereof. The antiplatelet agents are, preferably, cyclooxygenase inhibitors, ADP receptor antagonists, phosphodiesterase  
10 inhibitors and GP IIb/IIIa inhibitors. The treatment has enhanced antithrombotic efficacy and preferably, minimizes any increase in bleeding risk compared to administration of the antibody or agent alone.

Another aspect of invention concerns a method for identifying anti-tissue factor (anti-TF) antibodies with enhanced anticoagulant potency, comprising (a) subjecting a plurality of  
15 anti-TF antibodies to epitope mapping, and (b) selecting antibodies binding to an epitope comprising at least part of the C-terminal macromolecular substrate-binding region of tissue factor (TF). The tissue factor is preferably human, and the macromolecular substrate preferably is Factor X (FX) or Factor IX (FIX). In a particularly preferred embodiment, the antibody selected recognizes an epitope which includes a TF region directly interacting with  
20 substrate factor FX or FIX, preferably by binding to a site which prevents or blocks association of TF with a Gla domain of the substrate factor. In another preferred embodiment, the antibody selected binds an epitope comprising residues K165, K166 and K201 of hTF. In yet another preferred embodiment, the epitope further comprises residues N199, R200 and I152 of hTF. In a further preferred embodiment, the epitope additionally comprises residue  
25 Y156 of hTF. In a particular embodiment, the method is used to identify antibodies that bind essentially to the same hTF epitope as any of antibodies D3, 5G6 and TF8-5G9. In some instances, it might be advantageous to select antibodies that have the binding properties specified above, and do not interfere with the association of hTF and Factor VIIa (FVIIa). All antibodies identified in accordance with the present invention may be poly- or monoclonal  
30 antibodies (as hereinafter defined), and may be rodent, (e.g. murine), rabbit, humanized or human antibodies.

The invention also covers compositions comprising the antibodies identified in accordance with the present invention, and methods of using such antibodies to block a TF-FVIIa mediated or associated process or event, or to prevent or treat a TF-FVIIa related

disease or disorder, including but not limited to, thrombotic and coagulopathic disorders. Compositions may further comprise at least one additional anticoagulant and/or anti-platelet agent. Preferably, the anticoagulant agent is an agent that inhibits the formation or growth of a thrombus. The antiplatelet agent, preferably, is a GP IIb/IIIa inhibitor.

5 In another aspect, the invention concerns a method for producing an antibody having enhanced anticoagulant potency, comprising raising antibodies against an antigen comprising at least part of the C-terminal macromolecular substrate binding region of tissue factor (TF). Again, the antibodies may be poly- or monoclonal antibodies (as hereinafter defined), including rodent, e.g. murine, humanized and human antibodies. In a preferred embodiment,  
10 the antibodies are raised against an antigen comprising the entire C-terminal macromolecular substrate-binding region of TF, preferably human TF (hTF). Preferably, the antigen used to raise the antibodies comprises residues K165, K166 and K201, and optionally residues N199, R200 and I152 of hTF. The antigen may additionally contain residue Y156 of hTF.

In yet another aspect, the invention concerns an anti-tissue factor (anti-TF) antibody  
15 heavy chain variable domain comprising the amino acid sequence of SEQ ID NO: 1 (VH SEQUENCE OF MURINE D3, Figure 8) or SEQ ID NO: 2 (VH SEQUENCE OF HUMANIZED D3H44, Figure 8).

In a further aspect, the invention concerns an anti-tissue factor (anti-TF) light chain variable domain comprising the amino acid sequence of SEQ ID NO: 3 (VL SEQUENCE OF  
20 MURINE D3, Figure 9) or SEQ ID NO: 4 (VL SEQUENCE OF HUMANIZED D3H44, Figure 9).

In a further aspect, the invention concerns an anti-tissue factor (anti-TF) heavy chain variable domain comprising the amino acid sequence of SEQ ID NO: 5 (VH SEQUENCE OF  
25 MURINE 5G6 - Figure 15).

In a different aspect, the invention concerns an anti-tissue factor (anti-TF) light chain variable domain comprising the amino acid sequence of SEQ ID NO: 6 (VL SEQUENCE OF  
30 MURINE 5G6 - Figure 15).

In another aspect, the invention concerns isolated nucleic acid comprising a sequence encoding an anti-tissue factor (anti-TF) antibody heavy chain variable domain of SEQ ID NO:  
30 1, 2 or 5.

In yet another aspect, the invention concerns isolated nucleic acid comprising a sequence encoding an anti-tissue factor (anti-TF) antibody light chain variable domain of  
SEQ ID NO: 3, 4 or 6.

In a further aspect, the invention concerns a vector comprising, and capable of expressing, a nucleic acid as hereinabove defined, a recombinant host cell transformed with such vector, a cell culture comprising such recombinant host cell, and a method for expressing said nucleic acid to produce the encoded polypeptide.

5           The invention also concerns a humanized anti-tissue factor (anti-TF) antibody comprising a heavy and a light chain variable domain, wherein the heavy chain variable domain comprises hypervariable regions CDR-H1 having the sequence of GFNIKEYYM (SEQ ID NO:7), CDR-H2 having the sequence of LIDPEQGNTIYDPKFQD (SEQ ID NO:8) and CDR-H3 having the sequence of DTAAYFDY (SEQ ID NO:9). In a particular  
10           embodiment, the humanized anti-TF antibody of the present invention has a light chain variable domain comprising hypervariable regions CDR-L1 having the sequence of RASRDIKSYLN (SEQ ID NO: 10), CDR-L2 having the sequence of YATSLAE (SEQ ID NO:11) and CDR-L3 having the sequence of LQHGESPT (SEQ ID NO:12). Preferably, both the heavy and light chain hypervariable regions are provided in a human framework  
15           region. Particular antibodies that are within the scope of the present invention include, without limitation: (a) murine antibody D3 (D3Mur), (b) humanized antibody D3H44, (c) murine antibody 5G6, and (d) antibodies specifically binding essentially the same epitope as any one of antibodies (a) - (c).

20           In another aspect, the invention concerns isolated nucleic acid comprising a sequence encoding a humanized anti-TF antibody heavy or light chain variable domain as hereinabove defined, a vector comprising and capable of expressing such nucleic acid, a recombinant host cell transformed with such vector, a cell culture comprising such recombinant host cell, and a method for expressing said nucleic acid to produce the encoded polypeptide.

25           In another aspect, the invention concerns a composition comprising an anti-tissue factor (anti-TF) antibody identifiable by the method of claim 1, in admixture with a pharmaceutically acceptable carrier. The antibody preferably is an anti-hTF antibody, and is preferably humanized or human. The composition may, for example, comprise an antibody selected from the group consisting of (a) murine antibody D3 (D3Mur), (b) humanized antibody D3H44, (c) murine antibody 5G6, and (d) an antibody specifically binding  
30           essentially the same epitope as any one of antibodies (a)-(c), in admixture with a pharmaceutically acceptable carrier.

          The invention also concerns diagnostic methods, diagnostic kits and articles of manufacture comprising one or more antibodies of the present invention, optionally in

combination with one or more further active ingredients useful in the desired diagnostic or therapeutic application.

#### Brief Description of the Drawings

Fig. 1 Inhibitory characteristics of anti-tissue factor antibodies. (a) Amidolytic activity of sTF:FVIIa towards the small synthetic substrate Chromozym t-PA.

Antibodies were incubated together with sTF (10nM) and FVIIa (10nM) in HBS buffer, 5mM CaCl<sub>2</sub> for 15 min. Chromozym t-PA (0.5mM) was added and the rates of substrate cleavage were measured. The results were expressed in percent of control rates (average of 3 experiments  $\pm$  SD). (b) Prolongation of human plasma clotting by anti-TF antibodies. The antibodies were incubated with TF reagent (Innovin) for 15 min, then added to human citrated plasma. The increase in clotting times is reported as the ratio of clotting times in the presence of antibody and baseline values. The results are the average of two independent experiments. (filled circle) D3, (open circle) D3 Fab, (x) 5G6, (filled square) 7G11, (open square) 6B4, (filled triangle) HTF1, (open triangle) isotype-matched control Ab.

Fig. 2 Inhibition of fibrinopeptide A (FPA) generation by anti-TF antibodies in a human ex-vivo blood flow system. The FPA concentrations in plasma are expressed in percent of control values. Each value is the average of 3-8 experiments except for the highest concentrations of D3 Fab and 5G6 (n=1 and n=2, resp.) and for two 6B4 concentrations (n=2 for 1.5 $\mu$ g/ml and 15 $\mu$ g/ml). The average of all control values (n=36) was 1348.6  $\pm$  46.1 ng/ml ( $\pm$ SEM). filled circle, D3; open circle, D3 Fab; x, 5G6; open square, 6B4; filled triangle, HTF1.

Fig. 3 Effects of anti-tissue factor antibodies on TF-dependent FX activation in human plasma. The inhibited rates of FXa generation during the initial phase of 45 sec were calculated and expressed as fractional activity (vi/v<sub>0</sub>). (filled circle) D3, (x) 5G6, (filled square) 7G11, (open square) 6B4, (filled triangle) HTF1, (open triangle) isotype-matched control Ab.

Fig. 4 Prolongation of prothrombin time (PT) by anti-tissue factor antibodies. Prolongation of clotting times are reported as the ratio of clotting times in the presence of antibody and baseline values. The results are the average of two independent experiments. (filled circle) D3, (x) 5G6, (filled square) 7G11, (open square) 6B4, (filled triangle) HTF1.

Fig. 5 Effects of sTF mutations on antibody binding. The changes in binding affinities are expressed as the  $K_D$  ratios of sTF mutants and sTF wildtype ( $K_D(\text{mut})/K_D(\text{wt})$ ). The  $K_D$  values were calculated from surface plasmon resonance measurements with immobilized antibodies.

Fig. 6 Localization of the antibody epitopes on the crystal structure of the sTF:FVIIa complex. FVIIa is colored with the light chain in orange and the heavy chain in green. The active site inhibitor (D-Phe-L-Phe-Arg chloromethyl ketone) is in red and the calcium atoms in yellow. Tissue factor (grey) is in a solvent accessible representation and the antibody epitope residues are shown in red color. The figures were produced using Insight II (MSI, San Diego).

Fig. 7 Crystal structure of murine D3 F(ab). Ribbon diagram of VH (dark grey) and VL (light grey) backbones is shown. Side chains of residues changed or investigated during the humanization are shown and labeled; side chain nitrogens and oxygens are dark grey. Spheres represent two internal water molecules.

Fig. 8 Sequence alignment of VH domains of murine D3 (D3Mur), consensus human subgroup III (HumVHIII), and humanized D3H44. CDRs are underlined and differences between sequences are noted by \*. CDR's are defined according to Kabat et al., Sequences of Proteins of Immunological Interest, 5<sup>th</sup> Ed. Public Health Service, National Institute of Health, Bethesda, MD (1991) except for CDR-H1 which was defined using a combination of CDR-H1 definitions from Kabat et al. (*supra*) and Chothia et al., Nature 342:877-833 (1989), i.e., CDR-H1 was defined as extending from residues H26-H35 in the heavy chain.

Fig. 9 Sequence alignment of VL domains of murine D3 (D3Mur), consensus human kappa subgroup I (HumκI), and humanized D3H44. CDRs are underlined and differences between sequences are noted by \*. Residue numbering is according to Kabat et al. (1991), *supra*.

Fig. 10 Inhibition of the rate of FX activation by antibody F(ab) using membrane TF(mTF):FVIIa complex. The antibodies were incubated with mTF and FVIIa for 20 min before FX was added. Aliquots were taken at different time points and quenched in 20 mM EDTA. In the second stage of the assay, a chromogenic substrate S-2765 was added and the amidolytic activity measured at 405 nm on a kinetic microplate reader. The initial rates are calculated and the inhibition expressed as fractional rates ( $v_i/v_o$ ) of FXa generation.



Fig. 11 Inhibition of the rate of F.IX activation by antibody F(ab) using membrane TF(mTF):FVIIa complex. The antibodies were incubated with mTF and FVIIa for 20 min before F.IX was added. Aliquots were taken at different time points and quenched in 30 mM EDTA-60% ethyleneglycol. In the second stage of the assay, a chromogenic substrate #299 was added and the amidolytic activity measured at 405 nm on a kinetic microplate reader. The initial rates are calculated and the inhibition expressed as fractional rates ( $v_i/v_o$ ) of FIXa generation.

Fig. 12 Effects of antibody F(ab) and F(ab')<sub>2</sub> on prothrombin time (PT) in human plasma. *E. coli* expressed F(ab) of D3C2 (chimeric F(ab)), D3H18, D3H31, D3H44 and F(ab')<sub>2</sub> of D3H44 were incubated in human plasma for 5 min. Clotting was initiated by addition of human tissue factor reagent (Innovin). Clotting times were measured on an ACL 300 instrument. The prolongation of the clotting time is expressed as the ratio of inhibited clotting (with antibody) and uninhibited clotting time (buffer control). The indicated antibody concentrations are the concentrations in plasma.

Fig. 13 Amino acid sequence of human tissue factor (hTF) (SEQ ID NO: 13).

Fig. 14 Ribbon representation of the structure of the extracellular portion of human tissue factor.

Fig. 15 Heavy chain variable domain sequence of murine anti-TF antibody 5G6 (SEQ ID NO: 5). Light chain variable domain sequence of murine anti-TF antibody 5G6 (SEQ ID NO: 6).

Fig. 16 Binding of anti-tissue factor antibodies to tissue factor. IgG1, IgG2, IgG4 and IgG4b.

Fig. 17 Prolongation of human plasma clotting time (PT) for the full length versions and Fab and F(ab')<sub>2</sub> versions of D3H44.

Fig. 18 Comparison of potency of AP-1 in *in vitro* PT, 2st-PT, and APTT coagulation assays. Various amounts of AP-1 were tested *in vitro* in various assays for effect on coagulation time: PT assay (—•—); APTT assay (—Δ—); and 2st-PT (—○—).

Fig. 19 *In vitro* characterization of inhibition of amidolytic activity of rabbit sTF/human VIIa complex by various amounts of AP-1 (—○—) (IC<sub>50</sub> 33nM) compared to various amounts of anti-human tissue factor antibody 6B4 (—□—).

Fig. 20 Effects of various doses of anti-rabbit tissue factor antibody AP-1, unfractionated heparin (UFH), and a platelet GPGP IIb/IIIa inhibitor (G4120) on thrombus formation and bleeding time in a rabbit model of arterial thrombosis induced by

severe balloon damage and high shear stress. Thrombus mass is reported as a percentage of the thrombus mass of the saline control (%C). Cuticle bleeding time is reported as a ratio of the post treatment cuticle bleeding time over the pretreatment cuticle bleeding time (post/pre). Panel A shows the effects of administration of different doses of AP-1 on thrombus mass (—●—) and cuticle bleeding times (—□—). Panel B shows the effects of administration of various doses of G4120 on thrombus mass (—●—) and cuticle bleeding time (—□—). Panel C shows the effects of administration of various doses of UFH on thrombus mass (—●—) and cuticle bleeding time (—□—). The asterisks denote significant differences as follows: \* $p \leq 0.05$ , \*\* $p \leq 0.01$ , \*\*\* $p \leq 0.001$  versus saline by Mann Whitney (thrombus mass) or Fisher's PLSD test (CBT).

Fig. 21 Effects of administration of combinations of AP-1 and G4120 with two different doses of UFH on thrombus mass and cuticle bleeding times in the rabbit model of arterial thrombosis as compared to UFH alone are shown. Panel A shows the effect of administration of combinations of agents on thrombus mass compared to UFH alone. Panel B shows the effect of administration of the same agents at the same doses on cuticle bleeding times compared to UFH alone. Data are mean  $\pm$  sem. The doses of UFH were 25 U/kg and 0.25 U/kg/min. and 50 U/kg and 0.5 U/kg/min. The dose of AP-1 was 0.5 mg/kg and the G4120 dose was 37.5  $\mu$ g/kg + 0.5  $\mu$ g/kg/min. The numbers in the bars represent the percent change from the respective UFH monotherapy. The asterisks denote significant differences as follows: \* $p \leq 0.05$ , \*\* $p \leq 0.01$  versus UFH by Mann-Whitney (Thrombus Mass) or T-test (CBT).

## Detailed Description of the Preferred Embodiments

### Definitions

Abbreviations used throughout the description include: FIXa for Factor IXa; FXIa for Factor XIa; FXa for Factor Xa; TF for tissue factor; FVII for zymogen factor VII; FVIIa for Factor VIIa; TF-FVIIa for tissue factor-Factor VIIa complex; FVII/FVIIa for FVII and/or FVIIa; sTF for soluble tissue factor composed of the extracellular domain residues 1-219 in the hTF sequence of Figure 13 (SEQ ID NO:13); hTFAA, the sTF variant containing Lys to Ala substitutions at positions 165 and 166 of the native hTF sequence; TF7I-C for the Kunitz type TF-FVIIa inhibitor of the same name in Dennis et al., (1994) J. Biol. Chem.

269(35):22129-22136;  $K_D$  for equilibrium dissociation constant; PT for prothrombin time; APTT for activated partial thromboplastin time.

The term "anticoagulant potency" is used to refer to the ability of a substance, e.g. an antibody herein, to prevent, inhibit or prolong blood coagulation in an *in vitro* or *in vivo* assay of blood coagulation. Blood coagulation assays are known in the art and include, for example, prothrombin time assays such as those described in Example 1 herein, the human *ex vivo* thrombosis model described by Kirchhofer *et al.*, Arterioscler. Thromb. Vasc. Biol. 15, 1098-1106 (1995); and Kirchhofer *et al.*, J. Clin. Invest. 93, 2073-2083 (1994), and assays based on the measurement of Factor X activation in human plasma, and in the examples as described in the present application.

The anticoagulant potency of an antibody of the present invention is "enhanced", if its ability to prevent, inhibit or prolong blood coagulation surpasses the ability of an anti-TF antibody that binds to a TF epitope other than an epitope comprising at least part of the C-terminal macromolecular substrate-binding region of TF, as determined in a standard *in vivo* or *in vitro* assay of blood coagulation, such as the assays referred to above. Preferably, the anti-TF with enhanced anticoagulant potency achieves the same effect (prevention, inhibition or prolongation) at a lower dose and/or in a shorter time than a reference antibody binding to a different TF epitope. Preferably, the difference between the potency of an antibody within the scope of the present invention and a reference antibody is at least about 1.5-fold, more preferably at least about 2-fold, even more preferably at least about 3-fold, most preferably at least about 5-fold, as determined by side-by-side comparison in a selected standard blood coagulation assay.

The term "antithrombotic efficacy" of an anti-TF antibody or other agent is used to refer to the ability of a substance, to prevent, inhibit or reduce the incidence and/or severity of a thrombus as compared to a control. Systems for measuring antithrombotic efficacy are known in the art and include the rabbit model of arterial thrombosis as described by Refino *et al.*, Arterio. Thromb. Vasc. Biol., 517-522 [2002] and guinea pig and rat models as described in Refino *et al.*, Thrombosis and Hemostasis, 82:1188-1195 [1999]. Prevention, reduction or inhibition of a thrombus can be determined by measuring a decrease in thrombus mass as described in Refino *et al.* (op.cite). An optimal dose is a dose that inhibits or reduces the incidence and/or severity of a thrombus about 80% to 100%. A suboptimal dose is a dose that results in less than about 80% reduction or inhibition of the incidence and/or severity of a thrombus.

The antithrombotic efficacy of an anti tissue factor antibody or agent is "enhanced" if, e.g. when administered in combination, the incidence and/or severity of a thrombus is prevented, reduced or inhibited more than when the same dose of anti-TF antibody, anticoagulant or antiplatelet agent is administered alone as determined in a standard assay.

5 Preferably, the anti-TF antibody in combination with at least one additional agent achieves the same effect at a lower dose and/or in a shorter time compared to the antibody or agent alone. More preferably, the combination of the anti-TF antibody and at least one additional anticoagulant or anti-platelet agent provides for at least about 30% or greater inhibition or reduction in incidence and/or severity of a thrombus compared to the same dose of antibody  
10 or agent alone.

The administration of a treatment comprising a dose of an anti-TF antibody and a dose of at least one additional anticoagulant or anti-platelet agent preferably provides a "minimal increase in bleeding risk". A "minimal increase in bleeding risk" refers to little or no significant increase in the incidence and/or severity of bleeding events compared to control.

15 Methods for assessing bleeding risk are known to those of skill in the art and include standard bleeding time assays, such as cuticle bleeding time, surgical blood loss, or template bleeding in animal models and the TIMI criteria as described in Menon et al, supra. Whether bleeding risk is significantly increased can readily be determined by one of skill in the art using accepted statistical analysis. For example, when bleeding risk is assessed for the combination  
20 treatments as described herein, bleeding times are prolonged about 25% or less compared to the administration of the antibody or agent alone. When bleeding risk is assessed clinically, a minimal increase in bleeding risk would be an increase of minor bleeding events of about 20% or less compared to control and/or an increase in major bleeding events of about 10% or less compared to control. In some cases, the bleeding risk may be decreased compared to  
25 control. Preferably, the dose of the antibody and/or agent is a dose that does not significantly prolong the bleeding time or significantly increase the bleeding risk compared to control.

The "C-terminal macromolecular substrate-binding region of TF" is defined as the C-terminal region within the three-dimensional structure of TF that is responsible for the interaction of TF with its macromolecular substrate Factor X (FX) or Factor IX (FIX). In  
30 hTF, the FX interaction region is located within the second FNIII module of the extracellular domain of hTF as defined by Muller et al., J. Mol. Biol. 256, 144-159 (1996), including the  $\beta$ 8<sub>A</sub> to  $\beta$ 16<sub>G</sub> shown in Figure 3 of Muller et al., supra, and in Figure 14 herein. The main portion of the macromolecular substrate binding region of hTF includes residues Lys 165, Lys 166 (Roy et al., (1991) supra; Ruf et al., (1992) J. Biol. Chem. 267:6375-6381;

Huang et al., (1996) J. Biol. Chem. 271:21752-21757), Tyr 157, Lys 159, Ser 163, Gly 164, Tyr 185 (Kirchhofer et al., (1999) Thromb. Haemost. Suppl. 300, abstract; Kirchhofer et al., (2000) Biochemistry, 39:7380-7387). There are additional hTF residues which contribute to F.X interaction such as Tyr 156, Trp 158, Lys 169, Asn 173, Glu 174, Asn 199, Arg 200, Lys 201 and Asp 204 . The substrate F.IX interacts with about the same hTF region, the main interaction region (Lys 165, Lys 166, Tyr 157, Lys 159, Ser 163, Gly 164, Tyr 185) being identical to that for F.X. The only difference observed concerned the hTF residues Trp 158 and Asp 204 both of which may be less important for F.IX interaction than for F.X interaction.

The term "epitope" is used to refer to binding sites for (monoclonal or polyclonal) antibodies on protein antigens.

Antibodies which bind to the C-terminal macromolecular substrate-binding region of TF are identified by "epitope mapping." There are many methods known in the art for mapping and characterizing the location of epitopes on proteins, including solving the crystal structure of an antibody-antigen complex, competition assays, gene fragment expression assays, and synthetic peptide-based assays, as described, for example, in Chapter 11 of Harlow and Lane, Using Antibodies, a Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1999. Competition assays are discussed below. According to the gene fragment expression assays, the open reading frame encoding the protein is fragmented either randomly or by specific genetic constructions and the reactivity of the expressed fragments of the protein with the antibody to be tested is determined. The gene fragments may, for example, be produced by PCR and then transcribed and translated into protein *in vitro*, in the presence of radioactive amino acids. The binding of the antibody to the radioactively labeled protein fragments is then determined by immunoprecipitation and gel electrophoresis. Certain epitopes can also be identified by using large libraries of random peptide sequences displayed on the surface of phage particles (phage libraries). Alternatively, a defined library of overlapping peptide fragments can be tested for binding to the test antibody in simple binding assays. The latter approach is suitable to define linear epitopes of about 5 to 15 amino acids.

An antibody binds "essentially the same epitope" as a reference antibody, when the two antibodies recognize identical or sterically overlapping epitopes. The most widely used and rapid methods for determining whether two epitopes bind to identical or sterically overlapping epitopes are competition assays, which can be configured in all number of different formats, using either labeled antigen or labeled antibody. Usually, the antigen is

immobilized on a 96-well plate, and the ability of unlabeled antibodies to block the binding of labeled antibodies is measured using radioactive or enzyme labels.

The term amino acid or amino acid residue, as used herein, refers to naturally occurring L amino acids or to D amino acids as described further below with respect to variants. The commonly used one- and three-letter abbreviations for amino acids are used herein (Bruce Alberts et al., *Molecular Biology of the Cell*, Garland Publishing, Inc., New York (3d ed. 1994)).

A "TF-FVIIa mediated or associated process or event", or equivalently, an "activity associated with plasma FVIIa", according to the present invention is any event which requires the presence of TF-FVIIa. The general mechanism of blood clot formation is reviewed by Ganong, in *Review of Medical Physiology*, 13th ed., Lange, Los Altos CA, pp. 411-414 (1987) and Bach (1988) *CRC Crit. Rev. Biochem.* 23(4):359-368. Coagulation requires the confluence of two processes, the production of thrombin which induces platelet aggregation and the formation of fibrin which renders the platelet plug stable. The process comprises several stages each requiring the presence of discrete proenzymes and procofactors. The process ends in fibrin crosslinking and thrombus formation. Fibrinogen is converted to fibrin by the action of thrombin. Thrombin, in turn, is formed by the proteolytic cleavage of prothrombin. This proteolysis is effected by FXa which binds to the surface of activated platelets and in the presence of FVa and calcium, cleaves prothrombin. TF-FVIIa is required for the proteolytic activation of FX by the extrinsic pathway of coagulation. Therefore, a process mediated by or associated with TF-FVIIa, or an activity associated with FVIIa includes any step in the coagulation cascade from the formation of the TF-FVII complex to the formation of a fibrin platelet clot and which initially requires the presence TF-FVIIa. For example, the TF-FVIIa complex initiates the extrinsic pathway by activation of FX to FXa, FIX to FIXa, and additional FVII to FVIIa. TF-FVIIa mediated or associated process, or FVIIa activity, can be conveniently measured employing standard assays such as those described in Roy, S., (1991) *J. Biol. Chem.* 266:4665-4668, and O'Brien, D., et al., (1988) *J. Clin. Invest.* 82:206-212 for the conversion of Factor X to Factor Xa in the presence of Factor VII and other necessary reagents.

A "TF-FVIIa related disease or disorder" is meant to include chronic thromboembolic diseases or disorders associated with fibrin formation including vascular disorders such as deep venous thrombosis, arterial thrombosis, acute coronary syndrome, including unstable angina (UA) and non-ST-segment elevation myocardial infarction (NSTEMI), stroke, tumor metastasis, thrombolysis, arteriosclerosis and restenosis following angioplasty, acute and

chronic indications such as inflammation, septic shock, septicemia, hypotension, adult respiratory distress syndrome (ARDS), disseminated intravascular coagulopathy (DIC) and other diseases. The TF-FVIIa related disorder is not limited to in vivo coagulopathic disorders such as those named above but includes ex vivo TF-FVIIa related processes such as coagulation that may result from the extracorporeal circulation of blood, including blood removed in-line from a patient in such processes as dialysis procedures, blood filtration, or blood bypass during surgery.

"Bleeding disorders" are characterized by a tendency toward hemorrhage, both inherited and acquired. Examples of such bleeding disorders are deficiencies of factors VIII, IX, or XI. Examples of acquired disorders include acquired inhibitors to blood coagulation factors e.g., factor VIII, von Willebrand factor, factors IX, V, XI, XII and XIII, hemostatic disorders as a consequence of liver disease which included decreased synthesis of coagulation factors, bleeding tendency associated with acute and chronic renal disease and hemostasis after trauma or surgery.

The terms "tissue factor protein" and "mammalian tissue factor protein" are used to refer to a polypeptide having an amino acid sequence corresponding to a naturally occurring mammalian tissue factor or a recombinant tissue factor as described below. Naturally occurring TF includes human species as well as other animal species such as rabbit, rat, porcine, non-human primate, equine, murine, and ovine tissue factor (see, for example, Hartzell et al., (1989) Mol. Cell. Biol., 9:2567-2573; Andrews et al., (1991) Gene, 98:265-269; and Takayenik et al., (1991) Biochem. Biophys. Res. Comm., 181:1145-1150). The amino acid sequence of human tissue factor is shown in Figure 13 (SEQ ID NO: 13). The amino acid sequence of the other mammalian tissue factor proteins are generally known or obtainable through conventional techniques.

As used herein, "treatment" is an approach for obtaining beneficial or desired clinical results. For purposes of this invention, beneficial or desired clinical results include, but are not limited to, alleviation of symptoms, diminishment of extent of disease, stabilized (i.e., not worsening) state of disease, delay or slowing of disease progression, amelioration or palliation of the disease state, and remission (whether partial or total), whether detectable or undetectable. "Treatment" can also mean prolonging survival as compared to expected survival if not receiving treatment. "Treatment" is an intervention performed with the intention of preventing the development or altering the pathology of a disorder. Accordingly, "treatment" refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already with the disorder as well as those in which

the disorder is to be prevented. Accordingly, "treatment" in the context of the present invention is an intervention performed with the intention of preventing a TF-FVIIa mediated or associated process or event, or a TF-FVIIa related disease or disorder, or a bleeding disorder, as hereinabove defined.

5 "Mammal" for purposes of treatment refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, horses, cats, cows, *etc.* Preferably, the mammal is human.

"Antibodies" (Abs) and "immunoglobulins" (Igs) are glycoproteins having the same structural characteristics. While antibodies exhibit binding specificity to a specific antigen,  
10 immunoglobulins include both antibodies and other antibody-like molecules that lack antigen specificity. Polypeptides of the latter kind are, for example, produced at low levels by the lymph system and at increased levels by myelomas.

"Native antibodies" and "native immunoglobulins" are usually heterotetrameric glycoproteins of about 150,000 daltons, composed of two identical light (L) chains and two  
15 identical heavy (H) chains. Each light chain is linked to a heavy chain by one covalent disulfide bond, while the number of disulfide linkages varies among the heavy chains of different immunoglobulin isotypes. Each heavy and light chain also has regularly spaced intrachain disulfide bridges. Each heavy chain has at one end a variable domain ( $V_H$ ) followed by a number of constant domains. Each light chain has a variable domain at one end  
20 ( $V_L$ ) and a constant domain at its other end; the constant domain of the light chain is aligned with the first constant domain of the heavy chain, and the light-chain variable domain is aligned with the variable domain of the heavy chain. Particular amino acid residues are believed to form an interface between the light- and heavy-chain variable domains.

The term "variable" refers to the fact that certain portions of the variable domains  
25 differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not evenly distributed throughout the variable domains of antibodies. It is concentrated in three segments called hypervariable regions both in the light chain and the heavy chain variable domains. The more highly conserved portions of variable domains are called the framework  
30 region (FR). The variable domains of native heavy and light chains each comprise four FRs (FR1, FR2, FR3 and FR4, respectively), largely adopting a  $\beta$ -sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some cases forming part of, the  $\beta$ -sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to



the formation of the antigen-binding site of antibodies (see Kabat *et al.*, *Sequences of Proteins of Immunological Interest*, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD (1991), pages 647-669). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody-dependent cellular toxicity.

The term "hypervariable region" when used herein refers to the amino acid residues of an antibody which are responsible for antigen-binding. The hypervariable region comprises amino acid residues from a "complementarity determining region" or "CDR" (*i.e.* residues 24-34 (L1), 50-56 (L2) and 89-97 (L3) in the light chain variable domain and 31-35 (H1), 50-65 (H2) and 95-102 (H3) in the heavy chain variable domain; Kabat *et al.*, *Sequences of Proteins of Immunological Interest*, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD. (1991)) and/or those residues from a "hypervariable loop" (*i.e.* residues 26-32 (L1), 50-52 (L2) and 91-96 (L3) in the light chain variable domain and 26-32 (H1), 53-55 (H2) and 96-101 (H3) in the heavy chain variable domain; Chothia and Lesk *J. Mol. Biol.* 196:901-917 (1987)). "Framework" or "FR" residues are those variable domain residues other than the hypervariable region residues as herein defined.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')<sub>2</sub> fragment that has two antigen-combining sites and is still capable of cross-linking antigen.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This region consists of a dimer of one heavy chain and one light chain variable domain in tight, non-covalent association. It is in this configuration that the three hypervariable regions of each variable domain interact to define an antigen-binding site on the surface of the V<sub>H</sub>-V<sub>L</sub> dimer. Collectively, the six hypervariable regions confer antigen-binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three hypervariable regions specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxyl terminus of the heavy chain CH1 domain including one or more cysteine(s) from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. F(ab')<sub>2</sub>

antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The "light chains" of antibodies (immunoglobulins) from any vertebrate species can be assigned to one of two clearly distinct types, called kappa ( $\kappa$ ) and lambda ( $\lambda$ ), based on the amino acid sequences of their constant domains.

Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), *e.g.*, IgG1, IgG2, IgG3, IgG4, IgA1, and IgA2. The heavy-chain constant domains that correspond to the different classes of immunoglobulins are called  $\alpha$ ,  $\delta$ ,  $\epsilon$ ,  $\gamma$ , and  $\mu$ , respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known.

The term "antibody" herein is used in the broadest sense and specifically covers monoclonal antibodies (including full length monoclonal antibodies), polyclonal antibodies, multispecific antibodies (*e.g.*, bispecific antibodies), and antibody fragments so long as they exhibit the desired biological activity.

"Antibody fragments" comprise a portion of a full length antibody, generally the antigen binding or variable domain thereof. Examples of antibody fragments include Fab, Fab', F(ab')<sub>2</sub>, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, *i.e.*, the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler *et al.*, *Nature* 256:495 (1975), or may be made by recombinant DNA methods (see, *e.g.*, U.S. Patent No. 4,816,567). The "monoclonal antibodies" may also be isolated from phage antibody libraries using the techniques described

in Clackson *et al.*, *Nature* 352:624-628 (1991) and Marks *et al.*, *J. Mol. Biol.* 222:581-597 (1991), for example.

The monoclonal antibodies herein specifically include "chimeric" antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U.S. Patent No. 4,816,567; and Morrison *et al.*, *Proc. Natl. Acad. Sci. USA* 81:6851-6855 (1984)).

"Humanized" forms of non-human (*e.g.*, murine) antibodies are chimeric antibodies which contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which hypervariable region residues of the recipient are replaced by hypervariable region residues from a non-human species (donor antibody) such as mouse, rat, rabbit or nonhuman primate having the desired specificity, affinity, and capacity. In some instances, framework region (FR) residues of the human immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues which are not found in the recipient antibody or in the donor antibody. These modifications are made to further refine antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable regions correspond to those of a non-human immunoglobulin and all or substantially all of the FRs are those of a human immunoglobulin sequence. The humanized antibody optionally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details, see Jones *et al.*, *Nature* 321:522-525 (1986); Reichmann *et al.*, *Nature* 332:323-329 (1988); and Presta, *Curr. Op. Struct. Biol.* 2:593-596 (1992).

"Single-chain Fv" or "sFv" antibody fragments comprise the V<sub>H</sub> and V<sub>L</sub> domains of antibody, wherein these domains are present in a single polypeptide chain. Generally, the Fv polypeptide further comprises a polypeptide linker between the V<sub>H</sub> and V<sub>L</sub> domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv see Pluckthun in *The Pharmacology of Monoclonal Antibodies*, vol. 113, Rosenberg and Moore eds. Springer-Verlag, New York, pp. 269-315 (1994).

The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy chain variable domain ( $V_H$ ) connected to a light chain variable domain ( $V_L$ ) in the same polypeptide chain ( $V_H - V_L$ ). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger *et al.*, *Proc. Natl. Acad. Sci. USA* 90:6444-6448 (1993).

The expression "linear antibodies" when used throughout this application refers to the antibodies described in Zapata *et al.* *Protein Eng.* 8(10):1057-1062 (1995). Briefly, these antibodies comprise a pair of tandem Fd segments ( $V_H - C_H1 - V_H - C_H1$ ) which form a pair of antigen binding regions. Linear antibodies can be bispecific or monospecific.

#### Methods for carrying out the invention

##### A. Antibody preparation

Methods for humanizing nonhuman TF antibodies and generating variants of anti-TF antibodies are described in the examples below. In order to humanize an anti-TF antibody, the nonhuman antibody starting material is prepared. Where a variant is to be generated, the parent antibody is prepared. Exemplary techniques for generating such nonhuman antibody starting material and parent antibodies will be described in the following sections.

##### (i) *Polyclonal antibodies*

Methods of preparing polyclonal antibodies are known in the art. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized, such as serum albumin, or soybean trypsin inhibitor. Examples of adjuvants which may be employed include Freund's complete adjuvant and MPL-TDM.

##### (ii) *Monoclonal antibodies*

Monoclonal antibodies may be made using the hybridoma method first described by Kohler *et al.*, *Nature*, 256:495 (1975), or may be made by recombinant DNA methods (U.S. Patent No. 4,816,567).

In the hybridoma method, a mouse or other appropriate host animal, such as a hamster or macaque monkey, is immunized as hereinabove described to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the protein

used for immunization. Alternatively, lymphocytes may be immunized *in vitro*.

Lymphocytes then are fused with myeloma cells using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding, *Monoclonal Antibodies: Principles and Practice*, pp.59-103, [Academic Press, 1986]).

5           The hybridoma cells thus prepared are seeded and grown in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, parental myeloma cells. For example, if the parental myeloma cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine  
10 (HAT medium), which substances prevent the growth of HGPRT-deficient cells.

Preferred myeloma cells are those that fuse efficiently, support stable high-level production of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. Among these, preferred myeloma cell lines are murine myeloma lines, such as those derived from MOP-21 and M.C.-11 mouse tumors available  
15 from the Salk Institute Cell Distribution Center, San Diego, California USA, and SP-2 or X63-Ag8-653 cells available from the American Type Culture Collection, Rockville, Maryland USA. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor, *J. Immunol.*, 133:3001 (1984); Brodeur *et al.*, *Monoclonal Antibody Production Techniques and*  
20 *Applications*, pp. 51-63, Marcel Dekker, Inc., New York, [1987]).

Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against the antigen. Preferably, the binding specificity of monoclonal antibodies produced by hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked  
25 immunoabsorbent assay (ELISA).

The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson *et al.*, *Anal. Biochem.*, 107:220 (1980).

After hybridoma cells are identified that produce antibodies of the desired specificity, affinity, and/or activity, the cells may be subcloned by limiting dilution procedures and  
30 grown by standard methods (Goding, *Monoclonal Antibodies: Principles and Practice*, pp.59-103 (Academic Press, 1986)). Suitable culture media for this purpose include, for example, DMEM or RPMI-1640 medium. In addition, the hybridoma cells may be grown *in vivo* as ascites tumors in an animal.

The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

5 DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (*e.g.*, by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of the monoclonal antibodies). The hybridoma cells serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as *E. coli* cells,  
10 simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. Recombinant production of antibodies will be described in more detail below.

(iii) *Humanized antibodies*

15 Example 2 below describes procedures for humanization of an anti-TF antibody.

Generally, a humanized antibody has one or more amino acid residues introduced into it from a non-human source. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers  
20 [Jones et al., *Nature*, 321:522-525 (1986); Riechmann et al., *Nature*, 332:323-327 (1988); Verhoeven et al., *Science*, 239:1534-1536 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody.

(iv) *Amino acid sequence variants of antibodies*

25 Example 2 also describes methodologies for generating amino acid sequence variants of an anti-TF antibody with enhanced affinity relative to the parent antibody.

Amino acid sequence variants of the anti-TF antibody are prepared by introducing appropriate nucleotide changes into the anti-TF antibody DNA, or by peptide synthesis. Such variants include, for example, deletions from, and/or insertions into and/or substitutions of, residues within the amino acid sequences of the anti-TF antibodies of the examples herein.

30 Any combination of deletion, insertion, and substitution is made to arrive at the final construct, provided that the final construct possesses the desired characteristics. The amino acid changes also may alter post-translational processes of the humanized or variant anti-TF antibody, such as changing the number or position of glycosylation sites.

A useful method for identification of certain residues or regions of the anti-TF antibody that are preferred locations for mutagenesis is called "alanine scanning mutagenesis," as described by Cunningham and Wells, Science, 244:1081-1085 (1989). Here, a residue or group of target residues are identified (*e.g.*, charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with TF antigen. Those amino acid locations demonstrating functional sensitivity to the substitutions then are refined by introducing further or other variants at, or for, the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation *per se* need not be predetermined. For example, to analyze the performance of a mutation at a given site, ala scanning or random mutagenesis is conducted at the target codon or region and the expressed anti-TF antibody variants are screened for the desired activity.

Amino acid sequence insertions include amino- and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues. Examples of terminal insertions include an anti-TF antibody with an N-terminal methionyl residue or the antibody fused to an epitope tag. Other insertional variants of the anti-TF antibody molecule include the fusion to the N- or C-terminus of the anti-TF antibody of an enzyme or a polypeptide which increases the serum half-life of the antibody (see below).

Another type of variant is an amino acid substitution variant. These variants have at least one amino acid residue in the anti-TF antibody molecule removed and a different residue inserted in its place. The sites of greatest interest for substitutional mutagenesis include the hypervariable regions, but FR alterations are also contemplated. Conservative substitutions are shown in Table 1 under the heading of "preferred substitutions". If such substitutions result in a change in biological activity, then more substantial changes, denominated "exemplary substitutions" in Table 1, or as further described below in reference to amino acid classes, may be introduced and the products screened.

**Table 1**

Original Residue	Exemplary Substitutions	Preferred Substitutions
Ala (A)	val; leu; ile	val
Arg (R)	lys; gln; asn	lys

Asn (N)	gln; his; asp, lys; arg	gln
Asp (D)	glu; asn	glu
Cys (C)	ser; ala	ser
Gln (Q)	asn; glu	asn
Glu (E)	asp; gln	asp
Gly (G)	ala	ala
His (H)	asn; gln; lys; arg	arg
Ile (I)	leu; val; met; ala; phe; norleucine	leu
Leu (L)	norleucine; ile; val; met; ala; phe	ile
Lys (K)	arg; gln; asn	arg
Met (M)	leu; phe; ile	leu
Phe (F)	leu; val; ile; ala; tyr	tyr
Pro (P)	ala	ala
Ser (S)	thr	thr
Thr (T)	ser	ser
Trp (W)	tyr; phe	tyr
Tyr (Y)	trp; phe; thr; ser	phe
Val (V)	ile; leu; met; phe; ala; norleucine	leu

Substantial modifications in the biological properties of the antibody are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target



site, or (c) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties:

- (1) hydrophobic: norleucine, met, ala, val, leu, ile;
- (2) neutral hydrophilic: cys, ser, thr;
- 5 (3) acidic: asp, glu;
- (4) basic: asn, gln, his, lys, arg;
- (5) residues that influence chain orientation: gly, pro; and
- (6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these  
10 classes for another class.

Any cysteine residue not involved in maintaining the proper conformation of the humanized or variant anti-TF antibody also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond(s) may be added to the antibody to improve its stability  
15 (particularly where the antibody is an antibody fragment such as an Fv fragment).

A particularly preferred type of substitutional variant involves substituting one or more hypervariable region residues of a parent antibody (e.g. a humanized or human antibody). Generally, the resulting variant(s) selected for further development will have improved biological properties relative to the parent antibody from which they are generated.

20 A convenient way for generating such substitutional variants is affinity maturation using phage display. Briefly, several hypervariable region sites (e.g. 6-7 sites) are mutated to generate all possible amino substitutions at each site. The antibody variants thus generated are displayed in a monovalent fashion from filamentous phage particles as fusions to the gene III product of M13 packaged within each particle. The phage-displayed variants are  
25 then screened for their biological activity (e.g. binding affinity) as herein disclosed. In order to identify candidate hypervariable region sites for modification, alanine scanning mutagenesis can be performed to identify hypervariable region residues contributing significantly to antigen binding. Alternatively, or in addition, it may be beneficial to analyze a crystal structure of the antigen-antibody complex to identify contact points between the  
30 antibody and human TF. Such contact residues and neighboring residues are candidates for substitution according to the techniques elaborated herein. Once such variants are generated, the panel of variants is subjected to screening as described herein and antibodies with superior properties in one or more relevant assays may be selected for further development.

Another type of amino acid variant of the antibody alters the original glycosylation pattern of the antibody. By altering is meant deleting one or more carbohydrate moieties found in the antibody, and/or adding one or more glycosylation sites that are not present in the antibody.

5 Glycosylation of antibodies is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these  
10 tripeptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-aceylgalactosamine, galactose, or xylose to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

15 Addition of glycosylation sites to the antibody is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the sequence of the original antibody (for O-linked glycosylation sites).

20 Nucleic acid molecules encoding amino acid sequence variants of the anti-TF antibody are prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the anti-TF antibody.

25 (v) *Human antibodies*

Human antibodies can be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, *J. Mol. Biol.*, 227:381 (1991); Marks et al., *J. Mol. Biol.*, 222:581 (1991)]. The techniques of Cole et al. and Boerner et al. are also available for the preparation of human monoclonal antibodies (Cole et al.,  
30 Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985) and Boerner et al., *J. Immunol.*, 147(1):86-95 (1991)]. Similarly, human antibodies can be made by introducing of human immunoglobulin loci into transgenic animals, e.g., mice in which the endogenous immunoglobulin genes have been partially or completely inactivated. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all

respects, including gene rearrangement, assembly, and antibody repertoire. This approach is described, for example, in U.S. Patent Nos. 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,661,016, and in the following scientific publications: Marks et al., Bio/Technology 10, 779-783 (1992); Lonberg et al., Nature 368 856-859 (1994); Morrison, Nature 368, 812-13 (1994); Fishwild et al., Nature Biotechnology 14, 845-51 (1996); Neuberger, Nature Biotechnology 14, 826 (1996); Lonberg and Huszar, Intern. Rev. Immunol. 13 65-93 (1995).

(vi) *Antibody fragments*

In certain embodiments, the humanized or variant anti-TF antibody is an antibody fragment. Various techniques have been developed for the production of antibody fragments. Traditionally, these fragments were derived via proteolytic digestion of intact antibodies (see, e.g., Morimoto et al., J. Biochem. Biophys. Methods, 24:107-117 (1992) and Brennan et al., Science, 229:81 (1985)). However, these fragments can now be produced directly by recombinant host cells. For example, Fab'-SH fragments can be directly recovered from *E. coli* and chemically coupled to form F(ab')<sub>2</sub> fragments (Carter et al., Bio/Technology, 10:163-167 (1992)). In another embodiment, the F(ab')<sub>2</sub> is formed using the leucine zipper GCN4 to promote assembly of the F(ab')<sub>2</sub> molecule. According to another approach, Fv, Fab or F(ab')<sub>2</sub> fragments can be isolated directly from recombinant host cell culture. Other techniques for the production of antibody fragments will be apparent to the skilled practitioner.

(vii) *Multispecific antibodies*

In some embodiments, it may be desirable to generate multispecific (e.g. bispecific) humanized or variant anti-TF antibodies having binding specificities for at least two different epitopes. Exemplary bispecific antibodies may bind to two different epitopes of the TF protein. Alternatively, an anti-TF arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (e.g., CD2 or CD3), or Fc receptors for IgG (FcγR), such as FcγRI (CD64), FcγRII (CD32) and FcγRIII (CD16) so as to focus cellular defense mechanisms to the TF-expressing cell. Bispecific antibodies may also be used to localize cytotoxic agents to cells which express TF. These antibodies possess a TF-binding arm and an arm which binds the cytotoxic agent (e.g., saporin, anti-interferon-α, vinca alkaloid, ricin A chain, methotrexate or radioactive isotope hapten). Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e.g., F(ab')<sub>2</sub> bispecific antibodies).

According to another approach for making bispecific antibodies, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the C<sub>H</sub>3 domain of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e.g., tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e.g., alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers. See WO96/27011 published September 6, 1996.

Bispecific antibodies include cross-linked or "heteroconjugate" antibodies. For example, one of the antibodies in the heteroconjugate can be coupled to avidin, the other to biotin. Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in US Patent No. 4,676,980, along with a number of cross-linking techniques.

Techniques for generating bispecific antibodies from antibody fragments have also been described in the literature. For example, bispecific antibodies can be prepared using chemical linkage. Brennan *et al.*, *Science* 229:81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate F(ab')<sub>2</sub> fragments. These fragments are reduced in the presence of the dithiol complexing agent sodium arsenite to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes. In yet a further embodiment, Fab'-SH fragments directly recovered from *E. coli* can be chemically coupled *in vitro* to form bispecific antibodies, e.g. Shalaby et al., *J. Exp. Med.* 175:217-225 (1992).

Various techniques for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny et al., *J. Immunol.* 148(5):1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers were reduced

at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger et al., Proc. Natl. Acad. Sci. USA 90:6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (V<sub>H</sub>) connected to a light-chain variable domain (V<sub>L</sub>) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the V<sub>H</sub> and V<sub>L</sub> domains of one fragment are forced to pair with the complementary V<sub>L</sub> and V<sub>H</sub> domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See Gruber et al., J. Immunol. 152:5368 (1994). Alternatively, the bispecific antibody may be a "linear antibody" produced as described in Zapata et al. Protein Eng. 8(10):1057-1062 (1995).

Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared. Tutt et al., J. Immunol. 147:60 (1991).

(viii) *Other modifications*

Other modifications of the humanized or variant anti-TF antibody are contemplated. It may be desirable to modify the antibody of the invention with respect to effector function, so as to enhance the effectiveness of the antibody for instance in treating cancer. For example, cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron et al., J. Exp Med. 176:1191-1195 (1992) and Shopes, B. J. Immunol. 148:2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff et al., Cancer Research 53:2560-2565 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson et al., Anti-Cancer Drug Design 3:219-230 (1989).

The invention also pertains to immunoconjugates comprising the antibody described herein conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (e.g., an enzymatically active toxin of bacterial, fungal, plant or animal origin, or fragments thereof), or a radioactive isotope (i.e., a radioconjugate).

Chemotherapeutic agents useful in the generation of such immunoconjugates have been described above. Enzymatically active toxins and fragments thereof which can be used

include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain,  $\alpha$ -sarcin, *Aleurites fordii* proteins, dianthin proteins, *Phytolacca americana* proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin and the tricothecenes. A variety of radionuclides are available for the production of radioconjugated anti-TF antibodies. Examples include  $^{212}\text{Bi}$ ,  $^{131}\text{I}$ ,  $^{131}\text{In}$ ,  $^{90}\text{Y}$  and  $^{186}\text{Re}$ .

Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein coupling agents such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutaraldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta *et al.*, *Science* 238:1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026.

In another embodiment, the antibody may be conjugated to a "receptor" (such as streptavidin) for utilization in tumor pretargeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a "ligand" (e.g., avidin) which is conjugated to a cytotoxic agent (e.g., a radionuclide).

The anti-TF antibodies disclosed herein may also be formulated as immunoliposomes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Epstein *et al.*, *Proc. Natl. Acad. Sci. USA* 82:3688 (1985); Hwang *et al.*, *Proc. Natl. Acad. Sci. USA* 77:4030 (1980); and U.S. Pat. Nos. 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in U.S. Patent No. 5,013,556.

Particularly useful liposomes can be generated by the reverse phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol and PEG-derivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin

*et al.*, *J. Biol. Chem.* 257:286-288 (1982) via a disulfide interchange reaction. A chemotherapeutic agent (such as Doxorubicin) is optionally contained within the liposome. See Gabizon *et al.*, *J. National Cancer Inst.* 81(19):1484 (1989).

The antibody of the present invention may also be used in ADEPT by conjugating the antibody to a prodrug-activating enzyme which converts a prodrug (*e.g.*, a peptidyl chemotherapeutic agent, see WO81/01145) to an active anti-cancer drug. See, for example, WO 88/07378 and U.S. Patent No. 4,975,278.

The enzyme component of the immunoconjugate useful for ADEPT includes any enzyme capable of acting on a prodrug in such a way so as to convert it into its more active, cytotoxic form.

Enzymes that are useful in the method of this invention include, but are not limited to, alkaline phosphatase useful for converting phosphate-containing prodrugs into free drugs; arylsulfatase useful for converting sulfate-containing prodrugs into free drugs; cytosine deaminase useful for converting non-toxic 5-fluorocytosine into the anti-cancer drug, 5-fluorouracil; proteases, such as serratia protease, thermolysin, subtilisin, carboxypeptidases and cathepsins (such as cathepsins B and L), that are useful for converting peptide-containing prodrugs into free drugs; D-alanylcarboxypeptidases, useful for converting prodrugs that contain D-amino acid substituents; carbohydrate-cleaving enzymes such as  $\beta$ -galactosidase and neuraminidase useful for converting glycosylated prodrugs into free drugs;  $\beta$ -lactamase useful for converting drugs derivatized with  $\beta$ -lactams into free drugs; and penicillin amidases, such as penicillin V amidase or penicillin G amidase, useful for converting drugs derivatized at their amine nitrogens with phenoxyacetyl or phenylacetyl groups, respectively, into free drugs. Alternatively, antibodies with enzymatic activity, also known in the art as "abzymes", can be used to convert the prodrugs of the invention into free active drugs (see, *e.g.*, Massey, *Nature* 328:457-458 (1987)). Antibody-abzyme conjugates can be prepared as described herein for delivery of the abzyme to a tumor cell population.

The enzymes of this invention can be covalently bound to the anti-TF antibodies by techniques well known in the art such as the use of the heterobifunctional crosslinking reagents discussed above. Alternatively, fusion proteins comprising at least the antigen binding region of an antibody of the invention linked to at least a functionally active portion of an enzyme of the invention can be constructed using recombinant DNA techniques well known in the art (see, *e.g.*, Neuberger *et al.*, *Nature* 312:604-608 [1984]).

In certain embodiments of the invention, it may be desirable to use an antibody fragment, rather than an intact antibody, to increase tumor penetration, for example. In this

case, it may be desirable to modify the antibody fragment in order to increase its serum half life. This may be achieved, for example, by incorporation of a salvage receptor binding epitope into the antibody fragment (*e.g.*, by mutation of the appropriate region in the antibody fragment or by incorporating the epitope into a peptide tag that is then fused to the antibody fragment at either end or in the middle, *e.g.*, by DNA or peptide synthesis). See WO96/32478 published October 17, 1996.

The salvage receptor binding epitope generally constitutes a region wherein any one or more amino acid residues from one or two loops of a Fc domain are transferred to an analogous position of the antibody fragment. Even more preferably, three or more residues from one or two loops of the Fc domain are transferred. Still more preferred, the epitope is taken from the CH2 domain of the Fc region (*e.g.*, of an IgG) and transferred to the CH1, CH3, or V<sub>H</sub> region, or more than one such region, of the antibody. Alternatively, the epitope is taken from the CH2 domain of the Fc region and transferred to the C<sub>L</sub> region or V<sub>L</sub> region, or both, of the antibody fragment.

In one most preferred embodiment, the salvage receptor binding epitope comprises the sequence: PKNSSMISNTP (SEQ ID NO:14), and optionally further comprises a sequence selected from the group consisting of HQSLGTQ (SEQ ID NO:15), HQNLSDGK (SEQ ID NO:16), HQNISDGK (SEQ ID NO:17), or VISSHLGQ (SEQ ID NO:18), particularly where the antibody fragment is a Fab or F(ab')<sub>2</sub>. In another most preferred embodiment, the salvage receptor binding epitope is a polypeptide containing the sequence(s): HQNLSDGK (SEQ ID NO:16), HQNISDGK (SEQ ID NO:17), or VISSHLGQ (SEQ ID NO:18) and the sequence: PKNSSMISNTP (SEQ ID NO:14).

Covalent modifications of the humanized or variant anti-TF antibody are also included within the scope of this invention. They may be made by chemical synthesis or by enzymatic or chemical cleavage of the antibody, if applicable. Other types of covalent modifications of the antibody are introduced into the molecule by reacting targeted amino acid residues of the antibody with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues. Exemplary covalent modifications of polypeptides are described in US Patent 5,534,615, specifically incorporated herein by reference. A preferred type of covalent modification of the antibody comprises linking the antibody to one of a variety of nonproteinaceous polymers, *e.g.*, polyethylene glycol, polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.



### B. Vectors, Host Cells and Recombinant Methods

The invention also provides isolated nucleic acid encoding the humanized or variant anti-TF antibody, vectors and host cells comprising the nucleic acid, and recombinant techniques for the production of the antibody.

5 For recombinant production of the antibody, the nucleic acid encoding it may be isolated and inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. In another embodiment, the antibody may be produced by homologous recombination, *e.g.* as described in US Patent 5,204,244, specifically incorporated herein by reference. DNA encoding the monoclonal antibody is readily isolated and sequenced using  
10 conventional procedures (*e.g.*, by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of the antibody). Many vectors are available. The vector components generally include, but are not limited to, one or more of the following: a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence, *e.g.*, as described in  
15 US Patent 5,534,615 issued July 9, 1996 and specifically incorporated herein by reference.

Suitable host cells for cloning or expressing the DNA in the vectors herein are the prokaryote, yeast, or higher eukaryote cells described above. Suitable prokaryotes for this purpose include eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as *Escherichia*, *e.g.*, *E. coli*, *Enterobacter*, *Erwinia*, *Klebsiella*,  
20 *Proteus*, *Salmonella*, *e.g.*, *Salmonella typhimurium*, *Serratia*, *e.g.*, *Serratia marcescans*, and *Shigella*, as well as *Bacilli* such as *B. subtilis* and *B. licheniformis* (*e.g.*, *B. licheniformis* 41P disclosed in DD 266,710 published 12 April 1989), *Pseudomonas* such as *P. aeruginosa*, and *Streptomyces*. One preferred *E. coli* cloning host is *E. coli* 294 (ATCC 31,446), although other strains such as *E. coli* B, *E. coli* X1776 (ATCC 31,537), and *E. coli* W3110 (ATCC  
25 27,325) are suitable. These examples are illustrative rather than limiting.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for anti-TF antibody-encoding vectors. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are  
30 commonly available and useful herein, such as *Schizosaccharomyces pombe*; *Kluyveromyces* hosts such as, *e.g.*, *K. lactis*, *K. fragilis* (ATCC 12,424), *K. bulgaricus* (ATCC 16,045), *K. wickerhamii* (ATCC 24,178), *K. waltii* (ATCC 56,500), *K. drosophilum* (ATCC 36,906), *K. thermotolerans*, and *K. marxianus*; *Yarrowia* (EP 402,226); *Pichia pastoris* (EP 183,070); *Candida*; *Trichoderma reesia* (EP 244,234); *Neurospora crassa*; *Schwanniomyces* such as

*Schwanniomyces occidentalis*; and filamentous fungi such as, e.g., *Neurospora*, *Penicillium*, *Tolytocladium*, and *Aspergillus* hosts such as *A. nidulans* and *A. niger*.

Suitable host cells for the expression of glycosylated anti-TF antibody are derived from multicellular organisms. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda* (caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* have been identified. A variety of viral strains for transfection are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda* cells. Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can also be utilized as hosts.

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham *et al.*, *J. Gen Virol.* 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub *et al.*, *Proc. Natl. Acad. Sci. USA* 77:4216 [1980]); mouse sertoli cells (TM4, Mather, *Biol. Reprod.* 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HeLa, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather *et al.*, *Annals N.Y. Acad. Sci.* 383:44-68 [1982]); MRC 5 cells; and FS4 cells.

Host cells are transformed with the above-described expression or cloning vectors for anti-TF antibody production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

The host cells used to produce the anti-TF antibody of this invention may be cultured in a variety of media. Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ((MEM) (Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium (DMEM) (Sigma) are suitable for culturing the host cells. In addition, any of the media described in Ham *et al.*, *Meth. Enz.* 58:44 (1979), Barnes *et al.*, *Anal. Biochem.*

102:255 (1980), U.S. Pat. Nos. 4,767,704; 4,657,866; 4,927,762; 4,560,655; or 5,122,469;  
WO 90/03430; WO 87/00195; or U.S. Patent Re. 30,985 may be used as culture media for  
the host cells. Any of these media may be supplemented as necessary with hormones and/or  
other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as  
5 sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleotides  
(such as adenosine and thymidine), antibiotics (such as GENTAMYCIN<sup>TM</sup> drug), trace  
elements (defined as inorganic compounds usually present at final concentrations in the  
micromolar range), and glucose or an equivalent energy source. Any other necessary  
supplements may also be included at appropriate concentrations that would be known to  
10 those skilled in the art. The culture conditions, such as temperature, pH, and the like, are  
those previously used with the host cell selected for expression, and will be apparent to the  
ordinarily skilled artisan.

When using recombinant techniques, the antibody can be produced intracellularly, in  
the periplasmic space, or directly secreted into the medium. If the antibody is produced  
15 intracellularly, as a first step, the particulate debris, either host cells or lysed fragments, is  
removed, for example, by centrifugation or ultrafiltration. Carter *et al.*, *Bio/Technology*  
10:163-167 (1992) describe a procedure for isolating antibodies which are secreted to the  
periplasmic space of *E. coli*. Briefly, cell paste is thawed in the presence of sodium acetate  
(pH 3.5), EDTA, and phenylmethylsulfonylfluoride (PMSF) over about 30 min. Cell debris  
20 can be removed by centrifugation. Where the antibody is secreted into the medium,  
supernatants from such expression systems are generally first concentrated using a  
commercially available protein concentration filter, for example, an Amicon or Millipore  
Pellicon ultrafiltration unit. A protease inhibitor such as PMSF may be included in any of  
the foregoing steps to inhibit proteolysis and antibiotics may be included to prevent the  
25 growth of adventitious contaminants.

The antibody composition prepared from the cells can be purified using, for example,  
hydroxylapatite chromatography, gel electrophoresis, dialysis, and affinity chromatography,  
with affinity chromatography being the preferred purification technique. The suitability of  
protein A as an affinity ligand depends on the species and isotype of any immunoglobulin Fc  
30 domain that is present in the antibody. Protein A can be used to purify antibodies that are  
based on human  $\gamma 1$ ,  $\gamma 2$ , or  $\gamma 4$  heavy chains (Lindmark *et al.*, *J. Immunol. Meth.* 62:1-13  
(1983)). Protein G is recommended for all mouse isotypes and for human  $\gamma 3$  (Guss *et al.*,  
*EMBO J.* 5:15671575 (1986)). The matrix to which the affinity ligand is attached is most  
often agarose, but other matrices are available. Mechanically stable matrices such as

controlled pore glass or poly(styrene-divinyl)benzene allow for faster flow rates and shorter processing times than can be achieved with agarose. Where the antibody comprises a C<sub>H</sub>3 domain, the Bakerbond ABX<sup>TM</sup> resin (J. T. Baker, Phillipsburg, NJ) is useful for purification. Other techniques for protein purification such as fractionation on an ion-exchange column, ethanol precipitation, Reverse Phase HPLC, chromatography on silica, chromatography on heparin SEPHAROSE<sup>TM</sup> chromatography on an anion or cation exchange resin (such as a polyaspartic acid column), chromatofocusing, SDS-PAGE, and ammonium sulfate precipitation are also available depending on the antibody to be recovered.

Following any preliminary purification step(s), the mixture comprising the antibody of interest and contaminants may be subjected to low pH hydrophobic interaction chromatography using an elution buffer at a pH between about 2.5-4.5, preferably performed at low salt concentrations (e.g., from about 0-0.25M salt).

### C. Pharmaceutical Formulations

Therapeutic formulations of the antibody are prepared for storage by mixing the antibody having the desired degree of purity with optional physiologically acceptable carriers, excipients or stabilizers (*Remington's Pharmaceutical Sciences* 16th edition, Osol, A. Ed. (1980)), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g., Zn-protein complexes); and/or non-ionic surfactants such as TWEEN<sup>TM</sup>, PLURONICS<sup>TM</sup> or polyethylene glycol (PEG).

The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other (see Section G below). For example, the

composition can include at least one additional anticoagulant or anti-platelet agent. Preferably, the anticoagulant is an agent that prevents the growth or formation of a thrombus such as thrombin inhibitors including heparin, low molecular weight heparin, pentasaccharide thrombin inhibitors and direct thrombin inhibitors. The antiplatelet agents  
5 are preferably cyclooxygenase inhibitors, ADP receptor antagonists, phosphodiesterase inhibitors, and GP IIb/IIIa inhibitors. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsule prepared, for example, by coacervation techniques or by interfacial polymerization, for example,  
10 hydroxymethylcellulose or gelatin-microcapsule and poly-(methylmethacrylate) microcapsule, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in *Remington's Pharmaceutical Sciences* 16th edition, Osol, A. Ed. (1980).

15 The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, *e.g.*, films, or  
20 microcapsule. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and  $\gamma$  ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the Lupron Depot<sup>TM</sup> (injectable microspheres composed of lactic acid-glycolic acid copolymer and  
25 leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in  
30 immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling

moisture content, using appropriate additives, and developing specific polymer matrix compositions.

D. Non-therapeutic Uses for the Antibody

5 The antibodies of the invention may be used as affinity purification agents. In this process, the antibodies are immobilized on a solid phase such as Sephadex resin or filter paper, using methods well known in the art. The immobilized antibody is contacted with a sample containing the TF protein (or fragment thereof) to be purified, and thereafter the support is washed with a suitable solvent that will remove substantially all the material in the sample except the TF protein, which is bound to the immobilized antibody. Finally, the support is washed with another suitable solvent, such as glycine buffer, pH 5.0, that will release the TF protein from the antibody.

10 Anti-TF antibodies may also be useful in diagnostic assays for TF protein, *e.g.*, detecting its expression in specific cells, tissues, or serum. Such diagnostic methods may be useful in the diagnosis of various disorders associated with the aberrant expression, *e.g.* over- or underexpression of TF. For example, overexpression and/or aberrant utilization of TF has been linked to the pathophysiology of both thrombosis and sepsis, and TF has been implicated in tumor metastasis. Accordingly, anti-TF antibodies may be useful in the diagnosis of these diseases.

20 For diagnostic applications, the antibody typically will be labeled with a detectable moiety. Numerous labels are available which can be generally grouped into the following categories:

(a) Radioisotopes, such as  $^{35}\text{S}$ ,  $^{14}\text{C}$ ,  $^{125}\text{I}$ ,  $^3\text{H}$ , and  $^{131}\text{I}$ . The antibody can be labeled with the radioisotope using the techniques described in *Current Protocols in Immunology*, Volumes 1 and 2, Coligen *et al.*, Ed. Wiley-Interscience, New York, New York, Pubs. 25 (1991) for example and radioactivity can be measured using scintillation counting.

(b) Fluorescent labels such as rare earth chelates (europium chelates) or fluorescein and its derivatives, rhodamine and its derivatives, dansyl, Lissamine, phycoerythrin and Texas Red are available. The fluorescent labels can be conjugated to the antibody using the techniques disclosed in *Current Protocols in Immunology*, *supra*, for 30 example. Fluorescence can be quantified using a fluorimeter.

(c) Various enzyme-substrate labels are available and U.S. Patent No. 4,275,149 provides a review of some of these. The enzyme generally catalyzes a chemical alteration of the chromogenic substrate which can be measured using various techniques. For example, the enzyme may catalyze a color change in a substrate, which can be measured

spectrophotometrically. Alternatively, the enzyme may alter the fluorescence or chemiluminescence of the substrate. Techniques for quantifying a change in fluorescence are described above. The chemiluminescent substrate becomes electronically excited by a chemical reaction and may then emit light which can be measured (using a chemiluminometer, for example) or donates energy to a fluorescent acceptor. Examples of enzymatic labels include luciferases (*e.g.*, firefly luciferase and bacterial luciferase; U.S. Patent No. 4,737,456), luciferin, 2,3-dihydrophthalazinediones, malate dehydrogenase, urease, peroxidase such as horseradish peroxidase (HRPO), alkaline phosphatase,  $\beta$ -galactosidase, glucoamylase, lysozyme, saccharide oxidases (*e.g.*, glucose oxidase, galactose oxidase, and glucose-6-phosphate dehydrogenase), heterocyclic oxidases (such as uricase and xanthine oxidase), lactoperoxidase, microperoxidase, and the like. Techniques for conjugating enzymes to antibodies are described in O'Sullivan *et al.*, Methods for the Preparation of Enzyme-Antibody Conjugates for use in Enzyme Immunoassay, in *Methods in Enzym.* (ed J. Langone & H. Van Vunakis), Academic press, New York, 73:147-166 (1981).

Examples of enzyme-substrate combinations include, for example:

- (i) Horseradish peroxidase (HRPO) with hydrogen peroxidase as a substrate, wherein the hydrogen peroxidase oxidizes a dye precursor (*e.g.*, orthophenylene diamine (OPD) or 3,3',5,5'-tetramethyl benzidine hydrochloride (TMB));
- (ii) alkaline phosphatase (AP) with para-Nitrophenyl phosphate as chromogenic substrate; and
- (iii)  $\beta$ -D-galactosidase ( $\beta$ -D-Gal) with a chromogenic substrate (*e.g.*, p-nitrophenyl- $\beta$ -D-galactosidase) or fluorogenic substrate 4-methylumbelliferyl- $\beta$ -D-galactosidase.

Numerous other enzyme-substrate combinations are available to those skilled in the art. For a general review of these, see U.S. Patent Nos. 4,275,149 and 4,318,980.

Sometimes, the label is indirectly conjugated with the antibody. The skilled artisan will be aware of various techniques for achieving this. For example, the antibody can be conjugated with biotin and any of the three broad categories of labels mentioned above can be conjugated with avidin, or *vice versa*. Biotin binds selectively to avidin and thus, the label can be conjugated with the antibody in this indirect manner. Alternatively, to achieve indirect conjugation of the label with the antibody, the antibody is conjugated with a small hapten (*e.g.*, digoxin) and one of the different types of labels mentioned above is conjugated with an

anti-hapten antibody (*e.g.*, anti-digoxin antibody). Thus, indirect conjugation of the label with the antibody can be achieved.

In another embodiment of the invention, the anti-TF antibody need not be labeled, and the presence thereof can be detected using a labeled antibody which binds to the TF antibody.

The antibodies of the present invention may be employed in any known assay method, such as competitive binding assays, direct and indirect sandwich assays, and immunoprecipitation assays. Zola, *Monoclonal Antibodies: A Manual of Techniques*, pp.147-158 (CRC Press, Inc. 1987).

Competitive binding assays rely on the ability of a labeled standard to compete with the test sample analyte for binding with a limited amount of antibody. The amount of TF protein in the test sample is inversely proportional to the amount of standard that becomes bound to the antibodies. To facilitate determining the amount of standard that becomes bound, the antibodies generally are insolubilized before or after the competition, so that the standard and analyte that are bound to the antibodies may conveniently be separated from the standard and analyte which remain unbound.

Sandwich assays involve the use of two antibodies, each capable of binding to a different immunogenic portion, or epitope, of the protein to be detected. In a sandwich assay, the test sample analyte is bound by a first antibody which is immobilized on a solid support, and thereafter a second antibody binds to the analyte, thus forming an insoluble three-part complex. See, *e.g.*, US Pat No. 4,376,110. The second antibody may itself be labeled with a detectable moiety (direct sandwich assays) or may be measured using an anti-immunoglobulin antibody that is labeled with a detectable moiety (indirect sandwich assay). For example, one type of sandwich assay is an ELISA assay, in which case the detectable moiety is an enzyme.

For immunohistochemistry, the tumor sample may be fresh or frozen or may be embedded in paraffin and fixed with a preservative such as formalin, for example.

The antibodies may also be used for *in vivo* diagnostic assays. Generally, the antibody is labeled with a radio nuclide (such as <sup>111</sup>In, <sup>99</sup>Tc, <sup>14</sup>C, <sup>131</sup>I, <sup>125</sup>I, <sup>3</sup>H, <sup>32</sup>P or <sup>35</sup>S) so that the tumor can be localized using immunoscintigraphy.



### E. Diagnostic Kits

As a matter of convenience, the antibody of the present invention can be provided in a kit, *i.e.*, a packaged combination of reagents in predetermined amounts with instructions for performing the diagnostic assay. Where the antibody is labeled with an enzyme, the kit will include substrates and cofactors required by the enzyme (*e.g.*, a substrate precursor which provides the detectable chromophore or fluorophore). In addition, other additives may be included such as stabilizers, buffers (*e.g.*, a block buffer or lysis buffer) and the like. The relative amounts of the various reagents may be varied widely to provide for concentrations in solution of the reagents which substantially optimize the sensitivity of the assay. Particularly, the reagents may be provided as dry powders, usually lyophilized, including excipients which on dissolution will provide a reagent solution having the appropriate concentration.

### F. Therapeutic Uses for the Antibody

For therapeutic applications, the anti-TF antibodies of the invention are administered to a mammal, preferably a human, in a pharmaceutically acceptable dosage form such as those discussed above, including those that may be administered to a human intravenously as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intra-cerebrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. The antibodies also are suitably administered by intra tumoral, peritumoral, intralesional, or perilesional routes, to exert local as well as systemic therapeutic effects. The intraperitoneal route is expected to be particularly useful, for example, in the treatment of ovarian tumors.

For the prevention or treatment of disease, the appropriate dosage of antibody will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the antibody is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the antibody, and the discretion of the attending physician. The antibody is suitably administered to the patient at one time or over a series of treatments.

The anti-TF antibodies are useful in the treatment of various neoplastic and non-neoplastic diseases and disorders, such as TF-FVIIa related diseases or disorders. Such diseases or disorders include, for example, chronic thromboembolic diseases or disorders associated with fibrin formation including vascular disorders such as deep venous thrombosis, arterial thrombosis, acute coronary syndrome including unstable angina and non-ST-segment myocardial infarction, stroke, tumor metastasis, thrombolysis, arteriosclerosis

and restenosis following angioplasty, acute and chronic indications such as inflammation, septic shock, septicemia, hypotension, adult respiratory distress syndrome (ARDS), disseminated intravascular coagulopathy (DIC). The TF-FVIIa related disorder is not limited to in vivo coagulopathic disorders such as those named above but includes ex vivo TF-FVIIa related processes such as coagulation that may result from the extracorporeal circulation of blood, including blood removed in-line from a patient in such processes as dialysis procedures, blood filtration, or blood bypass during surgery.

Depending on the type and severity of the disease, about 1 µg/kg to about 50 mg/kg (e.g., 0.1-20mg/kg) of antibody is an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A typical daily or weekly dosage might range from about 1 µg/kg to about 20 mg/kg or more, depending on the factors mentioned above. For repeated administrations over several days or longer, depending on the condition, the treatment is repeated until a desired suppression of disease symptoms occurs. However, other dosage regimens may be useful. The progress of this therapy is easily monitored by conventional techniques and assays, including, for example, radiographic tumor imaging.

G. Combinations of Anti-TF Antibodies and Anticoagulant and/or Anti-Platelet Agents

According to another embodiment of the invention, anti-TF antibodies administered, serially or in combination, with at least one additional anticoagulant and/or anti-platelet agent enhance the antithrombotic efficacy of the anticoagulant or antiplatelet agent. Preferably, the additional anticoagulant agent is selected from the group consisting of heparin, low molecular weight heparin, pentasaccharide thrombin inhibitors, and mixtures thereof.

Many therapies for thrombotic conditions, such as acute coronary syndrome, involve the administration of multiple agents. These agents act to inhibit platelet aggregation or the activity of thrombin. For example, a current therapy for acute coronary syndrome is administration of aspirin, heparin and a GP IIb/IIIa inhibitor. While this therapy has improved efficacy, the administration of heparin and GP IIb/IIIa inhibitors is associated with bleeding and thrombocytopenia. Heparin can have decreased effectiveness due to its indirect action on thrombin, binding to various plasma proteins, variable clearance rates and heparin resistance. Increasing the dose of heparin to enhance efficacy of a treatment is limited because of the increased bleeding risk and risk of overdosing. Minimizing the dose of

heparin in a treatment is a desirable outcome but can be difficult to achieve while maintaining therapeutic efficacy.

Anti-TF antibodies act a point much earlier in the coagulation pathway than the therapeutic agents described above and inhibit generation of thrombin by the extrinsic pathway of coagulation. Many inherited diseases that result in impaired function of clotting factors such as factor VIII, IX and X have impaired blood clotting and spontaneous bleeding. Thus, it might have been expected that any approach using agents that interfere with the activation of these clotting factors would not improve antithrombotic efficacy and/or would have an unacceptable risk of bleeding. The findings, as described and shown herein, show that the combination an anti-TF antibody with at least one additional anticoagulant and/or anti-platelet agent enhances antithrombotic efficacy and minimally, if at all, increases bleeding risk compared to administration of the antibody or agent alone.

Treatments comprising an anti-TF antibody and at least one additional anticoagulant and/or anti-platelet agent are useful in the treatment of TF-VIIa related diseases or disorders. Such diseases or disorders include, for example, chronic thromboembolic diseases or disorders associated with fibrin formation such as deep venous thrombosis, arterial thrombosis, acute coronary syndrome including unstable angina and non-ST-segment myocardial infarction, stroke, thrombolysis, tumor metastasis, arteriosclerosis and restenosis following angioplasty, acute and chronic indication such as inflammation, septic shock, septicemia, hypotension, adult respiratory distress syndrome (ARDS), and disseminated intravascular coagulopathy (DIC). Preferably, the treatment is useful in the treatment of acute coronary syndrome.

The anti-TF antibodies include those antibodies that neutralize the activity of tissue factor and thereby prevent formation of thrombin. Neutralizing anti-TF antibodies include those, for example, that inhibit the binding of Factor VII/VIIa and/or those that interfere with the interaction of macromolecular substrates such as Factor X or IX to tissue factor. Anti-TF antibodies which bind to an epitope overlapping with the C-terminal macromolecular substrate-binding region of TF, and thus interfere with the TF-substrate interaction, are the most potent anticoagulant agents. Preferably, the anti-tissue factor antibody is a humanized antibody that binds to an epitope comprising at least part of the C-terminal macromolecular substrate binding region of tissue factor, such as D3H44.

The additional anticoagulant agents useful in combination therapies are agents that prevent, inhibit or prolong blood coagulation and that are not anti-TF antibodies. The anticoagulant agents are, preferably, those agents that inhibit the formation or growth of a

thrombus. The anticoagulant agents include thrombin inhibitors such as heparin, low molecular weight heparin, pentasaccharide thrombin inhibitors, and direct thrombin inhibitors as well as factor X/Xa inhibitors and antibodies to coagulation factors. Low molecular weight heparin is heparin that has an average molecular weight less than about 5 6500 daltons and can be prepared by modification of natural sources of heparin or prepared synthetically. Low molecular weight heparins include enoxaparin, dalteparin, nadroparin (Fraxiparin), tinzaparin and are commercially available. Pentasaccharide thrombin inhibitors include an antithrombin III-binding fragment of heparin and other agents that inhibit Factor Xa. Pentasaccharide inhibitors include fondaparinux sodium, and SR 34006 . 10 Direct thrombin inhibitors act to inhibit the action of thrombin and include, for example, hirudin, bivalirudin (Hirulog), argatroban, and Lepirudin ([Leu 1, Thr 2]-63-disulfohirudin). The preferred anticoagulant agents are heparin, low molecular weight heparin, or pentasaccharide thrombin inhibitors.

Anti-platelet agents are agents that inhibit platelet activation or platelet aggregation. 15 Anti-platelet agents include cyclooxygenase inhibitors, ADP receptor antagonists, phosphodiesterase inhibitors and GP IIb/IIIa inhibitors. Cyclooxygenase inhibitors include acetylsalicylic acid, ibuprofen, indomethacin and sulfinpyrazone. ADP receptor antagonists include clopidogrel and ticlopidine. Phosphodiesterase inhibitors include cilostazol. The glycoprotein GP IIb/IIIa inhibitors include abciximab, G4120, eptifibatide and tirofiban. The 20 preferred anti-platelet agents are GP IIb/IIIa inhibitors.

The anti-TF antibodies and anticoagulant and/or anti-platelet agents can be administered at the same time or at different times. Typically, an anti-TF antibody is given as a single dose and the additional anticoagulant or antiplatelet agent is administered as a single dose followed by an infusion. The antibodies and agents are administered, preferably, at 25 suboptimal doses and achieve greater anti-thrombotic efficacy compared to administration of the same dose of agent or antibody alone. A suboptimal dose is a dose that results in less than about 80% reduction or inhibition of the incidence and/or severity of a thrombus compared to saline control.

The administration of the antibody and agents are effective to enhance antithrombotic efficacy and preferably, minimize an increase in bleeding risk compared to administration of 30 the antibody or agent alone. Preferably, the anti-TF antibody in combination with at least one additional agent achieves the same reduction or inhibition of a thrombus at a lower dose and /or in a shorter time compared to the antibody or agent alone. More preferably, the combination of the anti-TF antibody with an anticoagulant or antiplatelet agent provides for at

least about 30% or greater inhibition or reduction in incidence and/or severity of a thrombus compared to the same dose of antibody or agent alone. For example, in clinical trial settings, treatments that have at least about 30% or greater improvement in outcomes compared to control are viewed as having enhanced efficacy. As described herein, co-administration of suboptimal doses of an anti-TF antibody with unfractionated heparin results in about 70-75% more decrease in thrombus mass compared to administration of the same dose of unfractionated heparin.

The administration of a treatment comprising a dose of an anti-TF antibody and a dose of at least one additional anticoagulant or anti-platelet agent preferably provides a “minimal increase in bleeding risk”. A “minimal increase in bleeding risk” refers to little or no significant increase in the incidence and/or severity of bleeding events compared to control. Methods for assessing bleeding risk are known to those of skill in the art and include standard bleeding time assays, such as cuticle bleeding time, surgical blood loss, or template bleeding in animal models and the TIMI criteria as described in Menon et al, *supra*. Whether bleeding risk is significantly increased can readily be determined by one of skill in the art using accepted statistical analysis. For example, when bleeding risk is assessed for the combination treatments as described herein, bleeding times are prolonged about 25% or less compared to the administration of the antibody or agent alone. When bleeding risk is assessed clinically, a minimal increase in bleeding risk would be an increase of minor bleeding events of about 20% or less compared to control and/or an increase in major bleeding events of about 10% or less compared to control. In some cases, the bleeding risk may be decreased compared to control. Preferably, the dose of the antibody and/or agent is a dose that does not significantly prolong the bleeding time or significantly increase the bleeding risk compared to control.

Also, the antibody is suitably administered serially or in combination with radiological treatments, whether involving irradiation or administration of radioactive substances.

#### H. Articles of Manufacture

In another embodiment of the invention, an article of manufacture containing materials useful for the treatment of the disorders described above is provided. The article of manufacture comprises a container and a label. Suitable containers include, for example, bottles, vials, syringes, and test tubes. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition which is effective for treating the condition and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection

needle). The active agent in the composition is the anti-TF antibody. The label on, or associated with, the container indicates that the composition is used for treating the condition of choice. The article of manufacture may further comprise a second container comprising a pharmaceutically-acceptable buffer, such as phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, syringes, and package inserts with instructions for use.

The following examples are offered by way of illustration and not by way of limitation. The disclosures of all citations in the specification are expressly incorporated herein by reference.

## EXAMPLES

### EXAMPLE 1

This example describes the determination of the binding epitopes of 5 neutralizing anti-TF antibodies and establishes the respective roles of binding affinity and epitope location on the anticoagulant potencies in different systems. Interestingly, the results demonstrate that the anticoagulant potencies have no correlation with antibody binding affinities. Rather, potency is primarily determined by the precise location of the antibody-binding site on the TF molecules.

## MATERIALS AND METHODS

### Materials.

Fatty acid-free BSA was from Calbiochem (La Jolla, CA). Human recombinant FVIIa was a gift from Mark O'Connell (Genentech, Inc.). FX was from Haematologic Technologies Inc. (Essex Junction, VT). Thrombin inhibitor napsagatran was a gift from Dr. Kurt Hilpert (Roche, Switzerland). Chromozym t-PA was from Boehringer Mannheim (Indianapolis, IN). Truncated transmembrane tissue factor comprising residues 1-243 (TF<sub>1-243</sub>) was generated and relipidated as described (47,48). FX chromogenic substrate S2765 was from Diapharma Group Inc. (Columbus, OH).

### Preparation of murine D3 Fab fragments.

Fab fragments were prepared from the D3 antibody by digestion with papain in the presence of cysteine. A concentrated solution of the D3 Mab was prepared for digestion by dialysis versus 0.1 M sodium acetate pH 5.5, 1 mM EDTA. To this solution (11.6 mg/mL antibody) was added solid cysteine to a final concentration of 50 mM. Sufficient papain (Worthington Biochemical Corp., Lakewood, NJ) was added to give a 1:100 weight ratio to antibody and the solution was incubated at 37 °C. After 8 hours the digestion was quenched

by addition of 100 mM iodoacetamide to inactivate the papain. Residual intact antibody and Fc fragments were removed by passing the solution over a Protein A-Sepharose column. The Fab fragments in the flow-through fraction were further purified by affinity chromatography on a column of immobilized soluble TF<sub>1-219</sub> (sTF). The affinity column was prepared by using a 1 x 5 mL NHS-activated HiTrap column (Pharmacia Biotech, Piscataway, NJ) following the instructions supplied by the manufacturer. The final coupling density achieved was 25 mg of sTF per mL of resin. D3 Fab were eluted from this column by washing with a solution of 0.1 M acetate pH 3, 0.2 M NaCl and the Fab containing fractions were pooled and neutralized with 2 M Tris base.

#### Clotting assays.

For pre-incubation assays, 20 µl of antibody was added to 180 µl relipidated human tissue factor (Innovin, Dade Behring Inc., Newark, DE) and incubated at 37°C for 15 min. 100 µl of normal citrated human plasma (Peninsula Blood Bank, Burlingame, CA) was added and clotting times were measured using an MLA Electra 800 (Medical Laboratory Automation Inc.; Pleasant, NY).

#### Prothrombin time assays.

Prothrombin time (PT) assays, antibody was added to citrated human plasma. After 5min incubation, clotting was started by adding human tissue factor reagent Innovin. Clotting times were measured on an ACL300 using the PT mode (Coulter Corp., Miami, FL). For both assays, the antibody concentrations are reported as final concentrations in the reaction mixture (including the tissue factor reagent).

#### Site-directed mutagenesis, expression, and purification of sTF mutants.

Expression of sTF mutants (TF<sub>1-219</sub>) in E.coli and subsequent purification on a D3 antibody affinity column was carried out as described earlier (Kelley et al., [1995] Biochemistry 34: 10383-10392). For sTF mutants which did not bind to the D3 column (N199A:R200A and K201A:D204A), a 7G11 antibody column was used. This column was prepared by coupling the 7G11 antibody to CNBr-activated Sepharose 4B (Pharmacia, Piscataway, NJ) according to the manufacturer's instructions. Cell pellets were stored at -20°C for at least 1 hr. The osmotic shock supernatants were applied to the antibody affinity column which was equilibrated with 50 mM Tris-HCl, pH 8.0, 500 mM NaCl (buffer A). To remove nonspecifically bound proteins, the column was washed with 6 column volumes of buffer A and 50 mM Tris-HCl pH 8.0, 1.0 M NaCl, 0.5M tetramethylammonium chloride. sTF mutants were eluted with 0.1 M sodium acetate, pH 3.0, 0.2 M NaCl. Fractions were

neutralized and peak fractions concentrated using a Centriprep 10 (Amicon, Beverly, MA). Protein concentrations were determined by absorbance measurements using an  $\lambda_{280}$  of 29.4  $\text{mM}^{-1} \text{cm}^{-1}$  calculated from quantitative amino acid analysis data. An  $\lambda_{280}$  of 24  $\text{mM}^{-1} \text{cm}^{-1}$  was used for the Trp to Phe mutants of sTF.

5                    Determination of anti-TF antibody's TF binding affinity and antibody epitope mapping.

10                    The binding affinity of sTF for immobilized antibody was determined by surface plasmon resonance (SPR) measurements on a Pharmacia BIAcore 2000 instrument (Pharmacia Biosensor). Each antibody was coupled to the sensor chip surface at a level of 2000-3000 resonance units using amine coupling chemistry (Pharmacia Biosensor). In a typical experiment, 4 different antibodies were immobilized on each of the 4 flow cells of the sensor chip so that sensorgrams could be recorded simultaneously for all 4 antibodies. Sensorgrams were recorded for sTF binding at concentrations ranging from 15.6 nM to 500 nM in 2-fold increments. The kinetic constants were determined by non-linear regression analysis according to a 1:1 binding model using software supplied by the manufacturer. Dissociation constants were calculated from the kinetic constants. In experiments to determine competition between the antibody and FVIIa for binding to sTF, the same sTF concentration series was prepared in the presence of 5  $\mu\text{M}$  human, recombinant FVIIa. These solutions were incubated at ambient temperature for 30 minutes prior to injection onto the sensor chip. The epitopes on sTF for binding the monoclonal antibodies were determined by measuring the effect of amino acid substitutions in sTF on the affinity for immobilized antibody. Affinities were determined by SPR measurements as described above for the wild-type protein.

25                    Monoclonal antibodies.

30                    Monoclonal antibody 7G11 was generated by immunizing female BALB/c mice subcutaneously 3 times, intraperitoneally 3 times with 20 $\mu\text{g}$  sTF in MPL/TDM adjuvant (Ribi Immunochem Research, Hamilton, MT), at 2 week intervals. These mice were further boosted 8 times into footpads with 10 $\mu\text{g}$  sTF in 100 $\mu\text{l}$  MPL/TDM Adjuvant every week. 5G6 was generated by immunizing female BALB/c mice via footpad with 10 $\mu\text{g}$  sTF in 100 $\mu\text{l}$  MPL/TDM adjuvant, 13 times every week. Four days after the last boost, popliteal lymph nodes were removed and fused with mouse myeloma cells P3X63Ag8U.1 (Yelton et al., [1978] Curr. Top. Microbiol. Immunol. 81: 1-7) using 35% polyethyleneglycol as described (Chuntharapai and Kim, [1997] Methods Enzymol. 288: 15-27). Hybridoma cell lines



secreting antibody specific for sTF, as determined by ELISA, were cloned twice by limiting dilution and further characterized. Ascites were produced in BALB/c mice and monoclonal antibodies were purified using protein G conjugated Sepharose 4B. The generation of D3 antibody was described previously (Paborsky et al., [1990] Prot. Engineering 3: 547-553) and the antibody 6B4 came from a separate immunization protocol. The HTF1 antibody was described by Carson et al. (Carson et al., [1987] supra).

FX activation in human plasma.

The antibodies were diluted in human citrated plasma from a donor plasma pool (Peninsula Blood Bank, Burlingame, CA) for 10min at room temperature. At the end of the incubation period the thrombin inhibitor napsagatran (Hilpert et al., [1994] J. Med. Chem. 37: 3889-3901; Gast and Tschopp [1995] Blood Coag. Fibrinolysis 6: 533-560) was added. FX activation was started with relipidated TF<sub>1-243</sub> in 20mM hepes, pH 7.5, 0.5% BSA (HBS buffer) containing 15mM CaCl<sub>2</sub>. The reaction mixture contained 33% plasma and the concentrations of relipidated TF<sub>1-243</sub>, napsagatran and CaCl<sub>2</sub> were 0.4nM, 0.5μM and 5mM, respectively. 50μl aliquots taken at 15sec intervals were quenched in 150μl of 20mM EDTA. In the second stage, 50μl of 1.5mM FXa chromogenic substrate S2765 was added and increase in absorbance at 405nm monitored on a kinetic microplate reader (Molecular Devices, Menlo Park, CA). The initial rates were calculated by the linear fit of the values over a 45 sec. period. The values of aliquots taken at later time points indicated that the linear phase of the reaction was limited to this short time period. Control experiments in which relipidated TF<sub>1-243</sub> was omitted showed that there was no increase in chromogenic activity, indicating the absence of any FXa generation without TF. Also, napsagatran had no effect on FXa amidolytic activity towards S2765 under the employed conditions, which is consistent with the reported high selectivity towards thrombin (Hilpert et al., [1994] supra). From standard curves with FXa incubated with plasma and all other components used in the assay, it was calculated that under non-inhibited conditions an average of 8.6 nM ± 0.9 FXa/min (±S.D.) was generated.

The amidolytic activities of several coagulation factors, such as factors IIa, VIIa, IXa and XIa were tested under identical assay conditions to test whether other coagulation factors generated during the reaction might have contributed to the amidolytic activity measured in the second stage of the assay. Only factor XIa displayed significant amidolytic activity towards S2765, which was about 25% of the FXa activity. To assess a possible contribution of factor XIa in our assay system, the inhibitory activity of the D3 antibody was examined in

factor XI-deficient plasma (George King Bio-Medical, Overland Park, KS). The IC<sub>50</sub> value of 5.2±0.1 µg/ml (±SD, n=4) was similar to the value determined in normal human plasma. In addition, experiments carried out in factor II-, and factor VIII-deficient plasmas (American Diagnostica) gave similar results (6.4±0.9 µg/ml and 7.6±1.5 µg/ml respectively). Together, these results strongly suggested that rates of S2765 cleavage accurately reflected the concentration of FXa generated by relipidated TF<sub>1-243</sub>:FVIIa complex in plasma.

Amidolytic activity of soluble TF:FVIIa complex.

The antibodies were incubated with sTF and FVIIa in HBS buffer containing 5mM CaCl<sub>2</sub> for 20min prior to addition of Chromozym t-PA. The final concentration of the reactants was as follows: 10nM sTF, 10nM FVIIa, 0.5mM Chromozym t-PA. The rates of amidolytic activity were measured at 405nm on a kinetic microplate reader (Molecular Devices). The background activity was defined as the amidolytic activity of FVIIa in the absence of sTF and was subtracted from the obtained values.

Human ex-vivo thrombosis model.

Tissue factor-expressing human J82 cells (epithelial carcinoma, ATCC HTB1) were grown on Thermanox plastic coverslips as described (Kirchhofer et al., [1995] Arterioscler. Thromb. Vasc. Biol. 15: 1098-1106). The coverslips with the cell monolayer were then positioned in parallel plate perfusion chambers and the entire system including tubings, mixing devices and parallel plate chambers was filled with DMEM-1% (w/v) BSA. The details of the experimental system were described recently (Kirchhofer et al., [1995] supra; Kirchhofer et al., [1994] J. Clin. Invest. 93: 2073-2083). Blood was then drawn from the antecubital vein of a healthy donor at a rate of 1 mL/min. Immediately before entering the mixing chambers the flowing blood was infused with the antibodies at a rate of 50 µL/min by use of an infusion pump (Infu 362, Datex AG, Switzerland). The homogenous blood-antibody mixture then entered three parallel plate perfusion devices containing the J82 cell monolayers. The blood flow of 1 mL/min resulted in a shear rate of 65 s<sup>-1</sup> on the coverslips which corresponded to venous blood flow conditions. After a 3.5 minute perfusion period the cell layer was washed and coverslips removed for visual inspection of deposited fibrin. Fibrinopeptide A (FPA) levels were measured in the blood leaving the perfusion device as described previously (Kirchhofer et al., [1994] and [1995] supra).

RESULTS

Functionally different anti-TF antibodies.

As seen in Fig.1a, the antibodies 7G11, 6B4 and HTF1 completely inhibited sTF:FVIIa-dependent activity towards Chromozym t-PA, indicating interference with the proper formation of the sTF:FVIIa complex. In contrast, 5G6 antibody did not inhibit at all, whereas D3 reduced activity by about 20%, reaching a plateau at higher concentration. The inhibition by D3 was also seen when the smaller antibody Fab was used (Fig.1a). The inhibitory effect of D3 was dependent on low sTF concentrations, since no inhibition occurred at high sTF concentrations (120-200nM) in the presence of molar excess antibody (data not shown). In agreement, D3 did not affect the amidolytic activity when relipidated TF<sub>1-243</sub> was used, which binds FVIIa with much higher affinity than sTF (data not shown). However, both D3 and 5G6 inhibited TF:FVIIa-mediated activation of macromolecular substrate as well as the other antibodies. This was shown by results obtained from clotting assays in which antibodies were pre-incubated with TF reagent. (Fig.1b).

Anticoagulant potencies of anti-TF antibodies.

The results of the amidolytic assays indicated two fundamentally different types of anti-TF antibodies. Two antibodies of each group (D3 and 5G6 vs 6B4 and HTF1) were selected and their anticoagulant potencies in a human ex-vivo blood flow system determined (Kirchhofer et al., [1994] and [1995] supra). In this system the antibodies were infused to flowing non-anticoagulated human blood, which then entered parallel plate devices containing a monolayer of TF-expressing J82 cells. The shear rate at the cell layer was  $65 \text{ s}^{-1}$  simulating venous blood flow conditions. In controls this resulted in the generation of fibrinopeptide A (FPA) and the deposition of polymerized fibrin onto the cell monolayer. The average FPA levels of 36 control samples was  $1348 \pm 46.1 \text{ ng/ml plasma } (\pm \text{SEM})$ , which was similar to earlier reported FPA concentrations using the same system ( $1192 \pm 69 \text{ ng/ml}$ ; (Kirchhofer et al., [1995] supra)). Infusion of D3 and 5G6 potently inhibited FPA generation with  $\text{IC}_{50}$  values of  $16 \mu\text{g/ml}$  and  $50 \mu\text{g/ml}$ , respectively (Fig.2). Compared to full length D3, the inhibition by the D3 Fab was weaker ( $\text{IC}_{50} 36 \mu\text{g/ml}$ ), most likely due to reduced avidity for surface TF as compared to the bivalent full length D3 antibody. Surprisingly, HTF1 antibody did not inhibit at the highest tested concentration of  $50 \mu\text{g/ml}$ , while 6B4 showed rather weak inhibitory activity with about 40% inhibition at  $150 \mu\text{g/ml}$  (Fig.2). Consistent with the observed reduction of FPA levels by D3, D3 Fab and 5G6, a visual inspection of the cell layers showed that only little if any fibrin was deposited, while HTF1 and 6B4-treated samples were indistinguishable from controls.

Next, the measurement of FX activation in human plasma was used as another way to quantify anticoagulant potencies of the antibodies. Similar to the blood flow system, where antibody is not pre-equilibrated with TF but infused to the flowing blood, the antibodies were added to plasma and coagulation was triggered with relipidated TF<sub>1-243</sub>. We found that the tested antibodies inhibited the initial rates of FX activation in a concentration-dependent manner (Fig.3). The antibodies D3 and 5G6 were more potent than HTF1, 6B4 and 7G11. The concentrations which inhibited the rates by 50% were as follows:  $7.2 \pm 1.0 \mu\text{g/ml}$  ( $\pm\text{SD}$ ,  $n=5$ ) for D3,  $15.5 \pm 1.3 \mu\text{g/ml}$  ( $\pm\text{SD}$ ,  $n=5$ ) for 5G6,  $43.4 \pm 6.8 \mu\text{g/ml}$  ( $\pm\text{SD}$ ,  $n=4$ ) for 6B4,  $147.8 \pm 8.6 \mu\text{g/ml}$  ( $\pm\text{SD}$ ,  $n=5$ ) for 7G11 and  $150.0 \pm 31.1 \mu\text{g/ml}$  ( $\pm\text{SD}$ ,  $n=4$ ) for HTF1.

Furthermore, similar potency differences between the antibodies were found when clotting times were measured in PT assays using the same incubation protocol as for FX activation rate assays (Fig.4). The antibody concentrations which prolonged the clotting time by 1.5-fold were  $10 \mu\text{g/ml}$  for D3,  $27 \mu\text{g/ml}$  for 5G6,  $133 \mu\text{g/ml}$  for 6B4 and  $500 \mu\text{g/ml}$  for 7G11. The highest tested concentration of HTF1 ( $40 \mu\text{g/ml}$ ) had no effect (Fig.4).

#### Determination of kinetic constants of anti-TF antibodies.

Because the inhibitory potencies of the examined antibodies could merely be a reflection of their binding affinities to sTF, we determined the kinetic constants of each antibody. A comparison of the calculated  $K_D$  values (Table 2) and the inhibitory activities of each antibody showed that there is no correlation between affinities and anticoagulant potencies. In fact, D3 was consistently the most potent anticoagulant, yet it displayed the weakest affinity for TF, while HTF1 and 7G11 were the strongest binders, but had the weakest anticoagulant activities. This lack of correlation was also seen when on-rates were compared which, with the exception of HTF1, were in a similar range ( $2.3 \times 10^5 - 6.0 \times 10^5 \text{ M}^{-1} \text{ sec}^{-1}$ ).

**Table 2**

Antibody	$k_{on}$ ( $10^5 M^{-1} s^{-1}$ )	$k_{off}$ ( $10^{-4} s^{-1}$ )	$K_D$ (nM)
D3	2.43	17.3	7.00
D3 Fab	2.40	27.5	11.50
5G6	3.00	5.00	1.70
7G11	6.00	1.20	0.20
6B4	2.26	13.4	5.90
HTF1	0.80	1.15	1.40

In addition, competition experiments with FVIIa and sTF showed that in the presence of molar excess of FVIIa (>100x), the antibodies 7G11, 6B4 and HTF1 did not bind to TF, whereas the affinity of D3 and 5G6 was only reduced by 4-5-fold. This was consistent with the results from amidolytic assays (fig.1a), indicating that D3 and 5G6 had a fundamentally different inhibitory mechanism than the other antibodies.

Determination of antibody epitopes.

The results so far indicated that the antibodies' anticoagulant potencies could be related to their specific inhibitory mechanism and, thus, to the precise binding site on TF. To determine the antibody epitopes, a large number of sTF mutants were generated by expression in E.coli and subsequent affinity purification on a D3 column (Kelley et al., [1995] supra). The binding of the antibodies to each sTF mutant was measured on a BIAcore instrument. The results, summarized in Fig.5, show the affinity loss expressed as the ratio of  $K_D$  values of sTF mutant and wild-type sTF. Residues that increased the ratio by more than 3-fold were considered important for antibody binding. The two double mutants sTF N199A:R200A and sTF K201A:D204A did not bind to the D3 affinity column and were purified on a 7G11 antibody affinity column. As expected, D3 had the largest loss in affinity to these two mutants ( $K_D$  (mut)/  $K_D$  (wt) >5000). Since sTF D204A alone had the same affinity to the antibodies as wildtype sTF, we concluded that K201 in the K201A:D204A double mutant was the critical residue for antibody binding. Other residues found to be important for D3 binding were I152, Y156 and K165:K166. With the exception of Y156L, the same sTF mutants also showed decreased binding to the 5G6 antibody.

In contrast to D3 and 5G6 which bound to the C-terminal TF domain, the three antibodies 7G11, 6B4 and HTF1 bound to residues located in the N-terminal TF domain. The

TF mutants which had the greatest loss in affinity to 7G11 were K46A (5000x) and Y51A (32x). Additional mutants with significant affinity losses were S47A, K48A, F50A and T52A. The sTF mutants that affected binding of the 6B4 antibody were Y10A, F76A, Y94A, E99A and L104A:E105A. Three of these mutants, F76A, Y94A and E99A, also reduced binding  
5 affinity of the HTF1 antibody (Fig.5).

Location of antibody epitopes on the crystal structure of the TF:FVIIa complex.

As seen in the crystal structure of the TF:FVIIa complex (Banner et al., [1996] supra), the 7G11 binding site is formed by a clearly defined patch of surface exposed residues with a calculated solvent accessible area of  $397\text{\AA}^2$  (Fig. 6). This is relatively small compared to  
10 buried surface areas of typical antigens (Huang et al., [1998] supra) and thus, the antibody binding region may not have been identified in its entirety. This region is important for FVIIa binding and the F50 residue makes a critical hydrophobic contact to the second epidermal growth factor domain of FVIIa (Banner et al., [1996] supra).

The TF residues that were important for 6B4 binding defined a large surface area  
15 located on the 'back side' of TF as compared to the 7G11 epitope (Fig.6). With the premise that the epitope is within the perimeter defined by the identified TF residues, the area of the hypothetical epitope was calculated to be  $594\text{\AA}^2$ . The epitope extended into a TF region which contacts the catalytic domain of FVIIa and included the residues F76 and Y94. For the HTF1 antibody only a relatively small number of TF mutants were examined and the epitope  
20 is somewhat less well defined (Fig.6). Nevertheless, the HTF1 and 6B4 epitopes were largely the same, since three identified binding residues including F76 and Y94 were shared by both antibodies.

The epitopes of the D3 and 5G6 antibodies were very similar, being located outside of the FVIIa-TF contact region. As shown in Figure 6, the epitope runs from the bottom to the  
25 top of the C-terminal TF domain and is approximately opposite to the main TF-FVIIa contact region. Accordingly, antibody binding may not interfere with TF:FVIIa complex formation.

DISCUSSION

By using a large panel of sTF mutants the binding epitopes of 5 anti-TF antibodies was determined and a clear picture of how they exert their anticoagulant effect gained. They  
30 bound to three distinct regions of TF and either interfered with FVIIa-TF association (7G11, 6B4, HTF1) or with TF-macromolecular substrate interaction (D3, 5G6). The anticoagulant potencies were determined in whole blood and plasma-based systems in which the antibodies and coagulation factors were allowed to simultaneously interact with TF. First, in the human

ex-vivo blood flow system D3 and 5G6 potentially inhibited the generation of FPA and the deposition of fibrin onto the J82 cell layer. In contrast, the antibodies 6B4 and HTF1 were at least one order of magnitude less potent and, in the case of HTF1, virtually inactive.

Secondly, since coagulation in the blood flow system was shown to proceed via direct  
5 TF:FVIIa mediated activation of FX (Kirchhofer et al., [1995] supra), it was reasoned that the differential effects on FPA generation should have a correlate in the inhibition of FX activation rates and prothrombin times in human plasma. Indeed, it was found that D3 and 5G6 were by far the most potent antibodies tested.

The apparent differences in anticoagulant activities could not be explained by  
10 differences in the binding affinities of the antibodies. In fact, D3 was the weakest binder ( $K_D$  7nM) and 7G11 the strongest ( $K_D$  0.2nM), yet D3 was about 20-fold more potent in inhibiting FX activation. Because the TF-initiated coagulation is a rapid process it seemed possible that the potency differences reflected differences in on-rates. However, with the exception of HTF1, the experimentally determined on-rates were within a narrow range and  
15 showed no correlation with the observed anticoagulant potencies. These findings suggested that the epitope locations rather than on-rate constants or binding affinities were the main determinants of anticoagulant potencies of the anti-TF antibodies.

Inspection of the epitopes on the crystal structure of the TF:FVIIa complex revealed that all antibodies bound to functionally important regions of TF, but impacted different  
20 aspects of TF function. The binding epitope of D3 and 5G6 overlapped with a TF region that does not contact FVIIa, but interacts with macromolecular substrate binding. This explained why the antibodies had little or no effect on amidolytic activity towards a small synthetic substrate. The epitope residues K165 and K166 were previously found to be critical for TF:FVIIa mediated activation of FX (Ruf et al., [1992] supra; Roy et al., [1991] supra; Kelley  
25 et al., [1995] supra) and FIX (Huang et al., [1996] supra). These two residues and the additional epitope residues Y156 and K201 are all part of a distinct surface-exposed TF region which directly interacts with substrates FX and FIX (Kirchhofer et al., [1999] *Thromb. Haemost. Suppl.* [abstract], 300; Kirchhofer et al., (2000) *Biochemistry* 39:7380-7387). This region may extend into the FVIIa Gla domain (Martin et al., [1993] supra; Ruf et al., [1999]  
30 supra) and most likely contacts the Gla domains of substrates (Huang et al., [1996] supra; Martin et al., [1993] supra). Thus, D3 and 5G6 by binding to this region will sterically prevent the association of the substrate Gla domains with TF and thus interfere with proper substrate orientation to form a productive ternary TF-FVIIa-substrate complex. This further provided a basis of explaining the excellent anticoagulant potencies of D3 and 5G6. First, since the

epitope is not within the TF-FVIIa contact region, these antibodies were able to bind to TF during and after the rapid formation of TF:FVII and TF:FVIIa complexes in plasma. Secondly, they competed with a rather low affinity TF-substrate interaction event. Moreover, the epitope residues K165 and K166 were shown to be important for the FVIIa and FXa-dependent activation of TF:FVII (Dittmar et al., [1997] *Biochem. J.* 321: 787-793). Therefore, both antibodies could have interfered with the activation of TF-bound zymogen FVII, a reaction which in all likelihood is the first activation step for coagulation in plasma (Rapaport and Rao [1995] *Thromb. Haemost.* 74: 7-17), as well as in the employed blood flow system with J82 cells (Sakai et al., [1989] *J. Biol. Chem.* 264: 9980-9988). In fact, D3H44-F(ab')<sub>2</sub> was found to strongly inhibit the FXa-mediated activation of zymogen FVII (Kirchhofer et al., (2001) *Biochemistry* 40:675-682). In agreement, the closely related antibody TF8-5G9 (Morrissey et al., [1988] *supra*; Ruf and Edgington [1991] *supra*; Huang et al., [1998] *supra*; Fiore et al., [1992] *supra*) which binds to the same TF region (Huang et al., [1998] *supra*), is a potent anticoagulant in plasma clotting assays (Ruf and Edgington [1991] *supra*) and inhibits TF-dependent FVII conversion to FVIIa (Fiore et al., [1992] *supra*). A comparison with TF8-5G9 revealed that all of the identified D3 and 5G6 epitope residues were also found as contact residues in the crystal structure of TF8-5G9 Fab bound to soluble TF (Huang et al., [1998] *supra*). Yet, despite having an apparently identical epitope the D3 and TF8-5G9 (Fiore et al., [1992] *supra*) antibodies differed from 5G6 in their ability to weakly inhibit the amidolytic activity (Chromozym t-PA) of the sTF:FVIIa complex at low sTF concentrations. These results indicated that there existed subtle differences in antibody binding to an apparently identical epitope.

The weaker anticoagulant potencies of 7G11, HTF1 and 6B4 were surprising, since they were as potent as D3 and 5G6 when allowed to pre-bind to TF in plasma clotting assays. The identification of the antibody binding sites on TF provided a basis for explaining these results. 7G11 bound to a TF region proximal to the light chain of FVIIa. One of the binding residues, F50, makes an important contact to the second epidermal growth factor domain of FVIIa (Banner et al., [1996] *supra*; Zhang et al., [1999] *J. Mol. Biol.* 285: 2089-2104) suggesting that the antibody interfered with the formation of the TF:FVIIa complex. Both 6B4 and HTF1 interfered with a shared FVIIa contact site, which was distinct from the 7G11 epitope. In agreement, 6B4 did not prevent binding of 7G11 to TF in cross-blocking experiments (data not shown). Epitope residues F76 and Y94 make direct contacts to the FVIIa catalytic domain residue Met306. Mutating this Met306 or TF residue Y94 strongly impaired macromolecular substrate activation (Dickinson et al., [1996] *supra*; Kelley et al.,



[1995] *supra*; Ruf et al., [1995] *Biochemistry* 34: 6310-6315), and binding of 6B4 or HTF1 should consequently have deleterious effects as well. This contact site is also important for the TF-dependent enhancement of FVIIa activity towards small synthetic substrates (Dickinson et al., [1996] *supra*), thus explaining the observed inhibitory effects of 6B4 and HTF1 in the  
5 amidolytic assay. The results are also consistent with a previous report demonstrating that HTF1 interfered with the binding of TF to FVIIa (Carson et al., [1987] *supra*). There is a possibility that 6B4 had additional effects on the macromolecular substrate-TF interaction. 6B4 did not bind to the main substrate interaction region around K165 and K166 (fig.6), but the epitope residues L104:E105 were proximal to residues N199:R200 which are part of the  
10 FX recognition region (Kirchhofer et al., [1999] *supra*). Thus, 6B4 binding could have resulted in additional steric effects on substrate-TF interaction.

A distinguishing characteristic of antibodies 7G11, 6B4 and HTF1 is their competition with local FVIIa contact sites within the context of an overall large contact surface (Banner et al., [1996] *supra*) and high affinity FVII-TF interaction. Whereas this inhibitory mechanism  
15 provided potent inhibition in pre-incubation experiments (Fig.1b), it did not do so under the non-equilibrium conditions of our experimental systems. One likely explanation is that once TF:FVIIa complexes were formed, the antibodies would have little inhibitory effect since inhibition would mainly be determined by the FVIIa/TF off-rate. Similar conclusions were made by Ruf and Edgington (1991, *supra*) and Fiore et al. (1992, *supra*) using different in-  
20 vitro systems to evaluate two antibodies which interfered with TF-FVIIa association. Nevertheless, at appropriate doses such types of antibodies demonstrated inhibition of the coagulation system in animal experiments (Taylor et al., [1991] *Circulatory Shock* 33: 127-134; Himber et al., [1997] *Thromb. Haemostasis* 78: 1142-1149; Pawashe et al., [1994] *Circ. Res.* 74: 56-63; Ragni et al., [1996] 93: 1913-1918; Thomas et al., [1993] *Stroke* 24: 847-854;  
25 Golino et al., [1996] *Nature Med.* 2: 35-40). A caveat to this comparison is that the antibodies used for the animal experiments have not been characterized in much detail and no epitope map information is available. Furthermore, the experiments would predict that an antibody like D3 or 5G6 should be efficacious at significantly lower doses. Consistent with this view the closely related TF8-5G9 antibody appeared extremely potent in inhibiting coagulation in a  
30 chimpanzee study (Levi et al., [1994] *J. Clin. Invest.* 93: 114-120) and was also very effective in a tumor metastasis model (Mueller et al., [1992] *supra*). However, a direct comparison of two well-defined, different type-antibodies has yet to be done.

The findings suggested that the anticoagulant potencies of anti-TF antibodies is not primarily determined by the binding affinity, but rather by the epitope location and

consequently by the particular mode of inhibition. Even though the translation of the results obtained from blood/plasma based in-vitro systems into an in-vivo setting has obvious limitations, the findings may nevertheless have some implications in regard to the use of anti-TF antibodies in anticoagulant therapy. As suggested by this study, the choice of an anti-TF antibody may be important in terms of the expected efficacy, since the epitope location will strongly influence the antibody's potency to inhibit thrombosis.

## EXAMPLE 2

### Humanization of a murine anti-human tissue factor monoclonal antibody D3.

#### MATERIALS AND METHODS

##### Cloning of Murine D3.

The murine anti-human TF mAb D3 was generated and cloned at Genentech (Paborsky et al., [1990] *Prot. Engineering* 3: 547-553). Protein sequence analysis of the purified antibody provided an N-terminal sequence for the heavy chain, EVQLQQSGAELVRPGALVKLSCKASGFNIKD (SEQ ID NO:19), and for the light chain, DIKMTQSPSSMSASLGESVTITCKASRDIK (SEQ ID NO:20). Total RNA was purified from D3 hybridoma cell line (1D4(14)\_D3) using the standard RNA STAT protocol (Tel-Test-Inc., Friendswood, TX). cDNA was made using Oligo dT and Superscript II RNase H-Reverse Transcriptase according to the manufacturers instructions (Gibco BRL, Gaithersburg, MA). PCR amplification was performed in a 50 µl reaction using 3 units of UITma DNA Polymerase (Perkin Elmer, Foster City, CA) with 1x buffer, 4 mM MgCl, 40 µM dNTPs, and 0.7 - 1.0 µM forward and reverse primers. Specific primers used were heavy chain forward, 5'-TACAAACGCGTACGCTGARGTNCARYTNCARCARWSNNGGNGC-3' (SEQ ID NO:21), heavy chain reverse, 5'-CAGTGGATAGACAGATGGGC CCGTCGTTTTGGC-3' (SEQ ID NO:22), light chain forward, 5'-GCATACGCTGAYATHAARATGACNCARWSNCC-3' (SEQ ID NO:23), light chain reverse, 5'-TG GTGCAGCCACGGTCCGTTTKAKYTCCARYTTKGT-3' (SEQ ID NO:24). Separate reactions were set up for heavy chain and light chain and cycled with the following conditions: 95° for 2 min, 30 cycles of (95° 20 sec, 60° 30 sec, 72° 1 min), 4° hold in a Perkin Elmer 9600. After purification on Qiaquick columns (Qiagen), 1/10 of the PCR reaction was cloned into pCR-Blunt (Invitrogen). Sequence analysis of the clones revealed that amino acid 7 of both the heavy and light chains were Arg instead of the expected Ser; the codon WSN used in the primers can result in Arg or Ser. The PCR was repeated on individual clones in pCR-Blunt to change the Arg to Ser. The same reverse primers were used and new heavy

chain forward primer, 5'-

AGGTACAAACGCGTACGCTGAAGTGCAACTCCAGCAAAGTGG-3' (SEQ ID NO:25)

and light chain forward primer, 5'-GCATACGCTGAT

ATAAAAATGACGCAGTCGCCATCC-3' (SEQ ID NO:26). PCR used UII<sup>Tma</sup> and the

5 same conditions as above. The heavy chain PCR fragment was digested with BsiWI and ApaI while the light chain PCR fragment was digested with EcoRV and RsrII. Each resulting digested fragment was cloned into a previously described F(ab) chimeric expression plasmid (Presta et al., Cancer Res. 57: 4593-4599 [1997]).

DNA sequence of heavy chain fragment BsiWI to ApaI (SEQ ID NO:27):

10 5'-

GTACGCTGAAGTGCAACTCCAGCAAAGTGGCGCTGAGCTTGTGAGGCCAGGGCC  
TTAGTCAAGTTGTCCTGCAAAGCTTCTGGCTTCAACATTAAAGACTACTATATGCA  
CTGGGTGAAGCAGAGGCCTGAACAGGGCCTGGAGTTGATTGGATGGATTGATCCT  
GAGAATGGTAATACTATTTATGACCCGAAGTTCAGGACAAGGCCAGTATAACA  
15 GCAGACACATCCTCCAACACAGCCTACCTGCAGCTCAGCAGCCTGACATCTGAGG  
AACTGCCGTCTATTACTGTGCTAGAGATACTGCGGCATACTTTGACTACTGGGG  
CCAAGGCACCACTCTCACAGTCTCCTCAGCCAAAACGACGGGCCC-3'

DNA sequence of light chain fragment EcoRV to RsrII (SEQ ID NO: 28):

5'-

20 GATATCAAAATGACGCAGTCGCCATCCTCCATGTCTGCATCGCTGGGAGAGAGTG  
TCACTATCACTTGCAAGGCGAGTCGGGACATTAAAAGCTATTTAAGCTGGTACCA  
GCAGAAACCATGGAAATCTCCTAAGACCCTGATCTATTATGCCACAAGCTTGGCG  
GATGGGGTCCCATCAAGATTCAGTGGCAGTGGATCTGGGCAAGATTATTCTCTAA  
CCATCAGCAGCCTGGAGTCTGACGATACAGCAACTTATTACTGTCTACAGCATGG  
25 TGAGAGCCCATTCACGTTTCGGCTCGGGGACAAAGTTGGAACCTCAAACGGACCG-3'

Computer Graphics Models of Murine and Humanized F(ab)s.

Sequences of the VL and VH domains (SEQ ID NOS: 3 and 1, respectively) were used to construct a computer graphics model of the murine D3 VL-VH domains. This model was used to determine which framework residues should be incorporated into the humanized antibody. A model of the humanized F(ab) was also constructed to verify correct selection of murine framework residues. Construction of models was performed as described previously (Carter et al., Proc. Natl. Acad. Sci. USA 89: 4285-4289 [1992]; Eigenbrot et al., J. Mol. Biol. 229: 969-995 [1993]).

Construction of Humanized F(ab)s.

The plasmid pEMX1 used for mutagenesis and expression of F(ab)s in *E. coli* has been described previously (Werther et al., J. Immunol. 157: 4986-4995 [1996]). Briefly, the plasmid contains a DNA fragment encoding a consensus human  $\kappa$  subgroup I light chain (VL $\kappa$ I-CL), a consensus human subgroup III heavy chain (VHIII-CH1) and an alkaline phosphatase promoter. The use of the consensus sequences for VL and VH has been described previously (Carter et al., *supra*).

To construct the first F(ab) variant of humanized D3, F(ab)-1, site-directed mutagenesis (Kunkel, Proc. Natl. Acad. Sci. USA 82: 488-492 [1985]) was performed on a deoxyuridine-containing template of pEMX1. The six CDRs were changed to the murine D3 sequence; the residues included in each CDR were from the sequence-based CDR definitions (Kabat et al., Sequences of proteins of immunological interest, Ed. 5, Public Health Service, National Institutes of Health, Bethesda, MD [1991] except for CDR-H1 which was defined using a combination of CDR-H1 definitions from Kabat et al. (*supra*) and Chothia et al., Nature 342:877-833 (1989), i.e., CDR-H1 was defined as extending from residues H26-H35 in the heavy chain). F(ab)-1 therefore consisted of a complete human framework (VL $\kappa$  subgroup I and VH subgroup III) with the six complete murine CDR sequences. Plasmids for all other F(ab) variants were constructed from the plasmid template of F(ab)-1. Plasmids were transformed into *E. coli* strain XL-1 Blue (Stratagene, San Diego, CA) for preparation of double- and single-stranded DNA. For each variant, DNA coding for light and heavy chains was completely sequenced using the dideoxynucleotide method (Sequenase, U.S. Biochemical Corp., Cleveland, OH). Plasmids were transformed into *E. coli* strain 16C9, a derivative of MM294, plated onto Luria broth plates containing 50  $\mu$ g/ml carbenicillin, and a single colony selected for protein expression. The single colony was grown in 5 ml Luria broth-100  $\mu$ g/ml carbenicillin for 5-8 h at 37°C. The 5 ml culture was added to 500 ml AP5-50  $\mu$ g/ml carbenicillin and allowed to grow for 20 h in a 4 L baffled shake flask at 30°C. AP5 media consists of: 1.5 g glucose, 11.0 g Hycase SF, 0.6 g yeast extract (certified), 0.19 g MgSO<sub>4</sub> (anhydrous), 1.07 g NH<sub>4</sub>Cl, 3.73 g KCl, 1.2 g NaCl, 120 ml 1 M triethanolamine, pH 7.4, to 1 L water and then sterile filtered through 0.1  $\mu$ m Sealkeen filter. Cells were harvested by centrifugation in a 1 L centrifuge bottle at 3000xg and the supernatant removed. After freezing for 1 h, the pellet was resuspended in 25 ml cold 10 mM Tris-1 mM EDTA-20% sucrose, pH 8.0 250  $\mu$ l of 0.1 M benzamidine (Sigma, St. Louis, MO) was added to inhibit proteolysis. After gentle stirring on ice for 3 h, the sample was centrifuged at

40,000xg for 15 min. The supernatant was then applied to a protein G-Sepharose CL-4B (Pharmacia, Uppsala, Sweden) column (0.5 ml bed volume) equilibrated with 10 mM Tris-1 mM EDTA, pH 7.5. The column was washed with 10 ml of 10 mM Tris-1 mM EDTA, pH 7.5, and eluted with 3 ml 0.3 M glycine, pH 3.0, into 1.25 ml 1 M Tris, pH 8.0. The F(ab) was then buffer exchanged into PBS using a Centricon-30 (Amicon, Beverly, MA) and concentrated to a final volume of 0.5 ml. SDS-PAGE gels of all F(ab)s were run to ascertain purity and the concentration of each variant was determined by amino acid analysis. F(ab)s were quantified by measuring OD<sub>280</sub> and amino acid analysis; concentrations used in assays were from the amino acid analysis.

A chimeric F(ab) was used as the standard in the binding assays. This chimeric F(ab) consisted of the entire murine D3 VH domain fused to a human CH1 domain at amino acid SerH113 and the entire murine D3 VL domain fused to a human CL domain at amino acid LysL107. Expression and purification of the chimeric F(ab) were identical to that of the humanized F(ab)s.

#### Construction and purification of D3H44-F(ab')<sub>2</sub>

D3H44-F(ab')<sub>2</sub> was generated by the addition of the heavy chain hinge (CPPCPAPELLGG) to the C-terminus of the D3H44-F(ab), followed by the GCN4 leucine zipper (51) and a (his)<sub>6</sub> tag for purification. D3H44-F(ab')<sub>2</sub> was expressed in E. coli and the cell paste was diluted 1:5 (w/v) in 20 mM sodium phosphate pH 7.4, 50 mM NaCl, then lysed using an M110Y microfluidizer (Microfluidics Corp., Newton, MA). Polyethylene imine (BASF Corp., Rensselaer, NY) was added to a final concentration of 0.2%, followed by centrifugation (4300xg, 30min) to remove cellular debris. The supernatant was filtered (0.2 µm) and loaded on to SP Sepharose FF (Amersham Pharmacia Biotech, Uppsala, Sweden) under conditions in which F(ab')<sub>2</sub> flowed through. The SP Sepharose FF flow through fraction was applied to Chelating Sepharose FF (Amersham Pharmacia Biotech, Uppsala, Sweden), charged with Cu<sup>2+</sup> and equilibrated in 2 mM imidazole, pH 7.0, 250 mM NaCl. D3H44-F(ab')<sub>2</sub> was eluted using 200 mM imidazole pH 7.0. The Chelating Sepharose FF elution pool was adjusted to pH 4.0, and the leucine zipper/(his)<sub>6</sub> tag was cleaved using pepsin. Following pepsin cleavage, D3H44-F(ab')<sub>2</sub> was applied to SP Sepharose High Performance (Amersham Pharmacia Biotech, Uppsala, Sweden) and eluted using a linear gradient from 0 to 0.12 M sodium acetate in 25 mM MES pH 5.6. SP Sepharose High Performance gradient fractions were analyzed by SDS-PAGE and pooled. Finally, D3H44-F(ab')<sub>2</sub> was formulated by ultrafiltration using a 10 kDa regenerated cellulose membrane (Millipore Corp., Bedford, MA), followed by diafiltration into 20 mM sodium acetate pH 5.5,

0.14 M NaCl. Formulated D3H44-F(ab')<sub>2</sub> purity was >99.9% by an E. coli protein impurity assay. The endotoxin level in the formulated D3H44-F(ab')<sub>2</sub> was <0.01 EU/mg.

Construction of Chimeric and Humanized IgG.

For generation of human IgG2 and IgG4 variants of humanized D3, the humanized VL and VH domains from (F(ab)-D3H44) were subcloned separately into previously described pRK vectors (Eaton et al., Biochemistry 25: 8343-8347 [1986]) containing the constant domains of human IgG2 or IgG4. The IgG4b variant includes a Ser H241 Pro change that improves formation of the inter-heavy chain disulfides in the hinge, resulting in a more homogeneous production of IgG4 antibody (Angal S, King DJ, Bodmer MW, Turner A, Lawson AD, Roberts G, Pedley b, Adair JR. A single amino acid substitution abolishes the heterogeneity of chimeric mouse/human (IgG4) antibody. Molec. Immunol. 1993;30:105-108; Bloom JW, Madanat MS, Marriot D, Wong T, Chan S-Y. Intrachain disulfide bond in the core hinge region of human IgG4. Prot. Sci. 1997;6:407-415). The DNA coding for the entire light and the entire heavy chain of each variant was verified by dideoxynucleotide sequencing. The IgG variants were purified using Protein A-Sepharose.

Construction and Purification of IgG1

Tissue Factor Binding Assay.

Maxisorp plates (Nunc, Roskilde, Denmark) were coated overnight at 4°C with 100 µl/well of 10 µg/ml human soluble tissue factor in coat buffer (50 mM carbonate buffer, pH 9.6). The plates were blocked with 150 µl/well blocking buffer (PBS, 0.5% BSA, pH 7.2) for 1 h at room temperature. The standard and samples were diluted in assay buffer (PBS, 0.5% BSA, 0.05% Tween20, pH 7.2) and incubated on the plates for 2 h at room temperature. 100 µl of 1:10,000 goat anti-human F(ab)-HRP (Cappel, Costa Mesa, CA) was added and the plates were incubated for 1 h at room temperature. 100 µl of the substrate 3,3',5,5'-tetramethyl benzidine (TMB) (Kirkegaard & Perry, Gaithersburg, MD) was added. After 5 min, 100 µl of 1 M H<sub>3</sub>PO<sub>4</sub> was added to stop the reaction. The plate was washed with wash buffer (PBS, 0.05% Tween 20, pH 7.2) between each step. The absorbance was read at 450 nm on a Titerek stacker reader (ICN, Costa Mesa, CA). The standard and samples were fit by Kaleidagraph 3.0.8 (Synergy Software, Reading, PA) using a four parameter fit regression. The OD<sub>450</sub> at the IC<sub>50</sub> of the standard was determined. The concentration of sample needed to obtain this OD was determined and the ratio of this value versus the IC<sub>50</sub> of the standard was calculated.

BIAcore™ Biosensor Assays.

TF binding of the humanized and chimeric F(ab)s were compared using a BIAcore™ biosensor (Karlsson et al., 1994). Concentrations of F(ab)s were determined by quantitative amino acid analysis. TF was coupled to a CM-5 biosensor chip through primary amine groups according to manufacturer's instructions (Pharmacia). Off-rate kinetics were measured by saturating the chip with F(ab) (35 µl of 2 µM F(ab) at a flow rate of 20 µl/min) and then switching to buffer (PBS-0.05% polysorbate 20). Data points from 0 - 4500 sec were used for off-rate kinetic analysis. The dissociation rate constant ( $k_{off}$ ) was obtained from the slope of the plot of  $\ln(R_0/R)$  versus time, where  $R_0$  is the signal at  $t=0$  and  $R$  is the signal at each time point.

On-rate kinetics were measured using two-fold serial dilutions of F(ab) (0.0625 - 2 µM). The slope,  $K_s$ , was obtained from the plot of  $\ln(-dR/dt)$  versus time for each F(ab) concentration using the BIAcore™ kinetics evaluation software as described in the Pharmacia Biosensor manual.  $R$  is the signal at time  $t$ . Data between 80 and 168, 148, 128, 114, 102, and 92 sec were used for 0.0625, 0.125, 0.25, 0.5, 1, and 2 µM F(ab), respectively. The association rate constant ( $k_{on}$ ) was obtained from the slope of the plot of  $K_s$  versus F(ab) concentration. At the end of each cycle, bound F(ab) was removed by injecting 5 µl of 50 mM HCl at a flow rate of 20 µl/min to regenerate the chip.

BioassaysReagents.

F.IX was from Haematologic Technologies Inc., (Essex Jct., VT) and F.X was from Enzyme Research Laboratories (South Bend, IN). Dioleoyl 1,2-diacyl-sn-glycero-3-(phospho-L-serine) (PS) and oleoyl 1,2-diacyl-sn-glycero-3-phosphocholine (PC) from Avanti Polar Lipids Inc. (Alabaster, AL). F.IXa chromogenic substrate #299 was from American Diagnostica (Greenwich, CT) and FXa chromogenic substrate S-2765 was from Diapharma Group Inc. (Columbus, Ohio). Innovin was obtained from Dade International Inc. (Miami, FL). Ethyleneglycol (analytical grade) was from Mallinckrodt Inc. (Paris, KY). Fatty acid-free BSA was from Calbiochem (Calbiochem-Novabiochem Corp., La Jolla, CA). TF (1-234) lacking the cytoplasmic domain was produced as described (Paborsky et al., (1989) Biochemistry 28:8072; Paborsky et al., (1991) J. Biol. Chem. 266:1911) and relipidated with PC/PC (7:3 molar ratio) according to Mimms et al., (1981) Biochemistry 20:833-840).

Activation of FIX by membrane tissue factor (mTF):FVIIa complex.

Membrane TF (mTF) was prepared from a human embryonic kidney cell line (293) expressing full length TF (1-263) (Kelley et al., Blood 89: 3219-3227 [1997]). The cells were washed in PBS, detached with 10 mM EDTA and centrifuged twice (2500 rpm for 10 min).

5 The cell pellet ( $4-5 \times 10^7$  cells/ml) was resuspended in Tris, pH 7.5, and homogenized in PBS using a pestle homogenizer, followed by centrifugation (2500 rpm on a Beckman GSA) for 40 min at 4°C. The protein concentration of the cell membrane fraction was determined and the membranes stored in aliquots at -80°C until use.

Prior to the addition of F.IX, the antibodies were incubated in microtiter tubes (8.8x45 mm, OPS, Petaluma, CA) together with mTF and FVIIa in HBSA buffer (20 mM Hepes, pH 7.5, 150 mM NaCl, 5 mM CaCl<sub>2</sub>, 0.5 mg/ml BSA) for 20 min at room temperature. The final concentration in the reaction mixture for the reactants were as follows: 150 µg/ml mTF (membrane protein concentration), 2 nM FVIIa and 400 nM F.IX in HBSA. 100 µl aliquots of the reaction mixture were taken at 30 s intervals and quenched in 96-well plates (Costar, 15 Corning Inc., Corning, NY) containing 125 µl of 30 mM EDTA-buffer-60% (v/v) ethyleneglycol. After adding 25 µl of 5 mM F.IX substrate #299, F.IXa amidolytic activity was measured at 405 nm on a kinetic microplate reader (Molecular Devices, Menlo Park, CA). Inhibition by the tested antibodies was expressed as fractional rates (vi/vo) of F.IXa generation.

20 Activation of FX by mTF:FVIIa complex.

The experiments were carried out in a similar fashion as described for F.IX activation. The concentration in the reaction mixture of the reactants were as follows: 200 nM FX, 150 µg/ml mTF, 30 pM FVIIa in HBSA. At 30 s intervals, 50 µl aliquots were quenched in 150 µl 20 mM EDTA and the FXa amidolytic activity measured by adding 50 µl of 1.5 mM S- 25 2765.

## RESULTS

Transplanting the murine D3 CDRs onto the human framework (VL $\kappa$  subgroup I, VH subgroup III) (Carter et al., Proc. Natl. Acad. Sci. USA 89: 4285-4289 [1992]; Presta et al., 30 Cancer Res. 57: 4593-4599 [1997]) resulted in a F(ab) which lacked binding to human TF. Based on the computer graphic model of murine D3 F(ab) (Fig. 7), several amino acid residues in the CDRs as well as framework region of light and heavy chains were altered using site-directed mutagenesis in order to optimize antigen binding. The engineered antibody



thus evolved, D3H44 F(ab), exhibited acceptable binding and efficacy in all of the biological assays, including the prothrombin time assays Figs. 10-12. D3H44 has four human-to-murine changes in its heavy chain framework: Gly H49, Ala H67, Ala H71, and Ala H78 (Fig. 8). D3H44 also has one human-to-murine change in its light chain framework, Tyr L71, as well as one change which is neither human nor murine, Val L46. In the CDRs, D3H44 has seven differences from the murine D3 parent: Glu H31 (CDR-H1), Leu H50 and Gln H54 (CDR-H2), Arg L24 and Asn L34 (CDR-L1), Glu L56 (CDR-L2), and Trp L96 (CDR-L3) (Figs. 8, 9).

Since a crystal structure of huTF-TF8-5G9 (Huang et al., J. Mol. Biol. 275: 873-894 [1998]) was available in the public Protein Data Bank crystal structure database (coordinates PDB1AHW), the effect of altering some of the sequence of the chimeric D3 F(ab) to that of TF8-5G9 was investigated. First, three residues in CDR-H3 were altered: D3Ch Thr96-Ala97-Ala98 to TF8 Asn96-Ser97-Tyr98. This resulted in a 20-fold reduction in binding ( $20.3 \pm 0.69$ ,  $n=2$ ). Given that these CDR-H3 residues interact with huTF in the crystal structure, the severe reduction in binding was unexpected. Second, in CDR-H2 D3Ch Asp H65 was changed to TF8 Gly; binding was reduced by 14-fold ( $14.2 \pm 2.7$ ,  $n=3$ ). Inspection of the huTF-TF8 crystal structure shows that residue H65 is not in contact with huTF and the change to Gly should not have affected binding. Taken together, these data suggest that the D3 antibody does not bind to huTF in the same manner as TF8-5G9.

Binding of anti-tissue factor antibodies (IgG1, IgG2, IgG4 and IgG4b) to tissue factor is shown in Figure 16. Each of E. coli produced IgG1 and CHO produced IgG2, IgG4 and IgG4b bound immobilized TF.

Inhibition of the rates of F.X and F.IX activation by full length versions and a F(ab')<sub>2</sub> version of the D3H44 antibody are shown in Table 3.

Table 3

Antibody	F.X activation IC <sub>50</sub> (nM)	F.IX activation IC <sub>50</sub> (nM)
D3H44 IgG1 (n = 3)	0.054	0.138
D3H44 IgG2 (n = 3)	0.073	0.160
D3H44 IgG4 (n = 3)	0.059	0.107
D3H44 IgG4 b(n = 3)	0.048	0.127
D3H44 F(ab') <sub>2</sub> (n = 3)	0.047	N/D

Antibodies were incubated with relip. TF (1-234) (0.04 nM) and F.VIIa (0.04 nM) for 20 min. and the reaction started by adding F.X (200 nM). Aliquots were taken at different time points and quenched in EDTA. In the second stage of the assay, F.Xa activity was measured by adding chromogenic substrate S2765 and monitoring absorbance at 405 nM on a kinetic microplate reader. IC50 values were calculated by non-linear curve fitting using fractional activities (vi/v0) of initial substrate activation rates vs. antibody concentration. For F.IX assays the concentration of reactants was 1 nM relip.TF(1-234), 1 nM F.VIIa, 400 nM F.IX. Reaction aliquots were quenched in EDTA-60% (v/v) ethyleneglycol. In the second stage of the assay, F.IXa activity was measured by adding chromogenic substrate #299 and monitoring absorbance at 405 nM on a kinetic microplate reader. IC50 values were calculated as described above for F.X.

Prolongation of clotting time for the full length versions and Fab and F(ab')2 versions of D3H44 are shown in Figure 17.

### EXAMPLE 3

Co-administration of anti-rabbit TF monoclonal antibody with heparin or a GP IIb/IIIa inhibitor (G4120) results in improved efficacy versus the respective monotherapies and has a better therapeutic profile than a heparin-G4120 combination in a rabbit thrombosis model.

#### Materials and Methods

##### Materials

Mouse monoclonal anti-rabbit TF antibody (AP-1) can be prepared as described in Pawashe et al., [1994] Circ. Res. 74:56. Rabbit tissue factor is available from Diagnostic Reagents (Diagen, Thame Oxon, England). Unfractionated heparin (UFH) is available from Elkins Sinn (Cherry Hills, NJ). Platelet GP IIb/IIIa inhibitor G4120 is prepared as described in Barker et al., J. Med. Chem. 35:2040-2048 (1992).

##### Rabbit Model of Arterial Thrombosis

The model used in this study is a vascular damage model as reported in Refino et al., Arterioscler. Thromb. Vasc. Biol. 22:517 (2002). The protocol used was as follows.

The left common carotid of an isoflurane-anesthetized male New Zealand (3.2 to 4.0 kg) rabbits were surgically isolated, and an ultrasonic flow probes (Transonics Systems) were placed proximal to the aortas. Treatment was initiated before the placement of the flow restrictor and 30 minutes before insertion of the balloon catheter. Lexan flow restrictors (1.2 mm internal diameter) were placed proximal to the segment of the artery to be damaged by the balloon catheter. Addition of the restrictor provided a defined diameter, which was used

to calculate the shear rate adjacent to the damaged arterial segment. Shear rate ( $\gamma$ ) was determined by using the equation  $\gamma=4Q/\pi r^3$ , in which Q = blood flow in cm<sup>3</sup>/s and r= the radius of the lumen in cm. The deflated catheter was inserted into the lumen of the carotid via a small side branch, which was distal to the flow restrictor. The tip of the catheter was guided to the restricted region and the balloon inflated so as to gently dilate the vessel. The catheter was then forcefully pulled back 2 cm and the balloon deflated. This procedure was repeated a total of six times in a 2-minute period. Flow measurements were recorded before and for 60 minutes after the balloon procedure. At 60 minutes, the artery was cross-clamped, and any thrombus present in the damaged section was removed, blotted on filter paper, and weighed.

Anti-thrombotic treatments were administered via the marginal ear vein as a single bolus (AP-1) or as a bolus followed by a continuous infusion (saline, heparin, G4120). Blood samples were collected from a femoral artery catheter before treatment, and at 1, 30, and 60 minutes after the balloon procedure. Blood samples were collected and processed as previously described in Refino et al., Thrombosis and Haemostasis [1999] 82:1188-1195.

Prothrombin times (PT), 2 stage prothrombin time (2st-PT), activated partial thromboplastin times (APTT), and platelet aggregation were determined. Although there were differences in coagulation times between groups, within-group PT, 2st-PT and APTT remained essentially unchanged during the 60 minutes after the balloon injury. Therefore, changes from the pretreatment determination were expressed as a ratio of the 1-minute post-balloon procedure value over the pretreatment value.

Cuticle transections were performed as described for rats and guinea pigs in Refino et al., Thrombosis and Haemostasis [1999] 82:1188-1195. Cuticle-bleeding times were determined before treatment, and at 5 and 30 minutes after the balloon procedure. The 5 minute and 30 minute values were meaned and a post-over pretreatment ratio was calculated from these two determinations.

#### Prothrombin Time Assays

For in vitro prothrombin time (PT) assays, increasing amounts of antibody AP-1 were added to 50 ul citrated rabbit plasma. Antibody concentrations ranged from 0.01 µg/ml to about 1000 µg/ml. After 5 minutes incubation, clotting was started by adding 100 ul thromboplastin (rabbit tissue factor and Ca<sup>++</sup>). Clotting times were measured on ACL300 using the PT mode (Coulter Corp., Miami, Fla.).

For ex vivo PT assays, citrated plasma was obtained from the treated rabbits as described above and clotting was started by adding thromboplastin. Clotting times were measured on Biodata-212 Microsampler Coagulation Analyzer (Horsham, P.A.).

For in vitro 2 stage prothrombin times (2st PT), increasing amounts of antibody AP-1 (0.01 µg/ml to 1000 µg/ml) were added to citrated rabbit plasma and rabbit tissue factor (Diagnostic Reagents, Thame Oxon, England) and incubated for 15 minutes at 37°C. After 15 minutes of incubation, clotting was initiated with calcium. Clotting times were measured on ACL300 using the PT mode (Coulter Corp., Miami, Fla.).

For ex vivo determinations, citrated plasma obtained from the treated animals was incubated with rabbit tissue factor. After 15 minutes, 50ul of 0.02M Ca<sup>++</sup> ions (Dade Behring, Deerfield, Ill.) was added and coagulation time measured on ACL300 using the PT mode (Coulter Corp., Miami, Fla.).

The activated partial thromboplastin times were also determined. Increasing amounts of antibody AP-1 were added to 53ul citrated rabbit plasma and 53 ul of activator (Actin FS) in the *in vitro* determination (Dade Behring, Deerfield, Ill.). Coagulation times were measured on ACL 300 using PT mode (Coulter Corp., Miami, Fla.).

For ex vivo determinations, 53 ul of citrated plasma from the treated rabbits were incubated with the activator. The mixture was incubated for 3 minutes at 37°C and then 53 ul of 0.02M of Ca<sup>++</sup> was added. Coagulation time was measured on Biodata-212 Microsampler Coagulation Analyzer (Horsham, PA).

#### Amidolytic Assay

Increasing amounts of antibody AP-1 or antibody 6B4 were incubated with 50nM soluble rabbit tissue factor and 10nM human FVIIa in HBS buffer containing 5mM CaCl<sub>2</sub> for 20 minutes at room temperature prior to the addition of Chromozym tPA. After 20 minutes incubation, Chromozym tPA was added. The rates of amidolytic activity were measured at 405 nm on a kinetic microplate reader (Molecular Devices). The background activity was defined as the amidolytic activity of FVIIa in the absence of sTF and was subtracted from the obtained values.

#### Cuticle Bleeding Time Assay

A standard cut was made at the apex of the nail cuticle by means of a nail clipper and blood was allowed to drop freely onto a gauze pad. The time that blood continued to flow freely from a transected cuticle was the cuticle bleeding time.

### Platelet Aggregation Assay

The effect of G4120 administration on ex-vivo platelet aggregation was determined as follows. Blood samples (2.7 ml/sample) collected in 3.8% Na<sub>3</sub>- citrate (9 parts blood to 1 part citrate) were processed to platelet rich (PRP) as previously described (Refino et al., Thromb Haemostas., 79:169-176 (1998)). Percent aggregation induced by 17  $\mu$ M ADP was determined for 200  $\mu$ l of each PRP sample in a PAP-4 Platelet Aggregation Profiler (Biodata Corp., Horsham PA). A sample of platelet poor rabbit plasma was used to calibrate the 100% aggregation setting. Data for each animal was expressed as a ratio of the mean aggregation of the post-treatment samples to the pre-treatment sample.

### Statistical Analysis

Data in table and graphs are represented as mean $\pm$ sem. Since thrombus mass appeared to have a non-normal distribution, a Mann-Whitney Test was used to test for differences between a given dose of a monotherapy or a combination treatment and saline control. This test was also used to test for differences in thrombus mass between combinations involving UFH and the relevant UFH monotherapy. For all other variables (CBT, ex-vivo coagulation tests and platelet aggregation), a Fisher's PLSD was used to test for differences between the various doses of a monotherapy and the saline control. A Dunnet's test was used to test for a significant effect (vs. saline) of combination therapies on CBT. The Fisher and Dunnet tests were used following a significant result ( $P \leq 0.05$ ) in a one-way ANOVA across the relevant treatment groups. A t-test (2-tail) was used to test for differences in CBT between combinations involving UFH and the relevant UFHF monotherapy.

### Effects of AP-1 on Coagulation Measured *In Vitro*

The effects of anti-tissue factor antibody on coagulation in vitro were measured in the prothrombin assay, the two stage prothrombin assay and activated partial thromboplastin assays. The results are shown in Figure 18. The effect of antibody AP-1 on amidolytic activity was also measured and shown in Figure 19.

As shown in Figure 18, AP-1 did not affect APTT and has only a small effect on the PT at concentrations above 100 ug/ml. AP-1 did increase coagulation times in the two-stage prothrombin time assay in a dose responsive manner. These results show that AP-1 did not affect the intrinsic pathway of coagulation and was a poor inhibitor of TF initiated

coagulation under non-equilibrium conditions. If AP-1 was preincubated with tissue factor before coagulation was initiated, as in the two-stage prothrombin assay, clotting time was potentially inhibited in a dose dependent manner ( 2 fold prolongation at 0.4ug/ml). These results suggest anti-TF antibody AP-1 inhibits the binding of factor VII/VIIa to tissue factor.

5 Amidolytic activity is an activity of the factor VII/VIIa tissue factor complex. The amidolytic activity of a complex of soluble rabbit tissue factor with human Factor VIIa was inhibited by AP-1 *in vitro*. The results are shown in Figure 19. The IC50 was 33nM or about 2 µg/ml. The 6B4 antibody was not effective to inhibit amidolytic activity in this assay because it is an antibody that binds human tissue factor and does not cross- react with rabbit  
10 tissue factor. Inhibition of amidolytic activity by AP-1 also supports the view that anti-TF antibody AP-1 interferes with the proper association of tissue factor with factor VII/VIIa.

#### Dose Response Experiments

15 As described previously, rabbits were treated with various doses of the anticoagulant and/or anti-platelet agents before flow restriction of the artery and 30 minutes before balloon damage. Thrombus mass was determined at 60 minutes after balloon procedure. The results are presented as a percentage of thrombus mass of the saline control (%C). Cuticle bleeding times were determined as described previously. The results are reported as a ratio of the post-cuticle bleeding time over the pre-cuticle bleeding time (post/pre). The results are shown in  
20 Figure 20. Blood samples taken from the treated rabbits were analyzed for *ex vivo* coagulation time and platelet aggregation. The results are shown in Table 4.

TABLE 4

Effects of AP-1 antibody, G4120 and heparin on ex-vivo coagulation and platelet aggregation in a rabbit thrombosis model. Data are mean  $\pm$  SEM

Treatment Dose (units)	N	APTT <sup>A</sup>	PT <sup>A</sup>	2sPT <sup>A</sup>	Platelet Aggregation <sup>A</sup>
<u>saline control</u>	24	1.07 $\pm$ 0.02	1.00 $\pm$ 0.00	1.05 $\pm$ 0.03	ND
<u>AP-1 (<math>\mu</math>g/kg)</u>					
500	5	1.18 $\pm$ 0.13	1.00 $\pm$ 0.01	3.33 $\pm$ 0.06	ND
1000	5	1.02 $\pm$ 0.01	0.98 $\pm$ 0.04	3.25 $\pm$ 0.12	ND
1500	5	1.21 $\pm$ 0.05	0.99 $\pm$ 0.00	3.67 $\pm$ 0.18	ND
<u>G4120 (B)</u>					
18.75 + 0.26	5	1.03 $\pm$ 0.02	1.01 $\pm$ 0.00	ND	0.92 $\pm$ 0.08
37.5 + 0.5	5	1.01 $\pm$ 0.02	0.99 $\pm$ 0.01	ND	0.56 $\pm$ 0.08
75 + 1.0	5	0.99 $\pm$ 0.01	1.00 $\pm$ 0.01	ND	0.39 $\pm$ 0.09
375 + 9.0	5	1.02 $\pm$ 0.04	1.01 $\pm$ 0.01	ND	0 $\pm$ 0
<u>Heparin (C)</u>					
25 + 0.25	11	1.67 $\pm$ 0.11	1.00 $\pm$ 0.00	1.01 $\pm$ 0.01	ND
50 + 0.5	8	2.92 $\pm$ 0.18	1.02 $\pm$ 0.01	1.08 $\pm$ 0.01	ND
100 + 1.0	7	8.97 $\pm$ 1.77	1.10 $\pm$ 0.02	1.22 $\pm$ 0.02	ND
150 + 1.5	5	18.6 $\pm$ 1.37	1.17 $\pm$ 0.03	1.26 $\pm$ 0.04	ND

A. data are expressed as the ratio of the post-treatment value (1 minutes post balloon procedure) over the pre-treatment value.

B. doses are bolus ( $\mu$ g/kg) followed by a continuous infusion ( $\mu$ g/kg/min)

C. doses are bolus (U/kg) followed by a continuous infusion (U/kg/min)

The results show that intravenous administration of anti-TF antibody AP-1 prior to vascular injury decreased thrombus mass in a dose dependent fashion. (Fig. 20, Panel A).

The dose that resulted in about 90% reduction or greater of thrombus mass (ED<sub>90</sub>) was a 1.5 mg/kg bolus of AP-1. The thrombus mass at that dose was less than 10% of the mass of the saline control. The ED<sub>90</sub> of AP-1 had no effect on coagulation times measured in ex vivo PT assay and APTT assay as shown in Table 4, but did prolong cuticle bleeding time to an extent similar to the ED<sub>90</sub> of unfractionated heparin (Fig. 20). Thus, the dose of AP-1 that was optimal to reduce thrombus mass also significantly increased cuticle bleeding time compared to saline control.

Increasing doses of AP-1 did increase coagulation times in the two stage prothrombin time assay measured *ex vivo* (Table 4). In the two stage assay, preincubation of anti-tissue factor antibody with tissue factor allows the antibody to bind to the tissue factor and inhibit the binding of factor VII and initiation of coagulation. As discussed previously with the *in vitro* assays, these results suggest that anti-TF antibody AP-1 inhibits the binding of factor VII to tissue factor.

Unfractionated heparin (UFH) was administered to the animals as a bolus followed by an infusion. Administration of UFH decreased thrombus mass in a dose dependent manner. (Figure 20, Panel C) The ED<sub>90</sub> dose was 100 units/kg bolus plus 1.0 unit/kg/min infusion. At the ED<sub>90</sub> dose, UFH significantly prolonged coagulation (Table 4) and cuticle bleeding times (Figure 20) compared to saline control. The optimal heparin dose increased the APTT about 8 fold and bleeding time was significantly prolonged. Suboptimal doses of heparin increased APTT less than about 3 fold and did not significantly prolong bleeding times compared to saline control.

Administration of G4120, GP IIa/IIIb inhibitor, also resulted in a dose dependent reduction of thrombus mass. (Figure 20, Panel B) Administration of G4120 at a dose of 375 µg/kg with an infusion 9.0 µg/kg/min reduced thrombus mass about 80%. None of the doses of G4120 tested were able to decrease thrombus mass 90% or greater. At the optimal dose of 375 µg/kg with an infusion 9.0 µg/kg/min and the next lower dose, cuticle bleeding times were significantly prolonged. (Figure 20, Panel B) Administration of G4120 did not affect coagulation times but did reduce platelet aggregation in a dose dependent manner as shown in Table 4.

Thus, each of the inhibitors reduced thrombus mass in a dose dependent fashion. Optimal doses of each of the inhibitors significantly prolonged cuticle bleeding time compared to saline control. Evidence from the 2 stage PT and amidolytic assays suggests that antibody AP-1 binds to tissue factor and interferes with the binding of Factor VII/VIIa.

#### Combination of Inhibitors

Although highly effective antithrombotic doses of each inhibitor were identified, a reduction of thrombus mass to about 80% or greater than that of the saline control was in each case associated with a significant increase in cuticle bleeding time. Therefore, additional studies were performed to determine if combinations of monotherapies at doses that did not decrease thrombus mass greater than about 80% and did not prolong cuticle bleeding times



were more efficacious than monotherapies. The monotherapies selected are shown in Table 5.

Table 5.

Effect of monotherapies on thrombus and bleeding. Data are mean $\pm$ sem

Treatment	N	Thrombus (%C)	% Reduction in thrombus mass	CBT post/pre
AP-1 (0.5 mg/kg)	5	60 $\pm$ 26	40%	1.13 $\pm$ 0.21
AP-1 (1.0 mg/kg)	5	77 $\pm$ 29	23%	1.30 $\pm$ 0.31
G4120 (37.5 $\mu$ g/kg + 0.5 $\mu$ g/kg/min)	5	75 $\pm$ 31	25%	1.19 $\pm$ 0.22
UFH (25 U/kg + 0.25 U/kg/min)	11	99 $\pm$ 14	1%	1.22 $\pm$ 0.19
UFH (50 U/kg + 0.5 U/kg/min)	8	33 $\pm$ 12	67%	1.39 $\pm$ 0.11

Combinations of inhibitors were administered to the rabbits before arterial damage was initiated as described previously. The results are shown in Table 6.

A suboptimal dose of AP-1 (0.5 mg/kg) was co-administered with two different suboptimal doses of unfractionated heparin. The amounts of heparin administered were 25 units/kg bolus and 0.25 units/kg/min infusion or 50 units/kg bolus plus 0.5 units/kg/min infusion. Suboptimal doses of AP-1 (0.5 or 1.0 mg/kg) were also co-administered with a suboptimal dose of 37.5  $\mu$ g/kg bolus plus 0.5  $\mu$ g/kg/min infusion of G4120.

Unfractionated heparin was also administered with G4120 as follows: G4120 was administered as 37.5  $\mu$ g/kg bolus plus 0.5  $\mu$ g/kg/min infusion with two different doses of UFH. UFH was administered as 25 units/kg bolus and 0.25 units/kg/min infusion or as 50 units/kg bolus plus 0.5 units/kg/min infusion.

Table 6.

Comparison of combination therapies to saline control (mean $\pm$ sem)

treatment (dose) $\dagger$ /treatment (dose) $\dagger$	n	Thrombus (%C)	CBT post/pre
Saline	24	100 $\pm$ 9.4	1.04 $\pm$ 0.04
AP-1(0.5)/UFH(25+0.25)	8	29.7 $\pm$ 12.3**	1.28 $\pm$ 0.10
AP-1(0.5)/UFH(50+0.5)	8	8.2 $\pm$ 5.4***	1.20 $\pm$ 0.10
AP-1(0.5)/G4120(37.5+0.5)	5	52.1 $\pm$ 34.7	1.13 $\pm$ 0.19
AP-1(1.0)/G4120(37.5+0.5)	8	27.1 $\pm$ 14.3**	1.35 $\pm$ 0.07 s
G4120(37.5+0.5)/UFH(25+0.25)	5	51.9 $\pm$ 36.6	1.35 $\pm$ 0.13
G4120(37.5+0.5)/UFH(50+0.5)	8	12.5 $\pm$ 8.2***	1.79 $\pm$ 0.11 s

$\dagger$  dose units are described in figure 20

\*  $P \leq 0.05$ , \*\*  $P \leq 0.01$ , \*\*\*  $P \leq 0.001$  vs. saline by Mann Whitney Test

s = significant difference vs saline by Mann-Whitney (thrombus) or Dunnell's Test (CBT)

The results show that the combination of AP-1 with unfractionated heparin resulted in a significant decrease in thrombus mass and did not significantly increase bleeding times at both doses of UFH when compared to the saline control. The combination of a 0.5 mg/kg bolus of AP-1 combined with a bolus 50 units/kg and infusion of 0.5 units/kg/min of unfractionated heparin reduced thrombus mass greater than 90% and did not significantly prolong bleeding time. When the combinations of anti-TF AP-1 and UFH are compared to the results with UFH alone, it can be seen that the combination reduced thrombus mass about 70%-75% more than UFH alone at both doses (Figure 21, Panel A) and did not significantly prolong bleeding times (Figure 21, Panel B). A bleeding time prolonged about 25% or less compared to the UFH alone was not significantly prolonged.

As shown in Table 6, the combination of AP-1 with G4120 also resulted in a significant decrease in thrombus mass and had a modest but significant effect on bleeding time at the higher dose of AP-1 when compared to the saline control. However, when compared to administration of G4120 alone, the combination of 1.0 mg/kg bolus of AP-1 with 37.5 µg/kg bolus and 0.5 ug/kg/min was effective to reduce thrombus mass about 64% and did not significantly prolong bleeding time.

The combination of 37.5 ug/kg bolus plus 0.5 ug/kg/min infusion of G4120 with the two different doses of UFH resulted in decrease in thrombus mass, but the most effective dose (UFH at 50 units/kg and infusion of 0.5 units/kg/min) significantly increased bleeding time when compared to the saline control. When the G4120 and heparin combinations were compared to administration of heparin alone as shown in Figure 21, it can be seen that the combination of G4120 with the higher dose of heparin was effective to decrease thrombus mass about 62% more than that dose of heparin alone. (Panel A) However, this dose of G4120 significantly increased the cuticle bleeding time (about 29%).

As discussed previously, anti-TF antibodies are effective anticoagulant agents. The antibodies described herein bind to epitopes that interfere with the association of TF and Factor VII/VIIa and/or TF and macromolecular substrates and inhibit the generation of thrombin. The results of amidolytic assay and the two stage prothrombin assay suggest that antibody AP-1 binds to an epitope that inhibits the interaction of Factor VII with tissue factor. This antibody is similar to antibodies 7G11, 6B4 and HPT1 described previously. Although these antibodies may be less potent than antibodies D3 and 5G6 under non-equilibrium conditions, antithrombotic efficacy of these antibodies can be enhanced when administered in combination with at least one additional anticoagulant and/or anti-platelet agent. These

results also suggest that the antithrombotic efficacy of treatments including anti-tissue factor antibodies that have enhanced anticoagulant potency, such as those that interfere with the interaction of tissue factor with macromolecular substrates, can also be enhanced by administering the antibody with at least one other anticoagulant or anti-platelet agent compared to administration of the antibody or agent alone.

The efficacy of the anti-tissue factor antibodies can be enhanced by co-administration of anti-TF antibody with UFH or G4120. The coadministration does not significantly increase bleeding time. The combination of an anti-TF antibody and unfractionated heparin has enhanced antithrombotic efficacy over unfractionated heparin alone. This combination provides for a greater decrease in thrombus mass and no significant increase in bleeding time as compared to the same dose of heparin alone. The combination of an anti-TF antibody and an anti-platelet agent also has enhanced antithrombotic efficacy over G4120 alone. This combination provides for a greater decrease in thrombus mass and no significant increase in bleeding time as compared to the same dose of G4120 alone. In contrast, the combination of UFH and G4120 results in a decrease in thrombus mass, but at the most effective dose significantly prolongs bleeding time compared to UFH alone. Combining an anti-tissue factor antibody with additional anticoagulant/ antiplatelet agents may be useful in the treatment of arterial thrombotic conditions, especially acute coronary syndrome.

All references cited throughout the specification, including the examples, and all references cited therein are hereby expressly incorporated by reference.

WHAT IS CLAIMED IS:

1. A method of enhancing the antithrombotic efficacy and minimizing an increase in  
5 bleeding risk of an anticoagulant comprising:  
administering a dose of anti-TF antibody and a suboptimal dose of at least  
one additional anticoagulant selected from the group consisting of heparin, low  
molecular weight heparin, pentasaccharide thrombin inhibitors and mixtures thereof.
- 10 2. A method according to claim 1, wherein the dose of the additional  
anticoagulant is a dose that does not prolong the activated partial prothrombin time (APTT)  
more than about 3 fold.
3. The method according to claim 1, wherein the anti-TF antibody binds to  
15 essentially the same epitope as an antibody selected from the group consisting of D3, 5G6 and  
T58-5G9.
4. The method of claim 1, wherein the anti-TF antibody binds to an epitope of  
human tissue factor that comprises amino acids K163, K166 and K201.
- 20 5. The method according to claim 1, wherein the anti-TF antibody binds to an  
epitope comprising at least part of the C-terminal macromolecular substrate-binding region of  
tissue factor.
- 25 6. The method according to claim 1, wherein the anti-TF antibody inhibits the  
interaction of tissue factor and factor VII/VIIa.
7. The method according to claim 6, wherein the antibody is selected from the  
group consisting of AP-1, 7G11, 6B4 and HPTI.
- 30 8. The method according to claim 1, wherein the anti-TF antibody is a humanized  
antibody comprising a light chain variable domain comprising hypervariable regions CDR-L1  
with a sequence of RASRDIKSYLN (SEQ ID NO: 10), CDR-L2 with the sequence  
YATSLAE (SEQ ID NO: 11), and CDRL3 with the sequence LQHGESPT (SEQ ID NO:  
35 9).

9. The method according to claim 1, wherein the anti-TF antibody is a humanized antibody comprising a heavy chain comprising hypervariable regions CDR-H1 with a sequence of GFWIKEYYMH (SEQ ID NO: 7), CDR-H2 with the sequence  
5 LIDPEQGNTIYDPKFQD (SEQ ID NO: 8) and CDR-H3 with the sequence DTAAYFDY (SEQ ID NO: 9).

10. The method according to claim 1, wherein the anti-TF antibody is a humanized antibody that binds to an epitope on human tissue factor comprising amino acids  
10 K165, K166 and K201.

11. A method according to claim 1, wherein the anti-TF antibody and the at least one additional anticoagulant agent are administered at the same time.

12. A method of treating a TF-VIIa related disease or disorder, comprising:  
administering to an individual a treatment comprising a dose of an anti-TF antibody and a dose of at least one additional anticoagulant agent selected from the group consisting of heparin, low molecular weight heparin, pentasaccharide thrombin inhibitors and mixtures thereof, wherein the treatment has enhanced antithrombotic efficacy and a minimal increase  
20 in bleeding risk compared to administration of the antibody or anticoagulant alone.

13. The method of claim 12, wherein the disease or disorder is a thrombotic or coagulopathic disorder.

14. The method of claim 13, wherein the disease or disorder is acute coronary syndrome.

15. The method of claim 14, wherein the disease or disorder is selected from the group consisting of unstable angina and non-ST-segment elevation myocardial infarction.

16. The method according to claim 12, wherein the anti-TF antibody binds to essentially the same epitope as an antibody selected from the group consisting of D3, 5G6 and T58-5G9.

17. The method of claim 12, wherein the anti-TF antibody binds to an epitope of human tissue factor that comprises amino acids K163, K166 and K201.

18. The method according to claim 12, wherein the anti-TF antibody binds to an epitope comprising at least part of the C-terminal macromolecular substrate-binding region of tissue factor.

19. The method according to claim 12, wherein the anti-TF antibody inhibits the interaction of tissue factor and factor VII/VIIa.

20. The method according to claim 19, wherein the antibody is selected from the group consisting of AP-1, 7G11, 6B4 and HPTI.

21. The method according to claim 12, wherein the anti-TF antibody is a humanized antibody comprising a light chain variable domain comprising hypervariable regions CDR-L1 with a sequence of RASRDIKSYLN (SEQ ID NO: 10), CDR-L2 with the sequence YATSLAE (SEQ ID NO: 11), and CDRL3 with the sequence LQHGESPW<sup>T</sup> (SEQ ID NO: 9).

22. The method according to claim 12, wherein the anti-TF antibody is a humanized antibody comprising a heavy chain comprising hypervariable regions CDR-H1 with a sequence of GFWIKEYYMH (SEQ ID NO: 7), CDR-H2 with the sequence LIDPEQGNTIYDPKFQD (SEQ ID NO: 8) and CDR-H3 with the sequence DTAAYFDY (SEQ ID NO: 9).

23. The method according to claim 12, wherein the anti-TF antibody is a humanized antibody that binds to an epitope on human tissue factor comprising amino acids K165, K166 and K201.

24. The method according to claim 12, wherein the dose of the anticoagulant agent is a dose that does not prolong the APTT more than about 3 fold.

25. The method according to claim 12, wherein the anti-TF antibody and at least one dose of the anticoagulant agent are administered serially.

26. A method according to claim 12, wherein the anti-TF antibody and at least one dose of anticoagulant agent are administered at the same time.

5 27. A method of treating a TF-VIIa related disease or disorder, comprising:  
administering to an individual a treatment comprising a dose of an anti-TF antibody and a dose of at least one antiplatelet agent, wherein the treatment has enhanced antithrombotic efficacy compared to administration of the antibody or agent alone.

10 28. A method according to claim 27, wherein the antiplatelet agent is selected from the group consisting of a cyclooxygenase inhibitor, ADP inhibitor, phosphodiesterase inhibitor and GP IIb/IIIa inhibitor.

15 29. A method according to claim 27, wherein the antiplatelet agent is acetylsalicylic acid.

30. A method according to claim 27, wherein the antiplatelet agent is a GP IIb/IIIa inhibitor.

20 31. The method of claim 27, wherein the disease or disorder is selected from the group consisting of unstable angina and non-ST-segment elevation myocardial infarction.

25 32. The method of claim 27, wherein the treatment has a minimal increase in bleeding risk compared to the antibody or agent alone.

33. The method according to claim 27, wherein the anti-TF antibody binds to an epitope comprising at least part of the C-terminal macromolecular substrate-binding region of tissue factor.

30 34. The method according to claim 27, wherein the anti-TF antibody inhibits the interaction of tissue factor and factor VII/VIIa.

35. The method according to claim 34, wherein the antibody is selected from the group consisting of AP-1, 7G11, 6B4 and HPTI.

36. The method according to claim 27, wherein the anti-TF antibody is a humanized antibody comprising a light chain variable domain comprising hypervariable regions CDR-L1 with a sequence of RASRDIKSYLN (SEQ ID NO: 10), CDR-L2 with the sequence YATSLAE (SEQ ID NO: 11), and CDRL3 with the sequence LQHGESPW (SEQ ID NO: 9).

37. The method according to claim 27, wherein the anti-TF antibody is a humanized antibody comprising a heavy chain comprising hypervariable regions CDR-H1 with a sequence of GFWIKEYYM (SEQ ID NO: 7), CDR-H2 with the sequence LIDPEQGNTIYDPKFQD (SEQ ID NO: 8) and CDR-H3 with the sequence DTAAYFDY (SEQ ID NO: 9).

38. The method according to claim 27, wherein the anti-TF antibody is a humanized antibody that binds to an epitope on human tissue factor comprising amino acids K165, K166 and K201.

39. A method according to claim 27, wherein the anti-TF antibody and antiplatelet agent are administered at the same time.

40. A method of treating a TF-VIIa related disease or disorder, comprising: administering to an individual a treatment comprising a dose of an anti-TF antibody and a dose of at least one additional anticoagulant agent or antiplatelet agent, wherein the antibody binds to an epitope comprising at least part of the C-terminal macromolecular substrate-binding region of tissue factor.

41. The method of claim 40, wherein the disease or disorder is a thrombotic or coagulopathic disorder.

42. The method of claim 40, wherein the disease or disorder is acute coronary syndrome.

43. The method of claim 40, wherein the disease or disorder is selected from the group consisting of unstable angina and non-ST-segment elevation myocardial infarction.



44. The method according to claim 40, wherein the anti-TF antibody binds to essentially the same epitope as an antibody selected from the group consisting of D3, 5G6 and T58-5G9.

45. The method of claim 40, wherein the anti-TF antibody binds to an epitope of human tissue factor that comprises amino acids K163, K166 and K201.

46. The method according to claim 40, wherein the anti-TF antibody is a humanized antibody comprising a light chain variable domain comprising hypervariable regions CDR-L1 with a sequence of RASRDIKSYLN (SEQ ID NO: 10), CDR-L2 with the sequence YATSLAE (SEQ ID NO: 11), and CDRL3 with the sequence LQHGESPW (SEQ ID NO: 9).

47. The method according to claim 40, wherein the anti-TF antibody is a humanized antibody comprising a heavy chain comprising hypervariable regions CDR-H1 with a sequence of GFWIKEYYM (SEQ ID NO: 7), CDR-H2 with the sequence LIDPEQGNTIYDPKFQD (SEQ ID NO: 8) and CDR-H3 with the sequence DTAAYFDY (SEQ ID NO: 9).

48. The method according to claim 40, wherein the anti-TF antibody is a humanized antibody that binds to an epitope on human tissue factor comprising amino acids K165, K166 and K201.

49. The method according to claim 40, wherein the anticoagulant agent is administered at a dose that does not prolong APTT more than about 3 fold..

50. A method according to claim 40, wherein the anti-TF antibody and at least one dose of anticoagulant agent or anti-platelet agent are administered at the same time.

51. A method according to claim 40, wherein the anticoagulant is an agent that inhibits formation or growth of a thrombus.

52. A method according to claim 51, wherein the anticoagulant is selected from the group consisting of heparin, low molecular weight heparin and direct thrombin inhibitors.

53. A method according to claim 52, wherein the anticoagulant is heparin or low molecular weight heparin.

54. A method according to claim 40, wherein the antiplatelet agent is selected from the group consisting of a cyclooxygenase inhibitor, ADP inhibitor, phosphodiesterase inhibitor and IIb/IIIa inhibitor.

55. A method according to claim 54, wherein the antiplatelet agent is acetyl salicylic acid.

56. A method according to claim 54, wherein the antiplatelet agent is a GP IIb/IIIa inhibitor.

57. The method of claim 40, wherein the treatment has enhanced antithrombotic efficacy and a minimal increase in bleeding risk compared to administration of the antibody or agent alone.

58. A composition comprising:

a) an anti-TF antibody; and

b) at least one additional anticoagulant or anti-platelet agent, wherein the anticoagulant agent inhibits the formation or growth of a thrombus;

in admixture with a pharmaceutically acceptable carrier.

59. The composition of claim 58, wherein the anti-TF antibody binds to an epitope on human tissue factor and inhibits the interaction of tissue factor and factor VII.

60. The composition of claim 58, wherein the anti-TF antibody binds to an epitope comprising at least part of the C-terminal macromolecular substrate binding region with tissue factor.

61. The composition of claim 58, wherein said region includes residues interacting with a Gla domain of said F.X or F.IX.

62. The composition of claim 58, wherein said epitope comprises residues K165, K166 and K201 of hTF.

5 63. The composition of claim 58, wherein said epitope additionally comprises residues N199, R200 and I152 of hTF.

64. The composition of claim 58, wherein said epitope additionally comprises residue Y156 of hTF.

10 65. The composition of claim 58, wherein the antibody binds essentially to the same hTF epitope as an antibody selected from the group consisting of D3, 5G6, and TF8-5G9.

15 66. The composition of claim 58, wherein the antibody binds essentially to the same epitope as 7G11, 6B4 and HPTI.

67. The composition of claim 58, wherein the antibody binds essentially to the same hTF epitope as antibody D3.

20 68. The composition of claim 58, wherein the antibody does not interfere with the hTF-Factor VIIa (FVIIa) association.

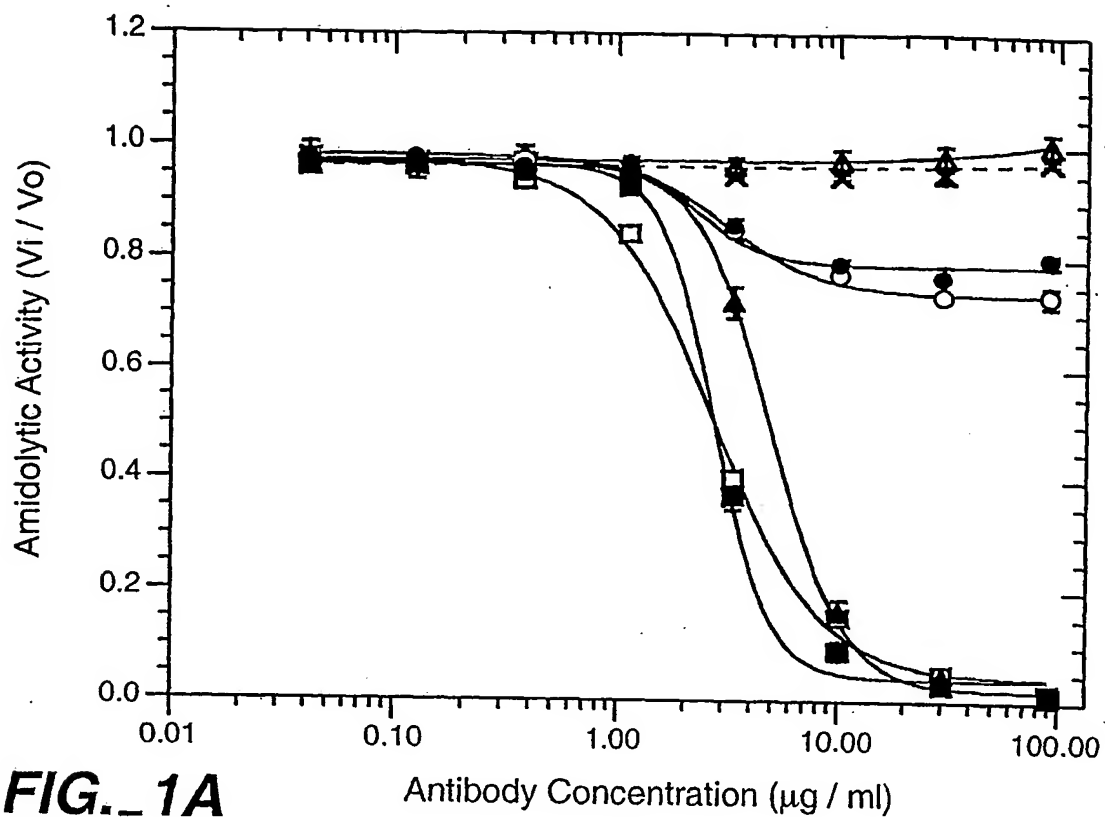
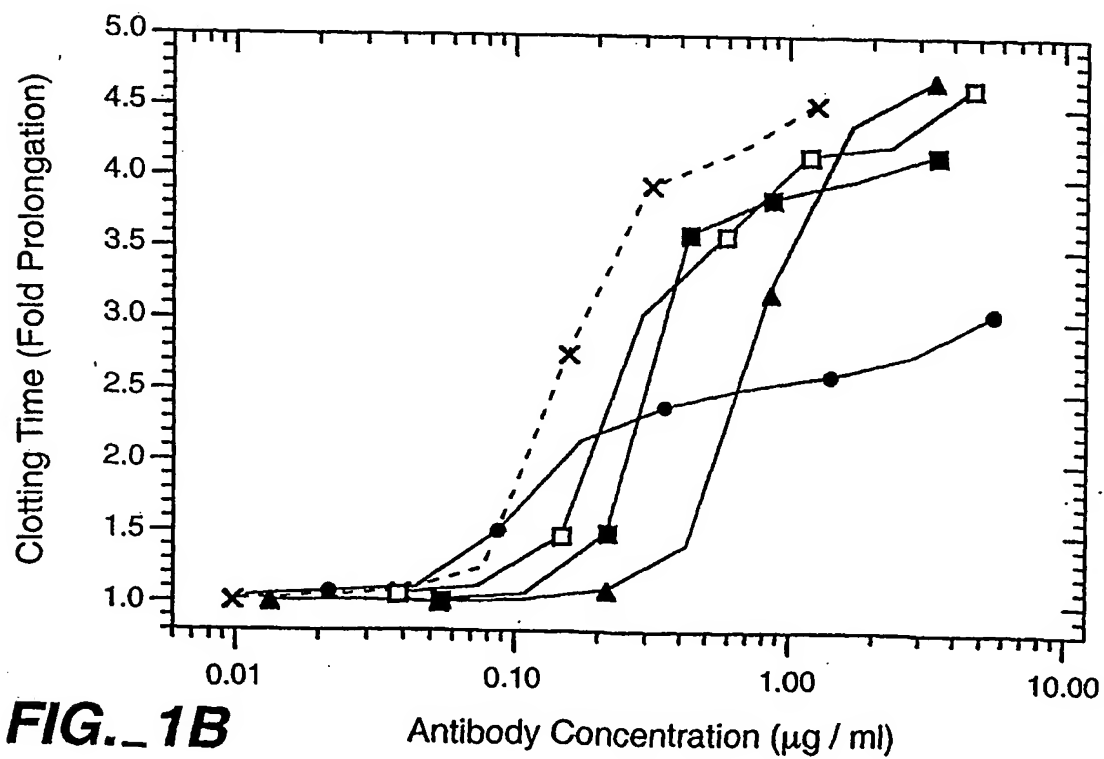
69. The composition of claim 58, wherein said antibodies are humanized.

25 70. The composition of claim 58, wherein said antibodies are human.

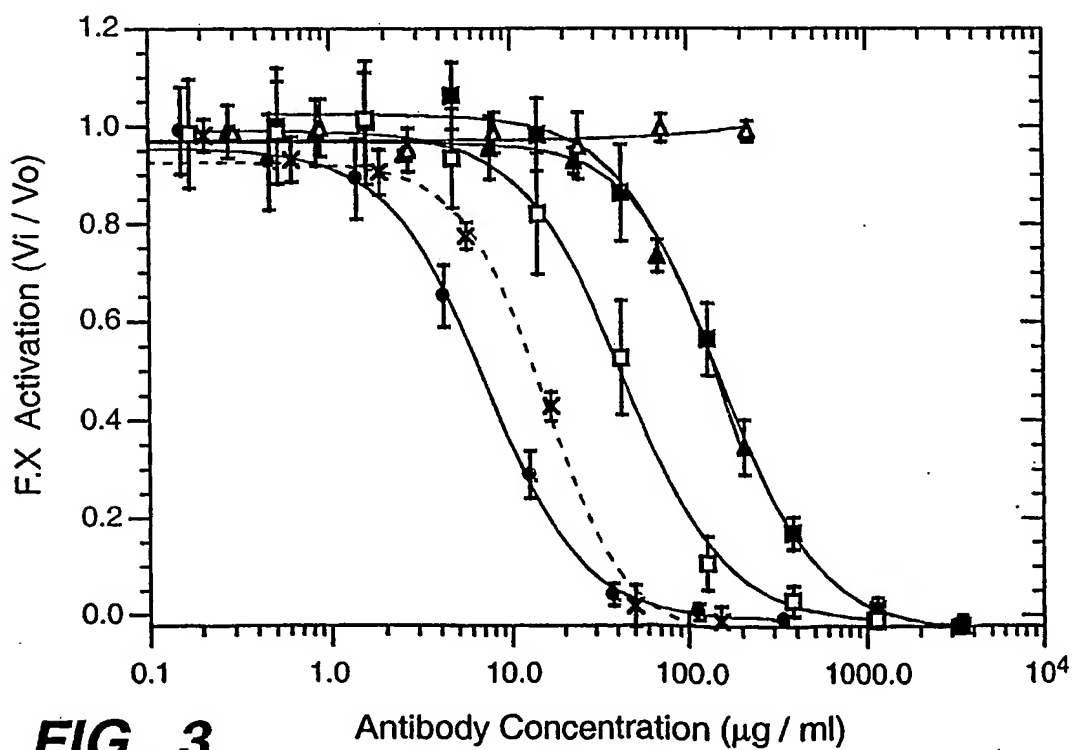
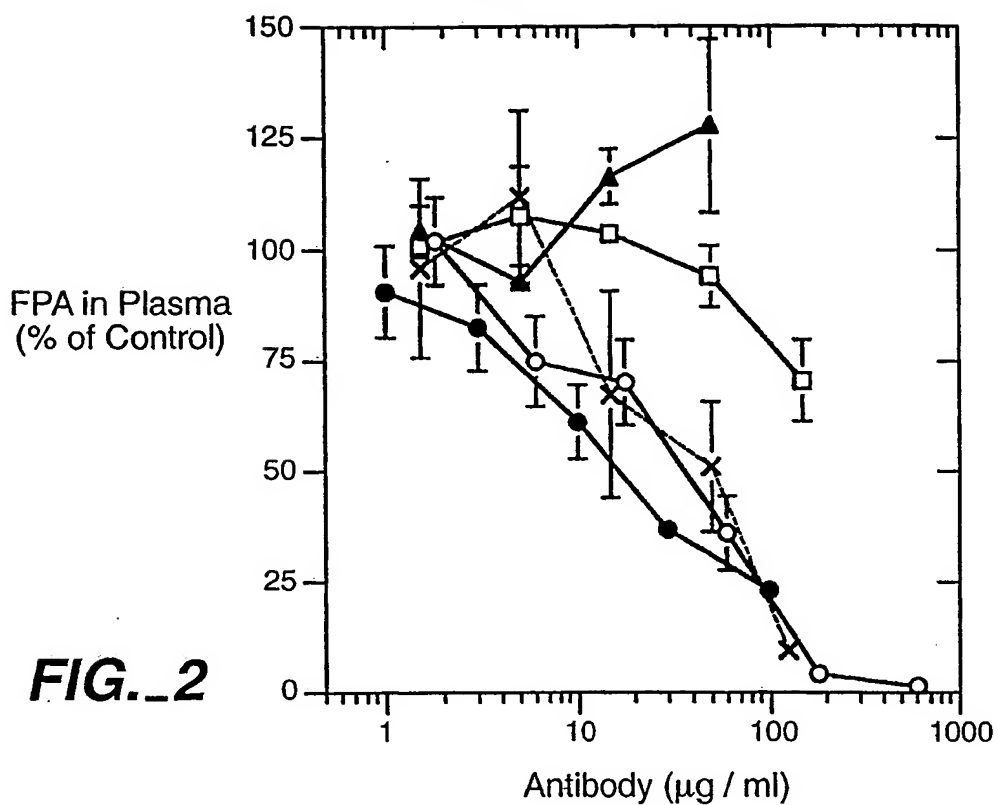
71. The composition of claim 58, wherein the anti-platelet agent is a platelet glycoprotein GP IIb/IIIa inhibitor.

30 72. The composition of claim 58, wherein the anticoagulant is selected from the group consisting of heparin, low molecular weight heparin, and direct thrombin inhibitors.

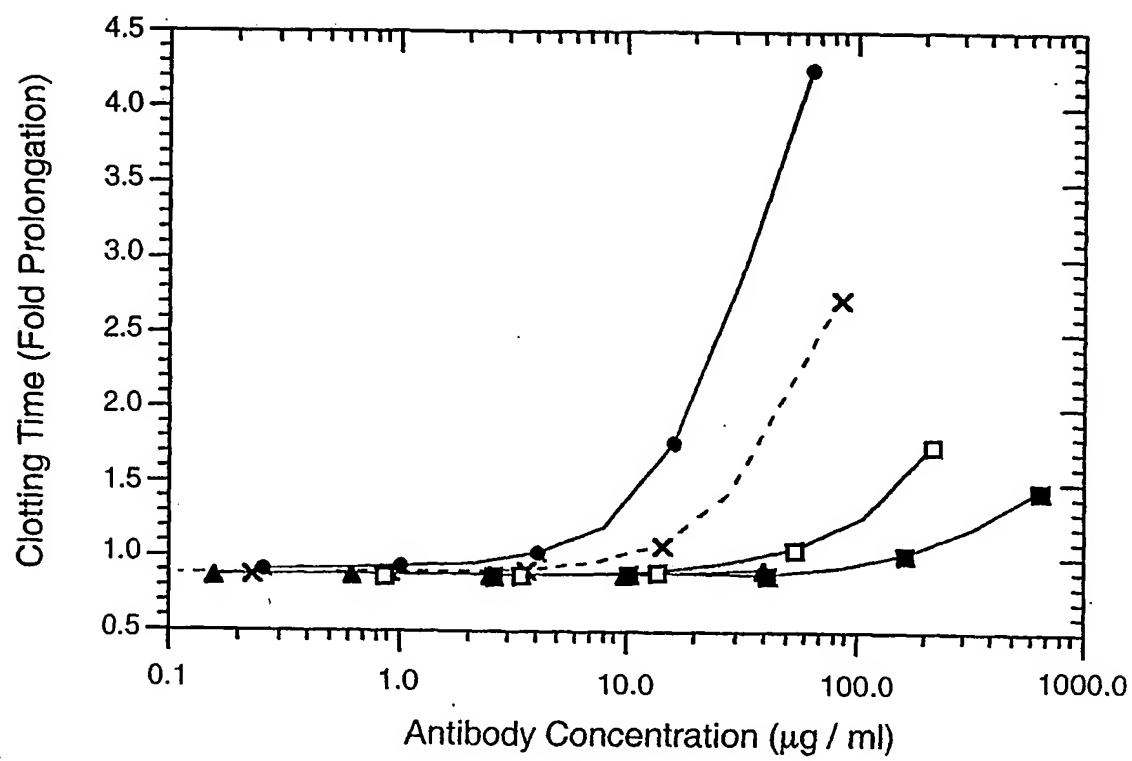
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**FIG. 1A****FIG. 1B**

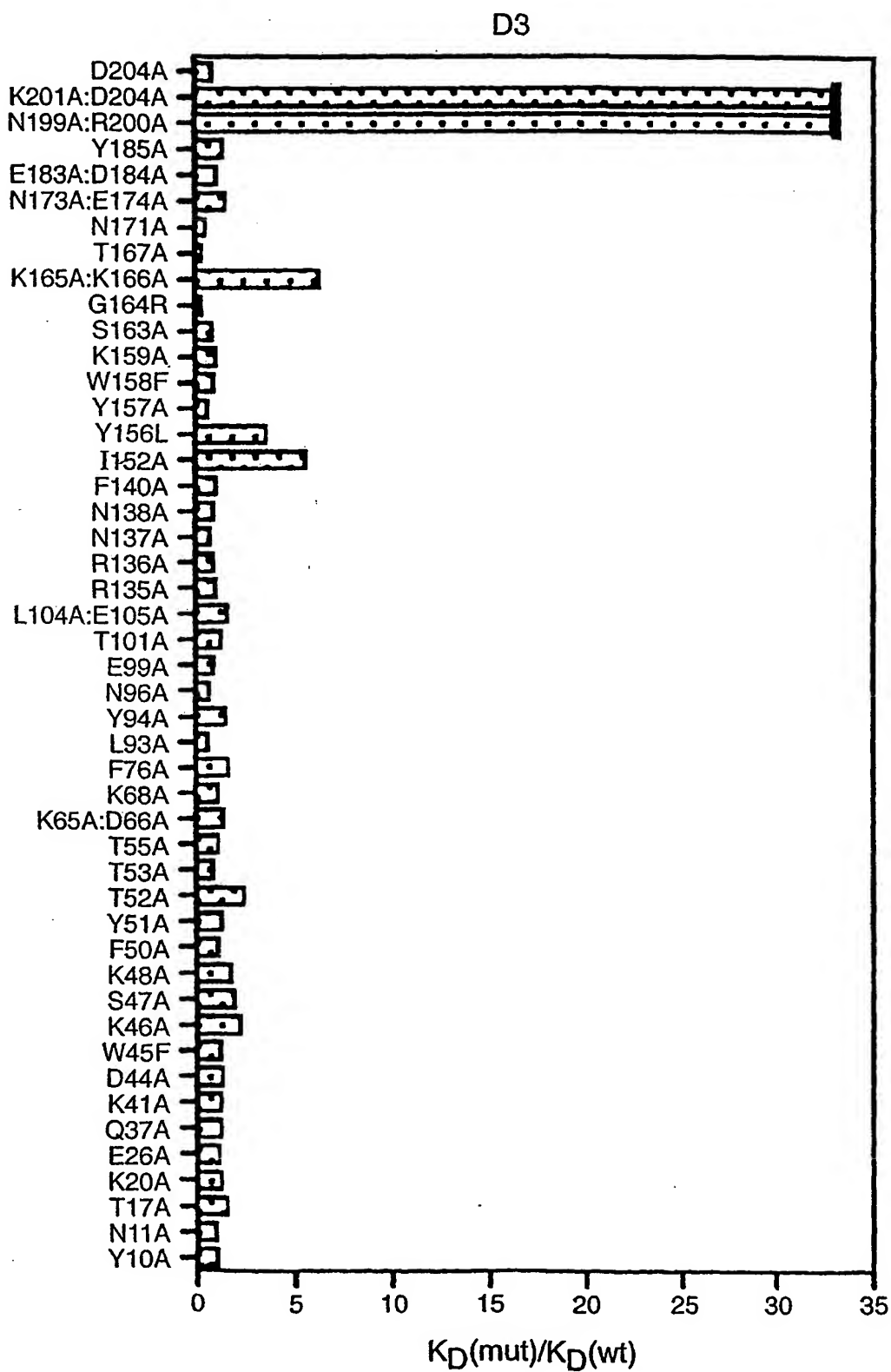
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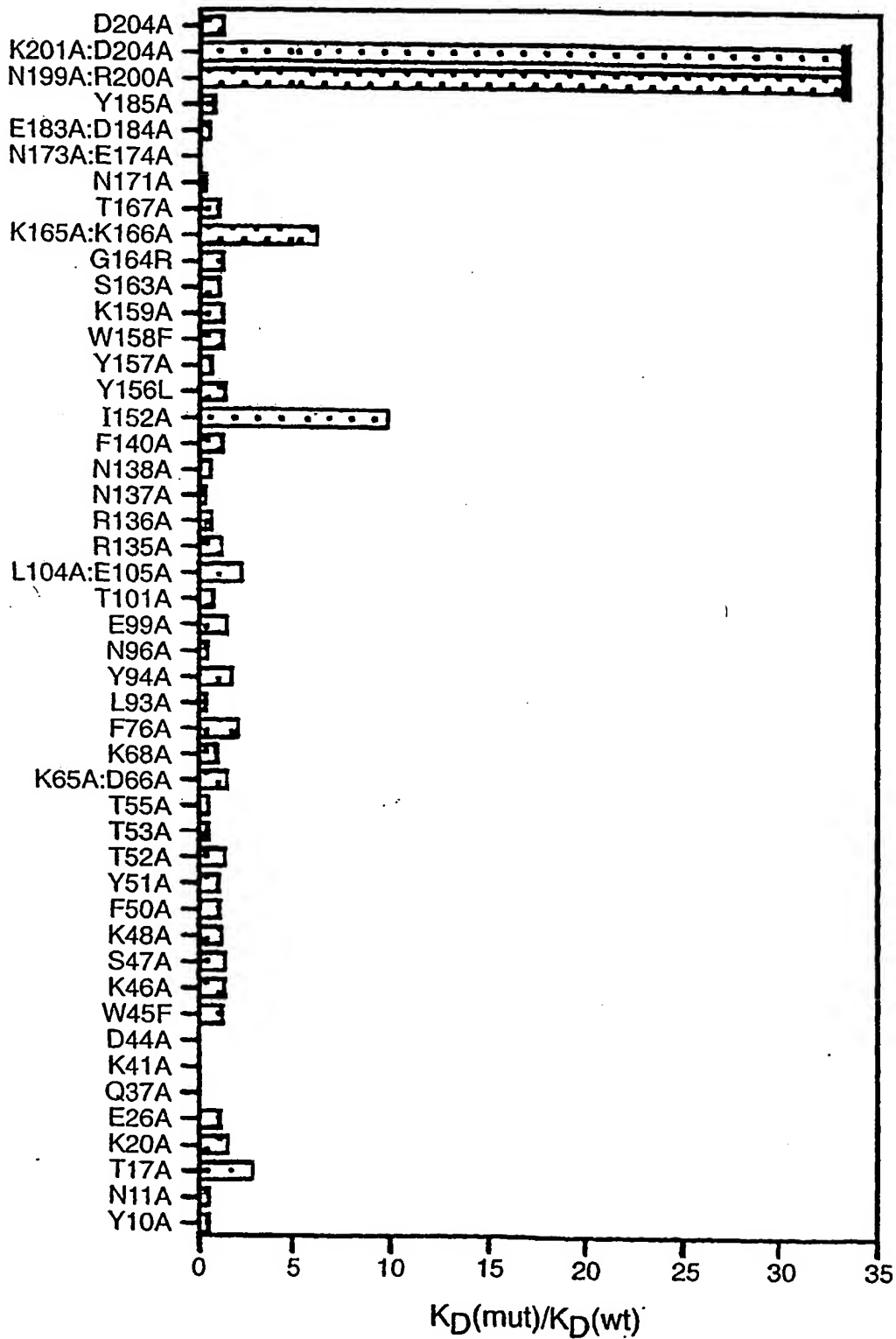
**FIG.\_4**

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**FIG. 5A**

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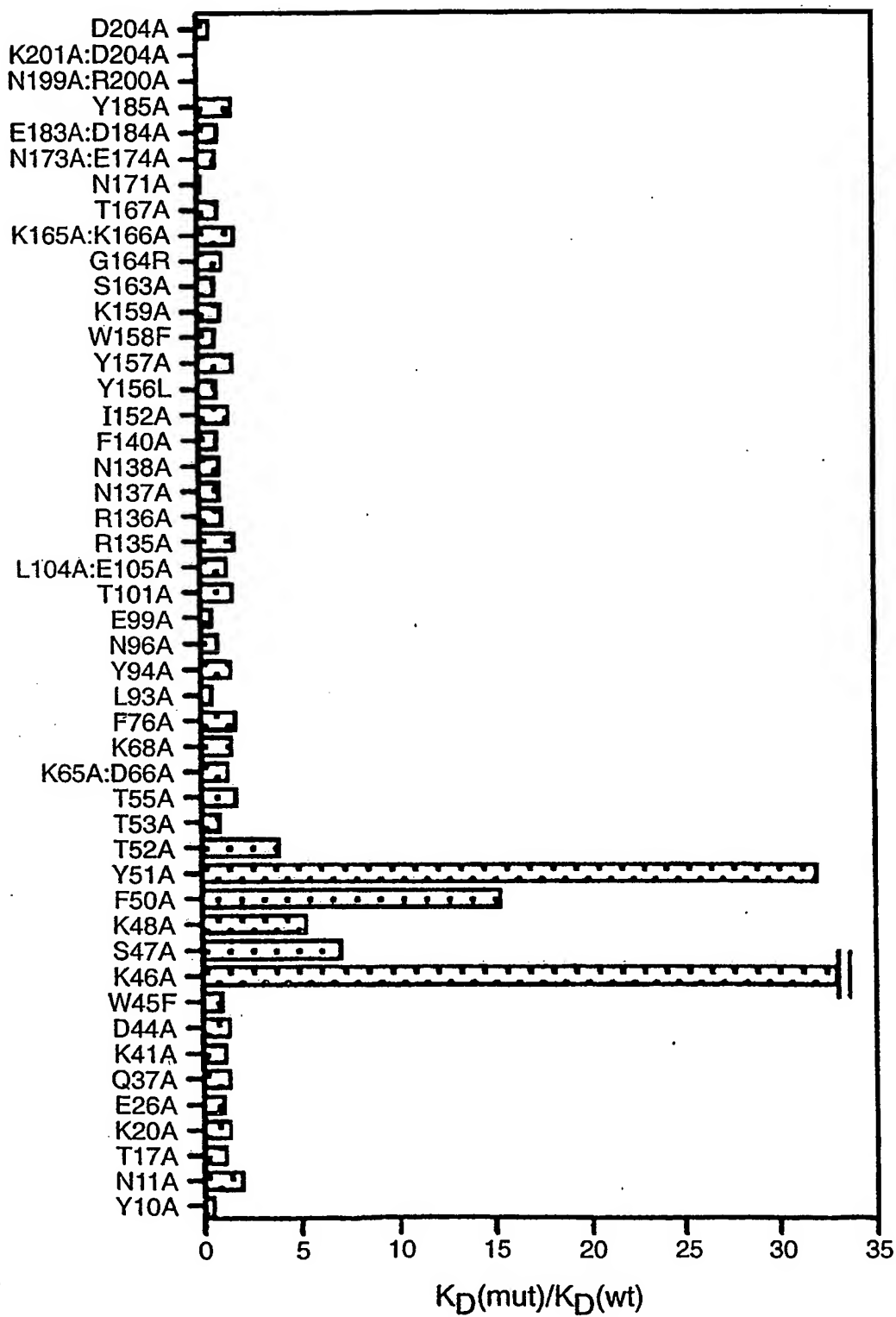
5G6

**FIG. 5B**



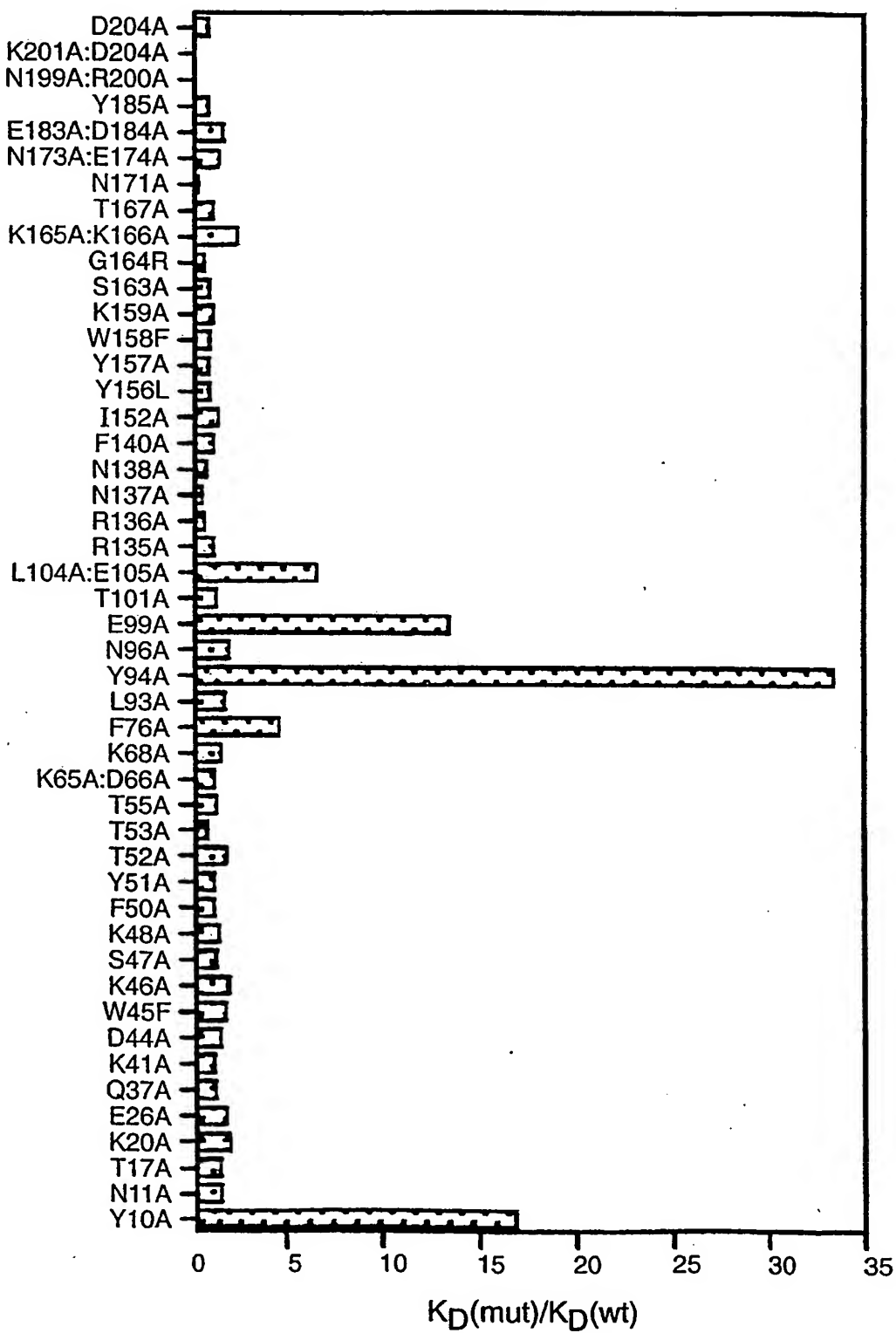
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7G11

**FIG. 5C**

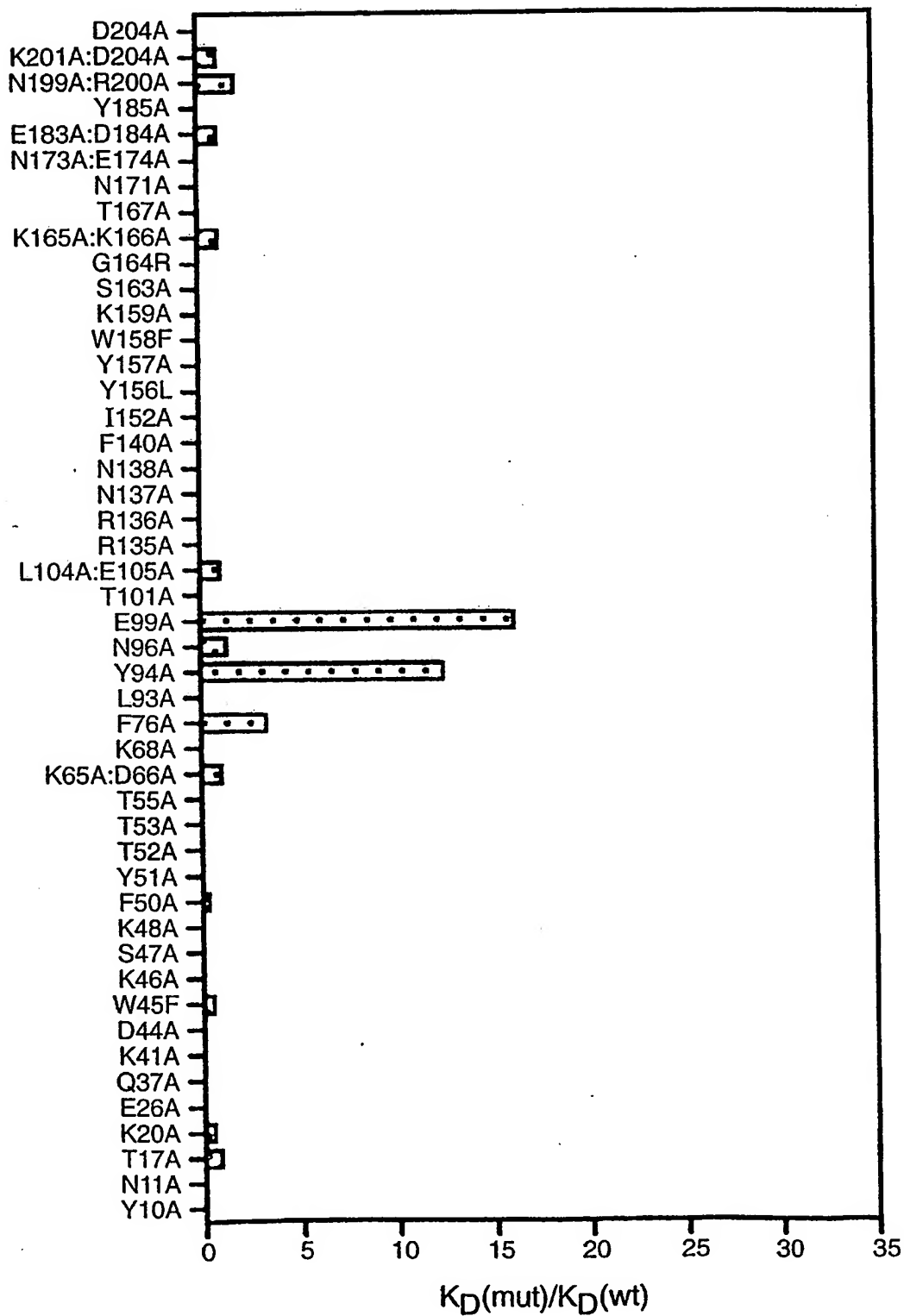
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6B4

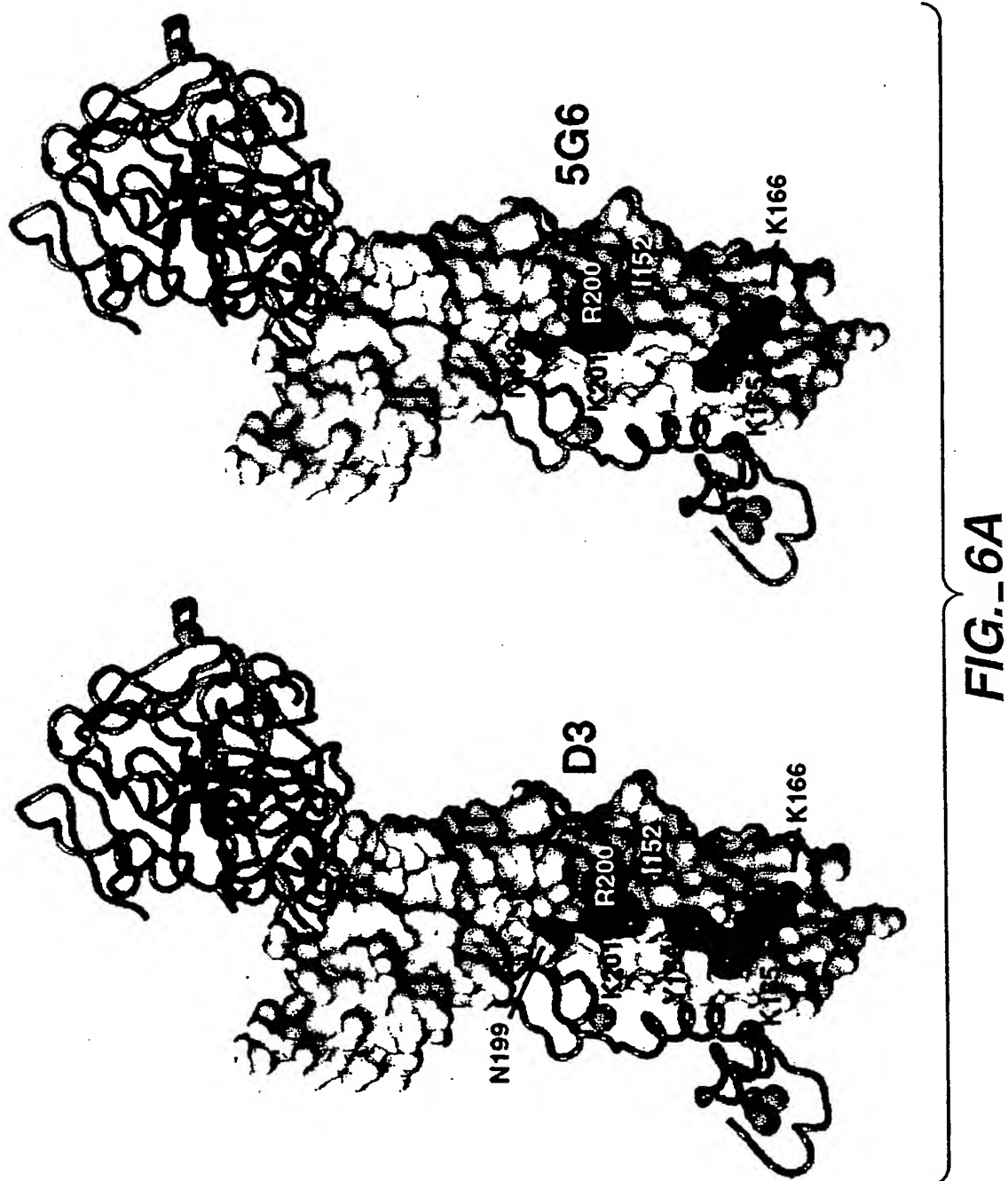
**FIG. 5D**

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HTF1

**FIG. 5E**

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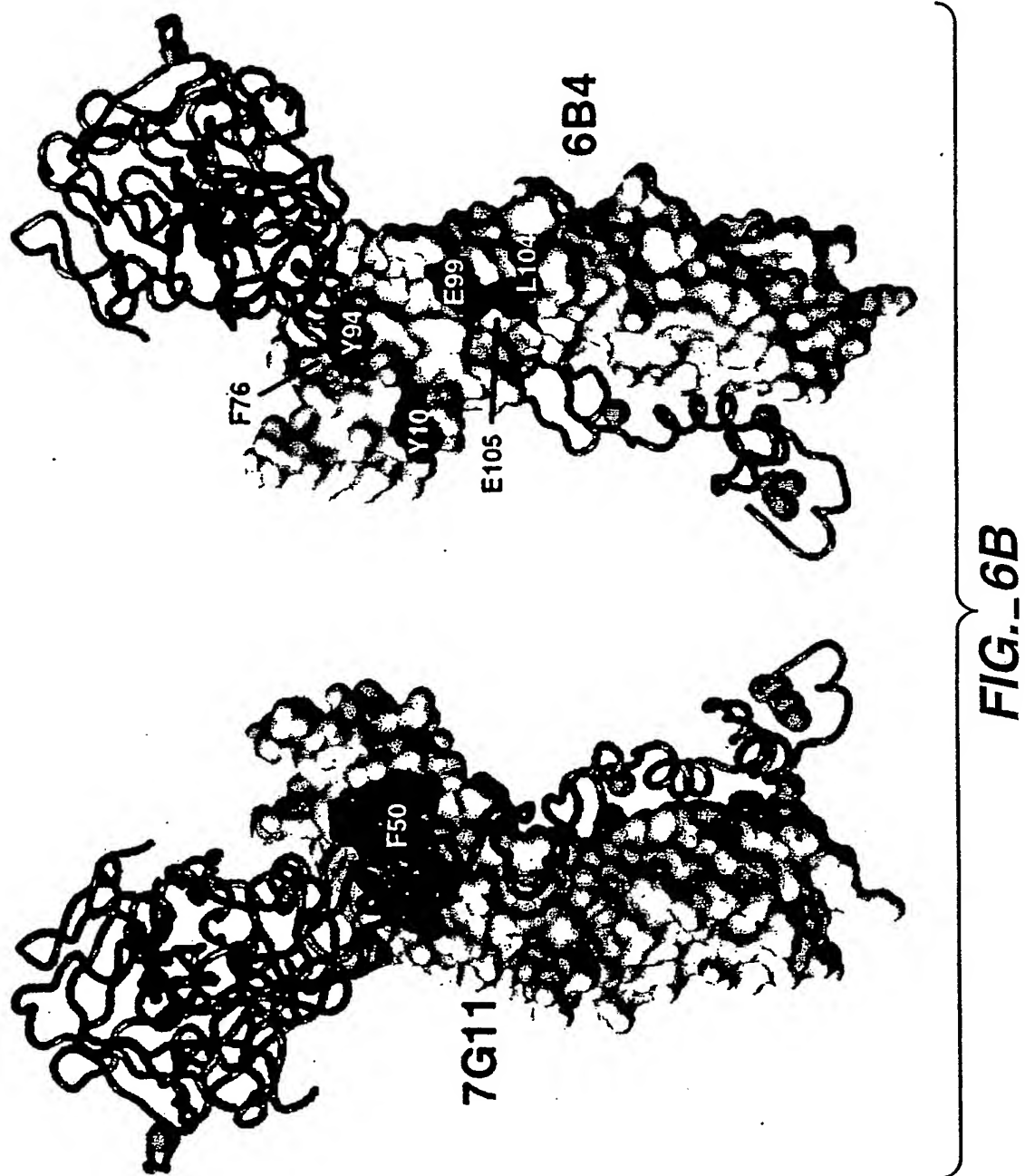




FIG. 7

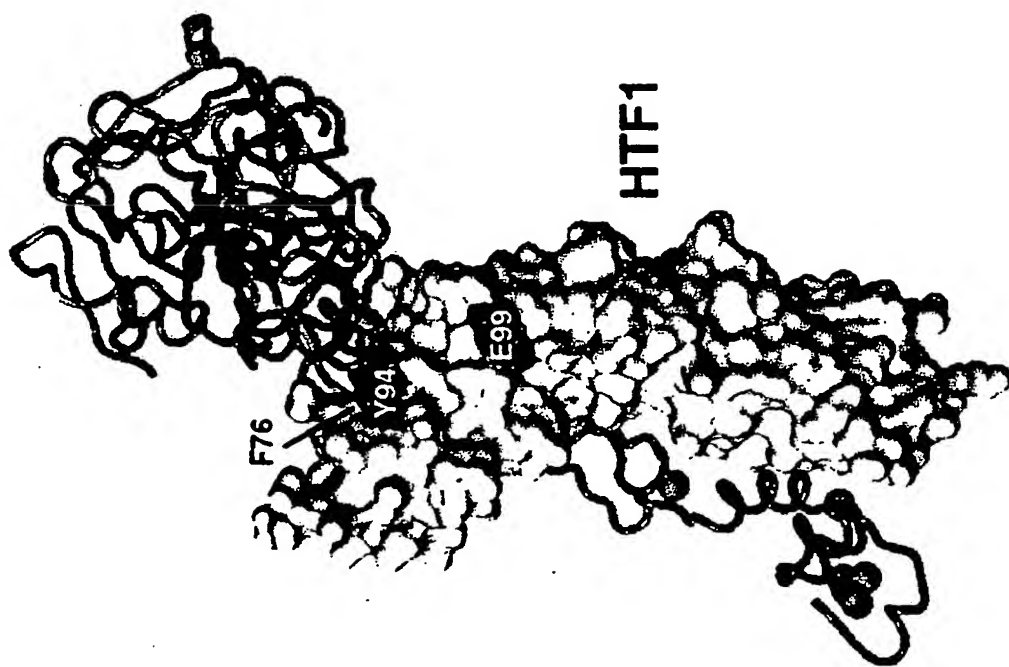


FIG. 6C

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## Variable Heavy

	10	20	30	40
D3Mur	EVQLQQSGAELVRPGALVKLSCKASGFN	IKDYIMHWVKQRPEQGLELIG		
	** ** *	**** *	*	* * * *
D3H44	EVQLVESGGGLVQPGGSLRLSCAASGFNI	KEYIMHWVRQAPGKGLEWVG		
		**** * *		*
HumVHIII	EVQLVESGGGLVQPGGSLRLSCAASGFTF	SSYAMSWVRQAPGKGLEWVA		
		-----		
		CDR-H1		

	50	a	60	70	80	abc	90
D3Mur	WIDPENGNTIYDPKFQDKASITADTSSNT	AYLQLSSLTSED	TAVYYCAR				
	*	*		* * * *		** **	
D3H44	LIDPEQGNTIYDPKFQDRATISADNSKNT	AYLQMN	SLRAEDTAVYYCAR				
	* **** *	* * * * *	*	*			
HumVHIII	VISGDGGSTYYADSVKGRFTISRDN	SKNTLYLQMN	SLRAEDTAVYYCAR				
		-----					
		CDR-H2					

	100	110	
D3Mur	DTAAYFDYWGQGTTLVSS		(SEQ ID NO: 1)
	**		
D3H44	DTAAYFDYWGQGTTLTVSS		(SEQ ID NO: 2)
	*****		
HumVHIII	GRGGGSDYWGQGTTLTVSS		
	-----		
	CDR-H3		

**FIG. 8**

## Variable Light

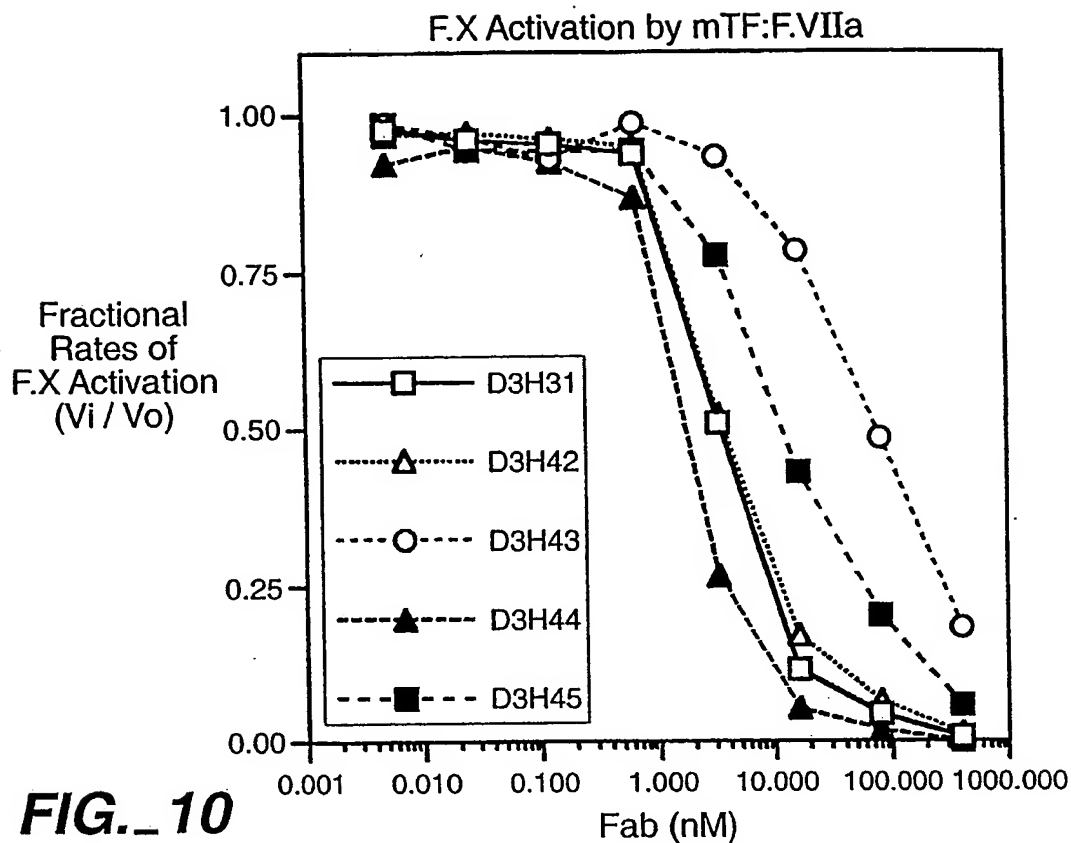
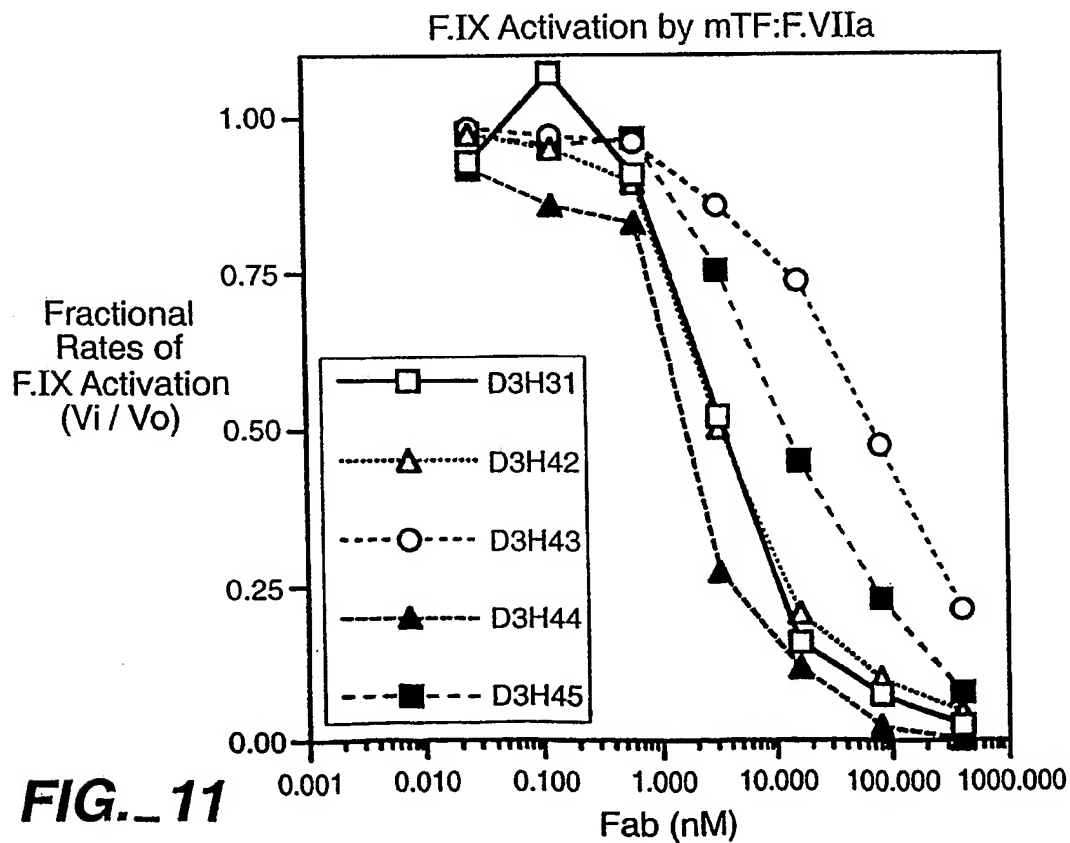
	10	20	30	40
D3Mur	DIKMTQSPSSMSASLGESVTITCKASRDIKS	YLSWYQQKPKWKSPKTLIY		
	*	* * *	*	* * *
D3H44	DIQMTQSPSSLSASVGDRVTITCRASRDIKS	YLNWYQQKPKGKAPKVLIIY		
		**** *		*
HumkI	DIQMTQSPSSLSASVGDRVTITCRAS-QISN	YLAWYQQKPKGKAPKLLIY		
		-----		
		CDR-L1		

	50	60	70	80	90
D3Mur	YATSLADGVPSRFSGSGSGQDYSLTISSLES	DDTATYYCLQHGES	PFTFG		
	*	*	*	*	*
D3H44	YATSLAEGVPSRFSGSGSGTDYTLTISSLQ	PEDFATYYCLQHGES	PWTFG		
	* * *	*		* * * *	
HumkI	AASSLESGVPSRFSGSGSGTDFTLTIS	SLQPEDFATYYCQYNSLPWTFG			
		-----		-----	
		CDR-L2		CDR-L3	

	100	
D3Mur	SGTKLELKRT	(SEQ ID NO: 3)
	* * *	
D3H44	QGTKVEIKRT	(SEQ ID NO: 4)
HumkI	QGTKVEIKRT	

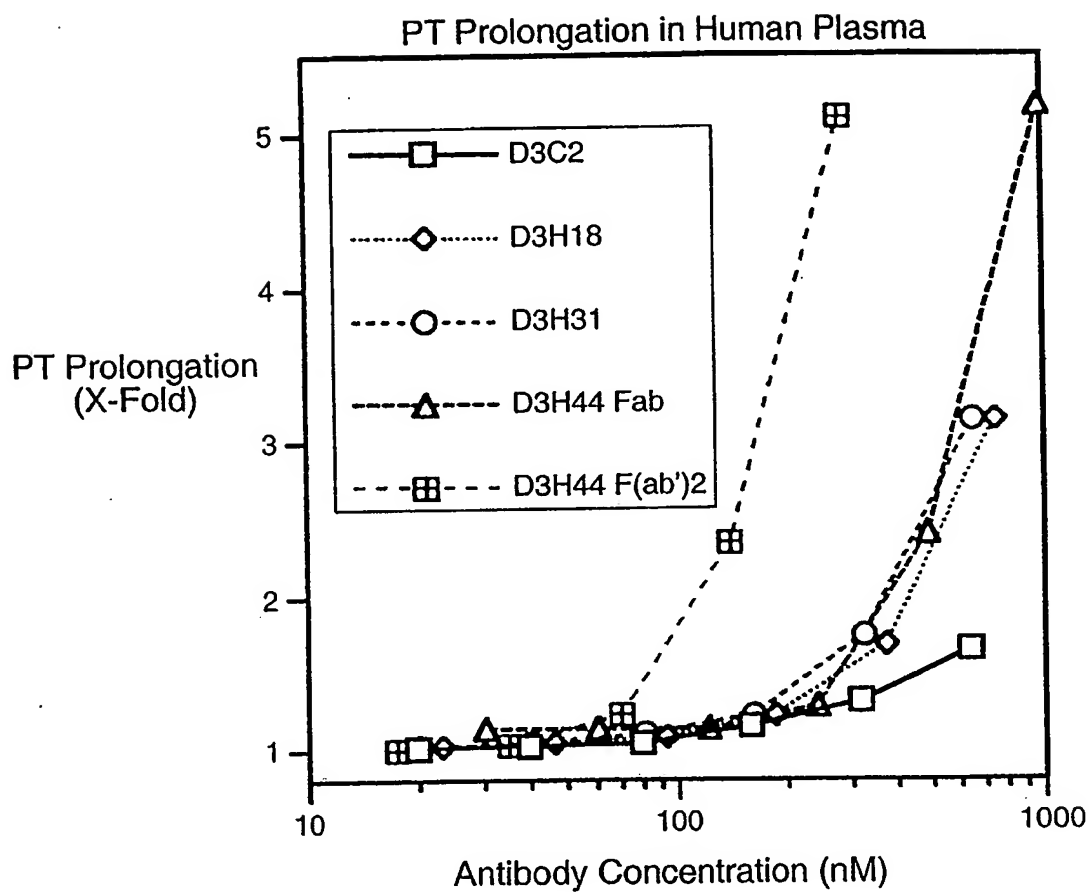
**FIG. 9**

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**FIG. 10****FIG. 11**



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**FIG. 12**

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Ser Gly Thr Thr Asn Thr Val Ala Ala Tyr Asn Leu Thr Trp Lys Ser Thr Asn Phe Lys Thr Thr Ile	22
Leu Glu Trp Glu Pro Lys Pro Val Asn Gln Val Tyr Thr Val Gln Ile Ser Thr Lys Ser Gly Asp	44
Trp Lys Ser Lys Cys Phe Tyr Thr Thr Asp Thr Glu Cys Asp Leu Thr Asp Glu Ile Val Lys Asp	66
Val Lys Gln Thr Tyr Leu Ala Arg Val Phe Ser Tyr Pro Ala Gly Asn Val Glu Ser Thr Gly Ser	88
Ala Gly Glu Pro Leu Tyr Glu Asn Ser Pro Glu Phe Thr Pro Tyr Leu Glu Thr Asn Leu Gly Gln	110
Pro Thr Ile Gln Ser Phe Glu Gln Val Gly Thr Lys Val Asn Val Thr Val Glu Asp Glu Arg Thr	132
Leu Val Arg Arg Asn Asn Thr Phe Leu Ser Leu Arg Asp Val Phe Gly Lys Asp Leu Ile Tyr Thr	154
Leu Tyr Tyr Trp Lys Ser Ser Ser Ser Gly Lys Lys Thr Ala Lys Thr Asn Thr Asn Glu Phe Leu	176
Ile Asp Val Asp Lys Gly Glu Asn Tyr Cys Phe Ser Val Gln Ala Val Ile Pro Ser Arg Thr Val	198
Asn Arg Lys Ser Thr Asp Ser Pro Val Glu Cys Met Gly Gln Glu Lys Gly Glu Phe Arg Glu Ile	220
Phe Tyr Ile Ile Gly Ala Val Phe Val Val Ile Ile Leu Val Ile Ile Leu Ala Ile Ser Leu	242
His Lys Cys Arg Lys Ala Gly Val Gly Gln Ser Trp Lys Glu Asn Ser Pro Leu Asn Val Ser	263

**FIG.-13**

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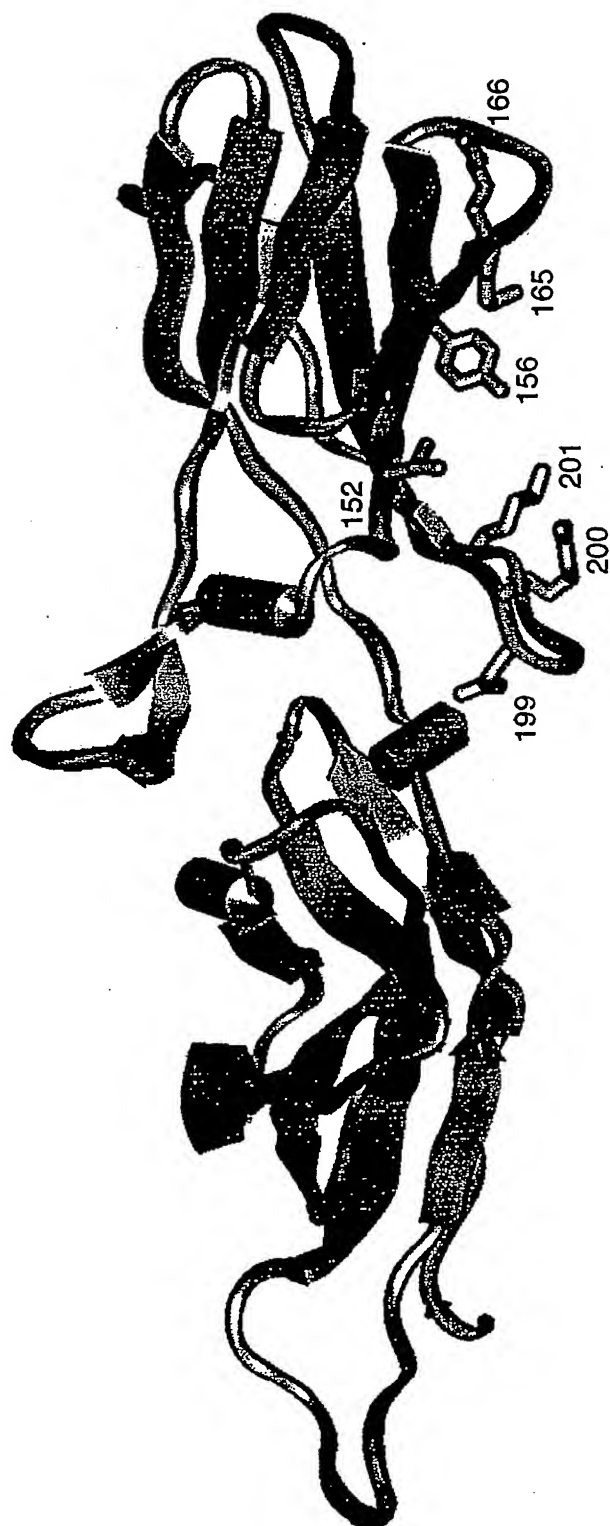


FIG. 14

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## 5G6 Heavy Chain

10 20 30 40  
5G6 VH EVLLQQSGPELVKPGASVKIPCKASGYTFTEYNMDWVKQSEHGSLEWIG

-----  
CDR-H1

50 a 60 70 80 abc 90  
5G6 VH DINPNNNGNTIYNQKFKGKATLTVDKSSTTAYLELRSLTSEDVAVYFCAR

-----  
CDR-H2

100 110  
5G6 VH DHDYYFDFWGQGTTTLTVSSA (SEQ ID NO: 5)

-----  
CDR-H3

## 5G6 Light Chain

10 20 30 40  
5G6 VL DIQMTQTPASQSASLGESVTITCLASQTIDTWLAWYQQKPGKSPQLLIY

-----  
CDR-L1

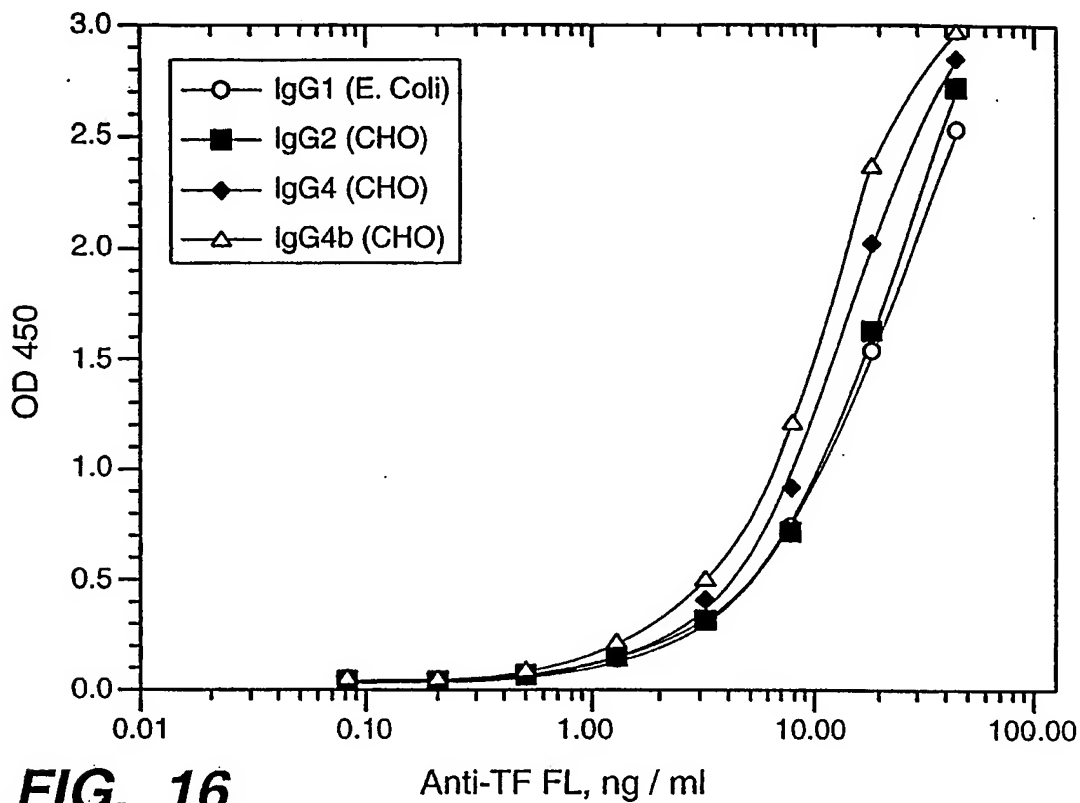
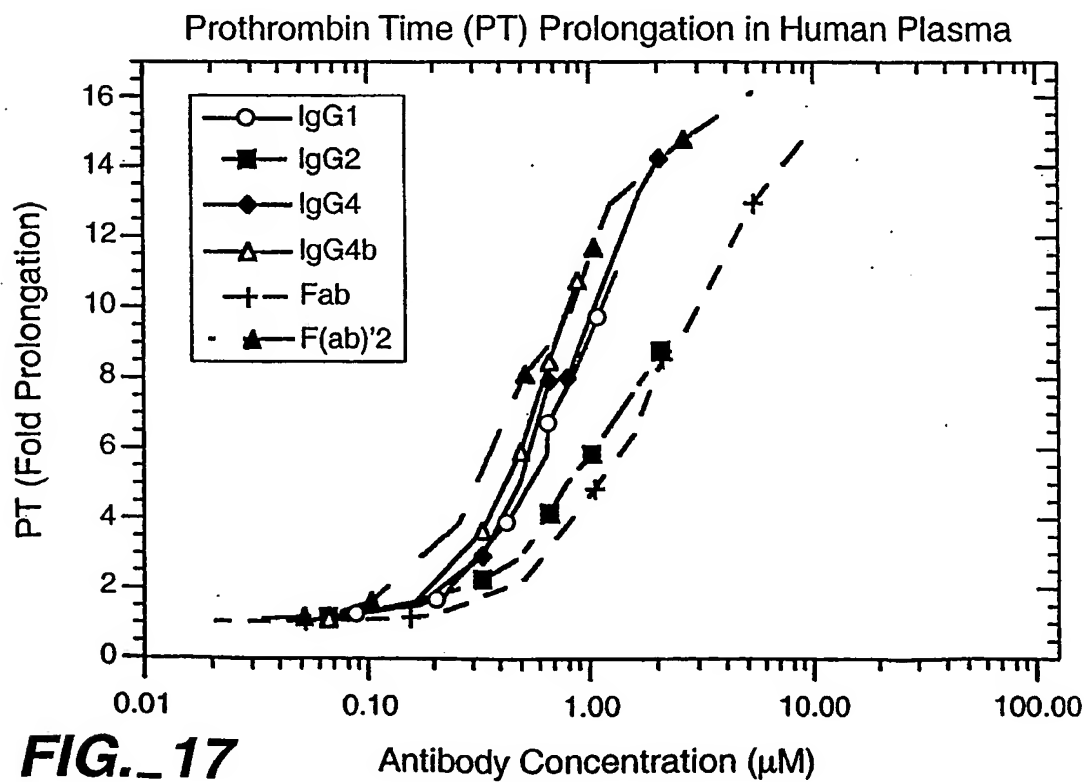
50 60 70 80 90  
5G6 VL AATSLADGVPSRFSGSGSGTKFSFKISSLQAEDFVSYCQQPYSSPYTF

-----  
CDR-L2-----  
CDR-L3

100  
5G6 VL GGGTKLELKRT (SEQ ID NO: 6)

**FIG.\_15**

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**FIG. 16****FIG. 17**

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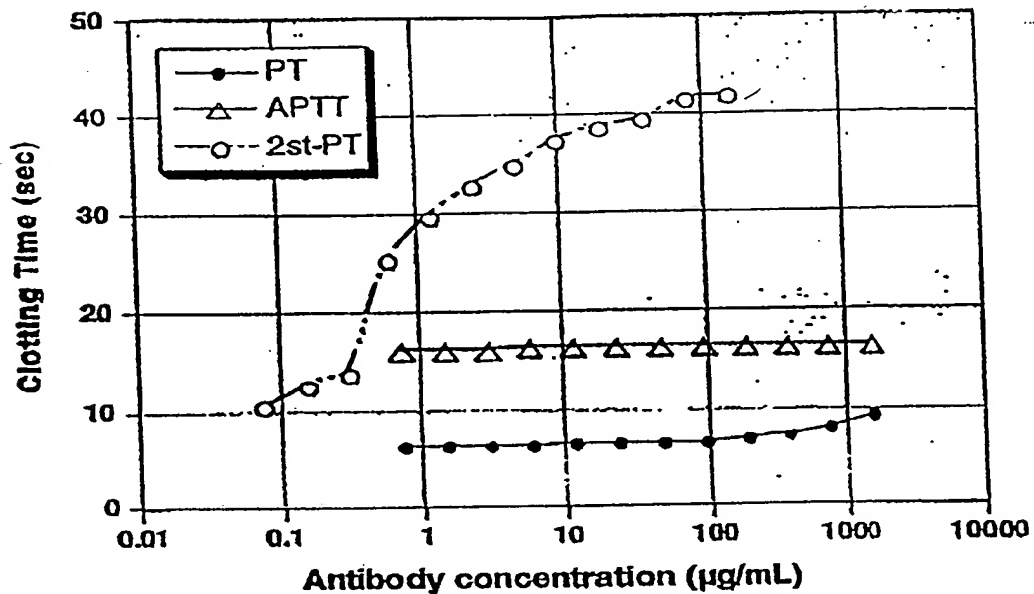
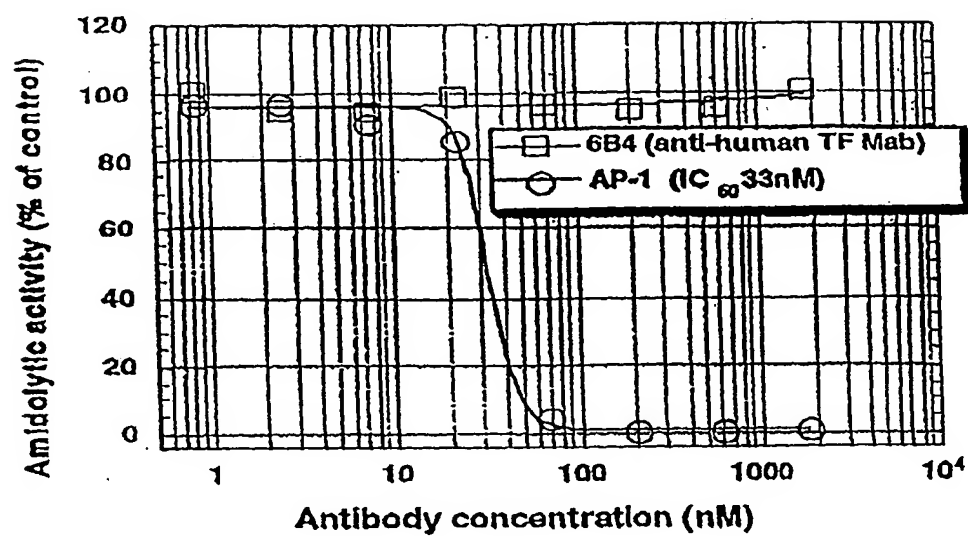
**FIG. 18****Comparison of potency of AP-1 in  
PT, 2st-PT and APTT coagulation assays**

FIG. 19

**In-Vitro Characterization****Inhibition of amidolytic activity of rabbit sTF/human VIIa complex**

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FIG. 20A

A.

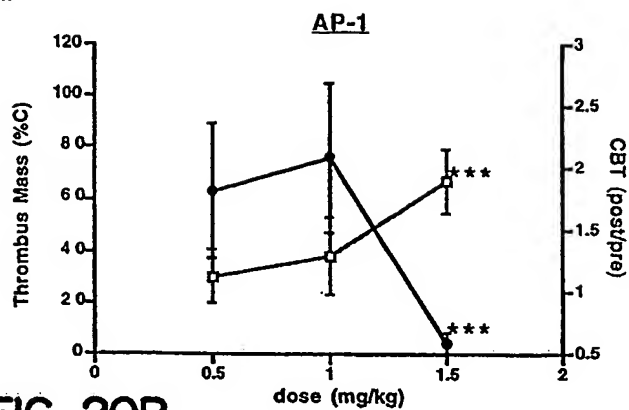


FIG. 20B

B.

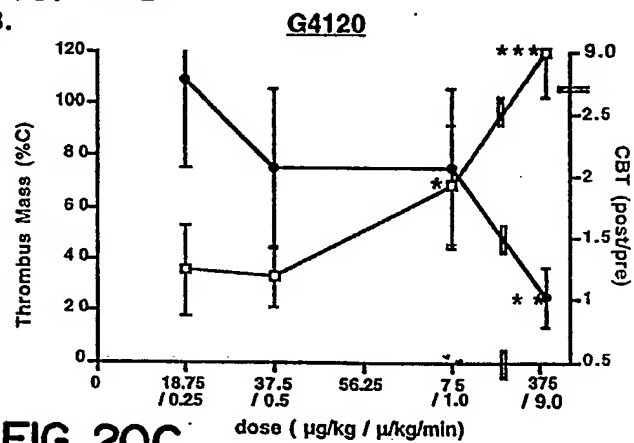
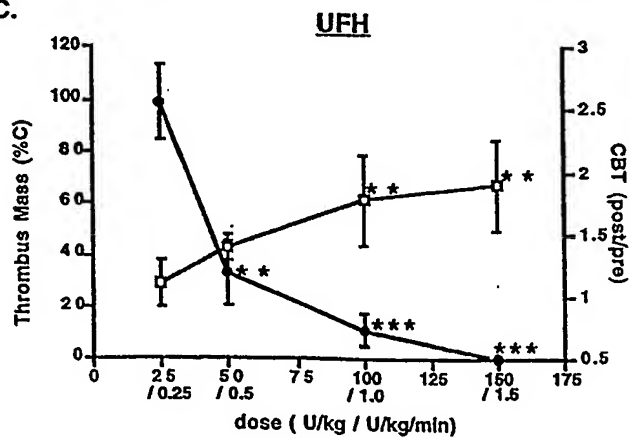


FIG. 20C

C.





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FIG. 2IA

A.

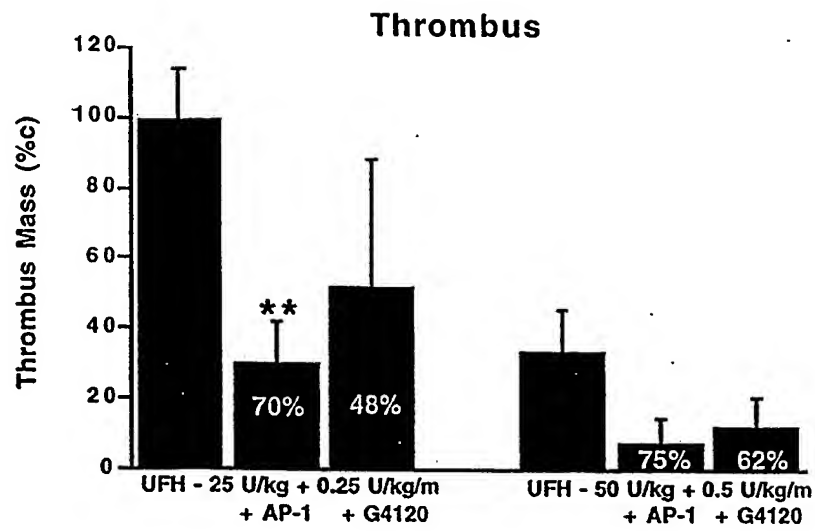
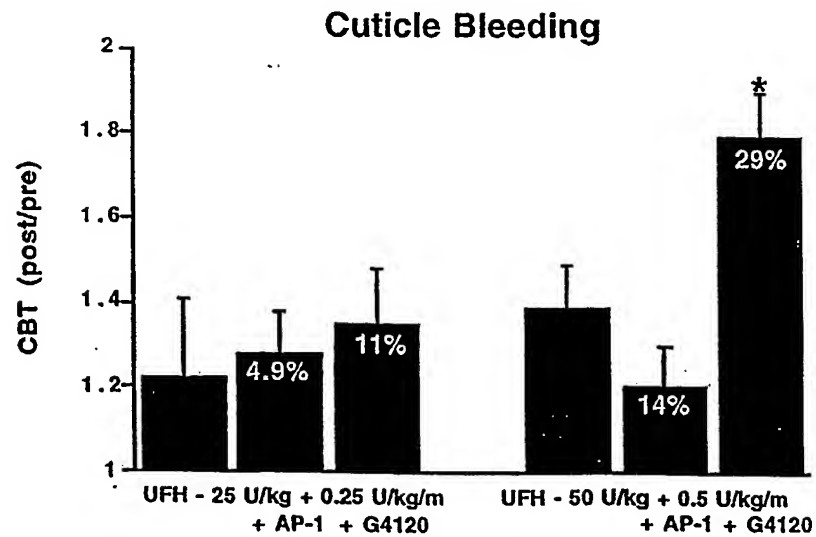


FIG. 2IB

B.



## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US03/17514

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : A61K 39/395; C07K 16/24

US CL : 424/130.1, 133.1, 141.1, 145.1, 158.1; 530/387.1, 387.3, 388.1, 388.3, 388.23

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/130.1, 133.1, 141.1, 145.1, 158.1; 530/387.1, 387.3, 388.1, 388.3, 388.23

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  
noneElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
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## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,437,864 A (EDGINGTON ET AL.) 01 August 1995 (01.08.1985), see entire document.	1-72



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Date of the actual completion of the international search

19 October 2003 (19.10.2003)

Date of mailing of the international search report

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Philip Gambel

Telephone No. 703-308-0196

INTERNATIONAL SEARCH REPORT

PCT/US03/17514

**Continuation of B. FIELDS SEARCHED Item 3:**

DIALOG, BIOSIS, CA, EMBASE, MEDLINE, WEST

search terms: refino, bunting, kirchhofer, tissue factor, tf, d3, 5g6, t58-5g9, ap-1, 7g11, gb4, hpt1, anticoagulant, heparin, VIIa, coagulation, infarction

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